

An Evidence-Based Systematic Review of Cinnamon (*Cinnamomum* spp.) by the Natural Standard Research Collaboration

Catherine Ulbricht, PharmD¹, Erica Seamon, PharmD², Regina C. Windsor, MPH², Nicole Armbruester, PhD³, J. Kathryn Bryan, BA², Dawn Costa, BA, BS², Nicole Giese, MS², Joerg Gruenwald, PhD⁴, Ramon Iovin, PhD², Richard Isaac, BA², Jill M. Grimes Serrano, PhD², Shaina Tanguay-Colucci, BS², Wendy Weissner, BA², Heeja Yoon, PharmD⁵, & Jie Zhang, MPH, PhD⁶

 ¹Massachusetts General Hospital, Boston, MA, USA, ²Natural Standard Research Collaboration, Somerville, MA, USA, ³Analyze & Realize, Berlin, Germany,
 ⁴Phytopharm Research, Berlin, Germany, ⁵Drake University, Des Moines, IA, USA,
 ⁶University of Alabama at Birmingham, Birmingham, AL, USA

ABSTRACT. An evidence-based systematic review of cinnamon (*Cinnamomum* spp.), including written and statistical analysis of scientific literature, expert opinion, folkloric precedent, history, pharmacology, kinetics/dynamics, interactions, adverse effects, toxicology, and dosing, by the Natural Standard Research Collaboration is discussed in this monograph.

KEYWORDS. Adverse effects, dosing, evidence-based, cinnamon (*Cinnamomum* spp.), interactions, pharmacodynamics, pharmacology, pharmacokinetics, systematic review

SYSTEMATIC AGGREGATION, ANALYSIS, AND REVIEW OF THE LITERATURE

Search Strategy

To prepare this Natural Standard review, electronic searches were conducted in several databases (including AMED, CANCERLIT, CINAHL, CISCOM, the Cochrane Library, EMBASE, HerbMed, International Pharmaceutical Abstracts, Medline, and NAPRALERT) from the inception of the study to October 2009. Search terms included the common name(s), scientific name(s), and all listed synonyms. Hand searches were conducted of 20 additional journals (not indexed in common databases) and of bibliographies from 50 selected secondary references. No restrictions were placed on language or quality of publications. Researchers in

Address correspondence to: Catherine Ulbricht, PharmD, Massachusetts General Hospital, Boston, MA, USA (E-mail: ulbricht@naturalstandard.com).

the field of complementary and alternative medicine (CAM) were consulted for access to additional references or ongoing research.

Selection Criteria

All literature was collected pertaining to efficacy in humans (regardless of study design, quality, or language), dosing, precautions, adverse effects, use in pregnancy/lactation, interactions, alteration of laboratory assays, and mechanism of action (in vitro, animal research, or human data). Standardized inclusion/exclusion criteria were utilized for selection.

Data Analysis

Data extraction and analysis were performed by healthcare professionals conducting clinical work and/or research at academic centers, using standardized instruments that pertained to each review section (defining inclusion/exclusion criteria and analytic techniques, including validated measures of study quality). Data were verified by a second reviewer.

Review Process

A blinded review was conducted by multidisciplinary research-clinical faculty at major academic centers with expertise in epidemiology and biostatistics, pharmacology, toxicology, CAM research, and clinical practice. In cases of editorial disagreement, a three-member panel of the Editorial Board addressed conflicts, and consulted experts when applicable. Authors of the studies were contacted when clarification was required (Natural Standard Systematic Review [www.naturalstandard.com] © 2011).

Synonyms/Common Names/Related Substances

- Cassia bark, cassia-bark tree, cassia cinnamon, cinnamal, cinnamaldehyde, cinnamate, cinnamic acid, cinnamic aldehyde, cinnamom-dhal chini, Cinnamomi cassiae, Cinnamomi cassiae cortex, Cinnamomi cevlanici cortex, Cinnamomi cortex, Cinnamomi flos, Cinnamomi osmophloeum, Cinnamomi ramulus, Cinnamomom, Cinnamomum aromaticum, Cinnamomum aromaticum Nees, Cinnamomum burmannii, Cinnamomum cassia, Cinnamomum cassia Blume, Cinnamomum cassia J. Presl, Cinnamomum cinnamon, Cinnamomum loureiroi, Cinnamomum mairei Levl., Cinnamomum migao, Cinnamomum obtusifolium, Cinnamomum osmophloeum clones (A and B), Cinnamomum osmophloeum Kaneh., Cinnamomum sieboldii, Cinnamomum sieboldii Meissn., Cinnamomum tamala, Cinnamomum tejpata, Cinnamomum verum, Cinnamomum verum J. Presl, Cinnamomum zeylanicum, Cinnamomum zeylanicum bark, Cinnamomum zeylanicum Blume, Cinnamomum zeylanicum Nees, cinnamon, cinnamon bark, cinnamon bark essential oil, cinnamon bark oil, cinnamon cortex, cinnamon leaf, cinnamon leaf essential oil, coca (Sanskrit), cocam (Sanskrit), Ceylon cinnamon, taj (Sanskrit).
- **Note**: This monograph focuses on cinnamon varieties that are edible and does not include *Cinnamomum camphora*, or the camphor tree, which may be lethal to humans in large doses, or *C. kotoense*, which is an ornamental species.

TABLE 1. Scientific evidence

Allergic rhinitis	С
Angina	С
Antioxidant	С
Bacterial infection	С
Candidiasis	С
Diabetes	С
Eye disorders	С
Helicobacter pylori infection	С
Insect repellant	С
Lung cancer	С
Metabolic syndrome (coronary heart disease)	С

CLINICAL BOTTOM LINE/EFFECTIVENESS

Brief Background

- Cinnamon has been used as a spice for centuries by countless cultural groups around the world. Individuals have also used cinnamon for its purported healing properties. It was traditionally used mainly as a stomachic and carminative medicine for gastrointestinal complaints and is still used for these conditions today [488]. The bark of *C. zeylanicum* and *C. cassia* is used as a spice (cinnamon bark). These two species are the only approved medicinal herbs of the genus *Cinnamonum*.
- At this time, high-quality human trials supporting the efficacy of cinnamon for any human indication are lacking. However, recent in-vitro and in-vivo research has discovered new potential properties of several cinnamon species.
- The treatment of diabetes (type 2) seems to be the most promising field of research for cinnamon [15, 39, 69, 101, 412]. Although there are conflicting results from two randomized studies, the results from in-vitro and animal studies indicate significant hypoglycemic effects. Cinnamon was shown to be highly effective in improving glucose and insulin metabolism. Researchers have recommended more studies for comparing the effectiveness of cinnamon in lowering A1C [131].
- Furthermore, due to the various potential effects of cinnamon and its constituents, including anti-inflammatory, antibacterial, antifungal, and antioxidant properties, it may prove effective in the supportive treatment of conditions such as cancer or severe viral infections (Table 1).

Natural Standard Evidence-Based Validated Grading RationaleTM

- Grades reflect the level of available scientific evidence in support of the efficacy of a given therapy for a specific indication.
- Expert opinion and historic/folkloric precedent are not included in this assessment, and are reflected in a separate section of each review ("Expert Opinion and Historic/Folkloric Precedent").
- Evidence of harm is considered separately; the grades presented in Table 2 apply only to evidence of benefit.

TABLE 2. Evidence-based grading and criteria

Level of evidence grade	Criteria
A (strong scientific evidence)	Statistically significant evidence of benefit from >2 properly conducted randomized controlled trials (RCTs), OR evidence from one properly conducted RCT AND one properly conducted meta-analysis, OR evidence from multiple RCTs with a clear majority of the properly conducted trials showing statistically significant evidence of benefit AND with supporting evidence in basic science, animal studies, or theory.
B (good scientific evidence)	Statistically significant evidence of benefit from 1–2 properly randomized trials, OR evidence of benefit from >1 properly conducted meta-analysis OR evidence of benefit from >1 cohort/case–control/non-randomized trials AND with supporting evidence in basic science, animal studies, or theory.
C (unclear or conflicting scientific evidence)	Evidence of benefit from >1 small RCT without adequate size, power, statistical significance, or quality of design by objective criteria, a OR conflicting evidence from multiple RCTs without a clear majority of the properly conducted trials showing evidence of benefit or ineffectiveness, OR evidence of benefit from >1 cohort/case–control/non-randomized trials AND without supporting evidence in basic science, animal studies, or theory, OR evidence of efficacy only from basic science, animal studies, or theory.
D (fair negative scientific evidence)	Statistically significant negative evidence (i.e., lack of evidence of benefit) from cohort/case–control/non-randomized trials, AND evidence in basic science, animal studies, or theory suggesting a lack of benefit.
F (strong negative scientific evidence)	Statistically significant negative evidence (i.e., lack of evidence of benefit) from >1 properly randomized adequately powered trial(s) of high-quality design by objective criteria. ^a
Lack of evidence ^b	Unable to evaluate efficacy due to lack of adequate available human data.

^aObjective criteria are derived from validated instruments for evaluating study quality, including the five-point scale developed by Jadad et al. [205], in which a score below 4 is considered to indicate lesser quality methodologically. ^bListed separately in the "Historical or Theoretical Uses That Lack Sufficient Evidence" section.

Historical or Theoretical Uses That Lack Sufficient Evidence

• Abdominal pain, abortifacient, abscess, acaricidal, acne, Alzheimer's disease [232, 323], analgesic [557], anesthetic, anthelmintic [290, 394], anticoagulant [543], antidepressant [198], anti-inflammatory [117, 132, 183, 195, 402, 452], antimutagenic [8, 102, 178, 207, 217, 218, 447, 447], antiparasitic [549], antiplatelet [206], antipyretic [256], antiseptic [99], antispasmodic, antiviral, arrhythmia [466, 471], arthritis, asthma [199], benign prostatic hyperplasia [327], bloating, blood purification, bronchitis, chronic bronchitis [559], chronic diarrhea, cognitive function [349], cold/flu, colic, cough, cystitis, dental caries [138, 414], deodorant [475], dermatitis, diarrhea [557], digestive aid [483], digestive disorders [51], diuretic [553], dyspepsia, eczema, emmenagogue, flavoring, food poisoning [441], food preservation [507, 544], food uses, gastric ulcer [483, 557], gastritis, gout [248, 552], gum disease [369], gynecologic disorders, HIV/AIDS [385], hypercholesterolemia [420], hypertension [517], hyperthyroid [134], immunostimulation [221,

244, 444], inflammatory conditions [277, 526], kidney disorders [24], lice [511], liver disease, long-term debility, loss of appetite, memory loss [251], movement disorders [117], muscle aches, nausea, neuralgia, neuroprotective [452], premature ejaculation, respiratory tract infection [202], rheumatoid arthritis [76], sciatica, sinusitis, skin conditions, snake repellent [89], sore throat, spermicide [44], toothache, tumors [171, 216, 257, 269, 355, 438], urethritis, urinary disorders [24], viral infections, weight gain, and wound healing [219].

Expert Opinion and Historic/Folkloric Precedent

- Cinnamon is a spice derived from the bark of the cinnamon tree. In western cuisine, cinnamon is often used with nutmeg, clove, and anise in baked goods, such as gingerbread. In Middle Eastern cuisine, it is often used in savory dishes. It has also been used in aromatherapy for supposed mood-enhancing effects [198]. Cinnamon is also a reported folk remedy in Pakistan [388]. *C. zeylanicum* has also been used in combination with other botanical species in order to treat kidney and urinary disorders in the tribal communities of the Ladakh region in India [24]. Guizhi (*C. zeylanicum*) decoctions have also been used in traditional Chinese medicine (TCM) [550], including the combination Shi-Quan-Da-Bu-Tang (Ten Significant Tonic Decoction) or SQT (Juzentaihoto, TJ-48) [548].
- Cinnamon may be used for various medical conditions [26, 277]. In a survey of parents in Germany determining the use of CAM in children with type 1 diabetes in four pediatric diabetes centers (located in Leipzig, Berlin, Stuttgart, and Bonn), 5.6% reported using cinnamon for this indication [98]. Cinnamon has been touted as having a positive effect on postprandial glucose metabolism [362]. Its ability to lower blood sugar in individuals with diabetes has been discussed [7]. Naturally occurring compounds found in cinnamon (*C. cassia*), including chromium and polyphenols, may improve insulin sensitivity [17]. Human data suggest that cinnamon exhibits "sweet" properties and may be used in strategies for reduction in sugar intake [32].
- As the sense of smell may be affected in individuals with Parkinson's disease (PD), cinnamon has been used in selective olfactory deficit tests to help diagnose PD-related hyposmia [114]. A cinnamon and citronellyl oil mixture has been used for the detection of allergy to perfumes [413].
- Cinnamaldehyde has been used as a filtering agent and a rubber-reinforcing agent. It is also used as a brightener in electroplating processes, as an animal repellent, an insect attractant, and an antifungal agent [5]. *Trans*-cinnamaldehyde is used as a flavor and fragrance ingredient.
- Cinnamon has been granted Generally Recognized as Safe (GRAS) status as a food additive by the US Food and Drug Administration (FDA). GRAS substances are considered safe and are not restricted, as is the case with other food additives. The FDA has sought fully up-to-date toxicology information on cinnamon (*Cinnamomum* species), including cinnamon bark oil, cinnamon oil, cinnamon leaf oil, and cinnamon oleoresin. The German Commission E and the European Scientific Cooperative on Phytotherapy (ESCOP) approved two medicinal herbs of the genus *Cinnamomum*: *C. zeylanicum* [35, 129] and *C. cassia* [35]. The bark is the only part of these plants that is used as a spice or for medical purposes (Cinnamomi cortex) [35].

Brief Safety Summary

- *Likely Safe*: When used orally and for short term (up to 6 weeks) in dosages up to 6 g daily [229].
- Possibly Unsafe: When used in patients taking drugs metabolized by cytochrome P450 (1A2 and 2E1, specifically), as cinnamon may alter agents metabolized by these enzymes based on in-vitro studies [87, 419, 527]. When used in patients using anticoagulant or antiplatelet agents, as cinnamon may decrease platelet counts and increase the risk of bleeding, based on animal study [206, 367]. When used in patients with diabetes or taking antidiabetic medications as, based on invitro and animal evidence, cinnamon has demonstrated lowering of blood glucose levels and acted as an insulin mimetic [6, 18, 29, 41, 56, 95, 121, 185, 200, 208, 223, 225, 228, 229, 252, 271, 309, 344, 367, 390–392, 405, 460, 461, 474, 481, 558]. Human data, however, have demonstrated conflicting results [13, 22, 33, 121, 242, 378, 476, 510]. When used in patients with autoimmune diseases or those who use immunosuppressants, as cinnamon has been found to have immunomodulatory effects in animal and in-vitro studies [244, 342, 343, 444]. When used in patients with liver damage or who are using hepatotoxic drugs, as in animal study of the essential oil of C. cassia stem bark, coumarin was isolated [87]. Coumarin may cause hepatotoxicity. When used in patients using antiarrhythmic agents, as cinnamon has demonstrated antiarrhythmic properties in animal studies [77, 466, 471, 473, 543, 553]. The effects of cinnamon with antiarrhythmic agents are not well understood. When used in patients using antilipemics, as based on animal evidence, cinnamon decreased serum total cholesterol and triglyceride concentrations and markedly increased HDL (high-density lipoprotein) cholesterol levels [273, 467]. In animals, cinnamate produced higher HDL cholesterol levels and lower atherogenic index compared with lovastatin, and inhibited hepatic 3-hydroxy-3-methylglutaryl CoA (HMG-CoA) reductase activity [273]. In in-vivo evidence in hamsters, a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels and inhibited cluster of differentiation 36 (CD36) and microsomal triglyceride transfer protein (MTTP) [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats [420].
- *Likely Unsafe*: When used in patients with a known allergy/hypersensitivity to cinnamon, its constituents, members of the Lauraceae family, or Balsam of Peru [3, 12, 37, 53, 91, 100, 116, 126, 127, 133, 150, 151, 156, 172, 173, 222, 240, 266, 298, 303, 325, 328, 329, 341, 358, 359, 373, 422, 433, 434, 493, 519, 525]. When used in large amounts (*C. aromaticum* (cassia)) (more than those found in foods), due to the potential for high levels of coumarin [299] and due to possible abortifacient effects (secondary sources).

DOSING/TOXICOLOGY

General

Listed doses are based on those most commonly used in available trials, on historical practice or on manufacturer recommendations. However, with natural

products, it is often not clear what the optimal doses are to balance efficacy and safety. Preparation of products may vary from manufacturer to manufacturer, and from batch to batch within one manufacturer. Because it is often not clear what the active components of a product are, standardization may not be possible, and the clinical effects of different brands may not be comparable.

Standardization

- Information about standardization of cinnamon products is lacking, but several studies have focused on processing and storage procedures for cinnamon. Gamma irradiation of cinnamon did not bring about any distinct qualitative or quantitative chemical changes based on spectrophotometric analysis [214]. However, another study demonstrated significant losses of total ascorbate in cinnamon as well as a significant decrease in carotenoid content 3 months after gamma irradiation [52]. Factors influencing the variation in constituents of cinnamon volatile oils, specifically in terms of their effect on aroma, have been investigated. In one study, compared with packaging material and storage duration, storage temperature has been suggested as the most important factor in altering cinnamon volatile oil aroma [245]. After disinfection by ethylene oxide and storage by ethylene oxide, a fast loss of residual ethylene oxide and ethylene glycol in cinnamon has been observed [63].
- Cinnamon bark has been confused with "yin xiang" [555]. "Yin xiang," according to secondary sources, may correspond with *C. burmannii* (Nees & T. Nees) Blume. A botany study was conducted that found that the pattern of morphology and distribution of calcium oxalate crystals may be an index for the identification of the crude drug of Cinnamomi cortex [535]. A *Compliance Policy Guide* has been published on the FDA's current Good Manufacturing Practices (cGMP) website regarding the levels that constitute legal action of insect infestation, mold, mammalian excreta, and rodent filth for whole cassia or ground cinnamon.

Dosing

Adult (age ≥ 18)
Oral.

- Antioxidant: Capsules containing 250 mg of an aqueous extract of cinnamon (Cinnulin PF®) twice daily for 12 weeks have been used with some evidence of benefit on the antioxidant status in overweight or obese individuals with impaired fasting glucose [411].
- Candidiasis: In a pilot study, eight lozenges of a commercially available cinnamon candy (not further specified) were taken daily for 1 week and were shown to be effective in three out of five HIV patients [393]. For oral candidiasis, a solution was made by cooking 250 g of cinnamon in 2,000 ml of water on medium heat until there was 500 ml of solution left (solution defined as 50% cinnamon solution) [55]. The treatment group gargled the solution 4–6 times a day, and each time with 20–30 ml of the solution.

- **Diabetes**: In a clinical trial, 1–6 g of cinnamon daily was used for 40 days [13, 33, 229]. In other such trials, cinnamon was administered in different ways: daily oral cinnamon (one capsule containing 333 mg of cinnamon extract three times daily) for 8 weeks [514], aqueous cinnamon extract corresponding to 3 g of cinnamon powder per day (duration unspecified) [309], 1.5 g of cinnamon cassia powder daily for 12 weeks [474], or cinnamon capsules 1 g daily for 90 days [95].
- *Helicobacter pylori* infection: In a clinical trial, 80 mg of cinnamon extract daily was used for 4 weeks [354].
- **Metabolic syndrome**: Two capsules (250 mg) of a water-soluble cinnamon extract, Cinnulin PF[®], twice daily [558]. According to the manufacturer, 500 mg of Cinnulin PF[®] is equivalent to around 10 g of cinnamon powder (20:1 extract). It contains approximately 1% double-linked polyphenol type-A polymers, which are considered to be the bioactive component of cinnamon.

Topical.

• **Insect repellant**: Single applications of: (1) cream containing 5% (w/w) cassia oil formulated with 5 g of cassia oil evaluated for up to 120 min, (2) 0.006–0.102 mg/cm² of *C. cassia* bark-derived extract (patch bioassay), (3) 0.013–0.153 mg/cm² of *trans*-cinnamaldehyde or cinnamyl alcohol (patch bioassay), (4) 0.006–0.102 mg/cm² of *C. cassia* bark-derived methanol extract (skin bioassay), or (5) 0.003–0.051 mg/cm² of *trans*-cinnamaldehyde or cinnamyl alcohol (skin bioassay) for up to 40 min [65].

Children (age < 18)

• Insufficient available evidence.

Toxicology

- Based on human study (two randomized trials, one controlled trial, and one pilot study) of the effects of cinnamon on type-2 diabetes, *H. pylori* infection, and candidiasis, no toxic effects were observed [229, 354, 393, 510].
- Ethanolic extracts of *C. zeylanicum* bark demonstrated no acute or chronic oral toxicity in mice [439]. However, *C. zeylanicum* treatment caused reduction in liver weight of the treated animals compared with the control. Hematological studies revealed a fall in hemoglobin level. The extract also induced an increase in reproductive organ weight, sperm motility, and sperm count, and it failed to illicit any spermatotoxic effect. The volatile oils of cinnamon and eugenol have also revealed a potent spermicidal action, whereas the fixed oils were devoid of action on spermatozoa [44].
- Cinnamon has been shown to affect xanthine dehydrogenase, aldehyde oxidase, and pyridoxal oxidase activity during development in *Drosophila melanogaster* [43]. High doses of cinnamon oil caused a depressive effect in rats, probably due to toxicity; the authors note that at the lowest dose, it caused weak or "doubtful" effects [148]. The ethanol extract of cinnamon has shown no in-vitro mutagenic activity [498]. *Trans*-cinnamaldehyde was found to induce lethal mutations

in *Drosophila melanogaster* ("fruit flies") [531]. Ceylon cinnamon (the bark of *C. zeylanicum*) has also displayed mutagenic activity [499]. Cinnamaldehyde, cinnamyl alcohol, methyl eugenol, eugenol, isoeugenol, as well as cinnamon bark oil, were positive in the *Bacillus subtilis* DNA-repair test (rec assay) without S9. All samples tested were negative in the *Escherichia coli* WP2 uvrA reversion test. The essential oil was positive in the DNA-repair test [435]. *C. mairei* extract was positive in the chromosomal aberration and micronucleus assays in mice [539]. *C. zeylanicum* bark showed low mutagenic activity in *B. subtilis* strains H17 (rec+) and M45 (rec-) [498]. *Trans*-cinnamaldehyde caused increased rates of structural and numerical chromosome abnormalities and increases in frequencies of cells containing 3n and 4n chromosomes [226].

- After two years of study, rats and mice given diets of 1,000, 2,100, or 4,100 ppm of *trans*-cinnamaldehyde showed no evidence of carcinogenic activity [5]. Exposure to *trans*-cinnamaldehyde resulted in olfactory epithelial pigmentation in male and female mice [5].
- Squamous cell papillomas and carcinomas of the forestomach were observed in male and female mice. In the 3-month studies, rats and mice were given diets containing 4,100, 8,200, 16,500, or 33,000 ppm of *trans*-cinnamaldehyde. The incidence of squamous epithelial hyperplasia of the forestomach was significantly increased in rats exposed to 8,200 ppm or greater and female mice exposed to 33,000 ppm. In rats, feed consumption was reduced in all exposed groups. In mice, feed consumption was reduced in the highest-dose groups. Body weights of all treated males were less than those of controls. Body weights were reduced in female rats exposed to 16,500 or 33,000 ppm and female mice exposed to 8,200 ppm or greater. All rats survived to the end of the study, but some male mice in the highest dose groups died due to inanition from unpalatability of the dosed feed. In mice, the incidence of olfactory epithelial degeneration of the nasal cavity was significantly increased in males and females exposed to 16,500 ppm and those to 33,000 ppm [5].
- Raw cinnamon (*C. zeylanicum*) has been shown to be tumorigenic in high doses [23]. A case report mentions a 24-year-old woman who developed a squamous cell carcinoma of the tongue following persistent and prolonged exposure to cinnamon-flavored gum [525].
- Cinnamon oil ingestion led to toxic manifestations in a child, according to a case report [379].
- Molecules similar to cinnamic acid, such as styrene and the related aldehyde, alcohol, and esters, are all considered more toxic than cinnamic acid [189]. Screening tests are performed to test food additives such as cinnamic aldehyde for mutagenicity and carcinogenicity [203].
- Cinnamon oil has been used recreationally by children and adolescents to "get high." Nausea or abdominal pain, but no systemic effects, have been reported [375, 432].
- In animal study of the essential oil of *C. cassia* stem bark, coumarin was isolated [87]. Coumarin may cause hepatotoxicity. Due to the potential for high levels of coumarin, European health agencies have recently warned against the consumption of large amounts of *C. aromaticum* (cassia) [299].

• Cinnamaldehyde and N,N-dimehtylcinnamylamine, chemical analogs of the neurotoxin N-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), lacked dopaminergic nigrostriatal neurotoxic effects in animal study [376].

ADVERSE EFFECTS/PRECAUTIONS/CONTRAINDICATIONS

Allergy

- Known allergy/hypersensitivity to cinnamon, its constituents, members of the Lauraceae family, or Balsam of Peru [1, 46, 50, 100, 103, 108, 113, 116, 128, 135, 151, 156, 190, 222, 227, 230, 240, 279, 304–307, 315, 319, 325, 348, 352, 353, 358, 415, 417, 426, 429, 465, 487, 508, 521]. However, scratch-chamber testing often leads to false-positive irritant reactions. A positive test to Balsam of Peru may indicate a spice allergy, but the absence of such reaction does not rule it out [113].
- Concomitant reactions to cinnamyl alcohol may be due to cross-sensitization in patients with contact allergy to ketoprofen [141, 155].
- One case of allergic contact dermatitis has been reported as a result of airborne exposure to cinnamon [9].
- Cinnamon is one of the 10 major food allergens [463]. Cinnamaldehyde seems to be considered the "true" allergen, while cinnamyl alcohol and cinnamic acid are transformed in the skin to cinnamaldehyde before contact allergic reactions may occur [194, 524]. Studies confirmed the sensitivity of patients to cinnamic aldehyde in a toothpaste [241, 407, 490]. Based on review, cinnamon constituents found in cosmetics, including cinnamic alcohol, eugenol, and coumarin, may be more likely to cause hypersensitivity and sensitization reactions than other compounds, such as cinnamic aldehyde [97, 307]. However, results from other studies are not consistent with these findings. Cinnamic aldehyde has been found to be a potent sensitizer and a strong cross-sensitizer [437].
- Immunologic reactions to spices such as cinnamon may be related to acute symptoms and lung function changes, but not to chronic changes [560]. Concerning allergic reactions to cinnamon dust, it may be the cellulose content that is responsible for the histological reactions [485]. Cinnamon powder has shown low cross-reactivity in patients with positive skin tests to birch or mugwort pollens and celery [464]. Alcohol as a vehicle was shown to have a higher sensitization potential than petrolatum when cinnamon bark oil was used in predictive tests [316].
- Allergic contact stomatitis to cinnamon in chewing gum was mistaken as facial angioedema [238].
- Cinnamon has produced nonimmunologic contact urticaria (NICU), which may occur without previous immunologic sensitization in exposed individuals [157, 162, 260–264]. Acetylsalicylic acid (aspirin) has been shown to inhibit this effect through prostaglandin bioformation [264]. Indomethacin and dexamethasone may also inhibit these reactions, based on animal study; however, the mechanism is not well understood [263]. In laboratory study, eugenol and d-limnonene may quench (inhibit) the sensitization potential of cinnamaldehyde through competitive inhibition at the receptor level [162].

• An ingredient in perfume that is a sensitizer may become hypoallergenic during the aging process of the perfume. Patients may tolerate an aged perfume that contains cinnamic aldehyde without acquiring allergic reactions [140].

Adverse Effects

- **General**: No adverse effects were observed in a pilot trial with five HIV patients [393]. In two randomized trials on the effects of cinnamon on type 2 diabetes, no adverse effects were observed [229, 510]. One controlled trial reported minor adverse effects in five out of 15 patients [354].
- As with any spice or drug, cinnamon may be contaminated by microorganisms during storage. The microbiological quality of cinnamon was evaluated in several studies. Cinnamon showed mainly satisfactory microbiological quality [404, 431]. However, contamination by aflatoxin-producing fungi may constitute health hazards in humans, as the aflatoxin level is not reduced by domestic cooking [27, 125, 302]. Furthermore, cinnamon may contain detectable ethylene oxide [142].
- **Dermatologic**: Allergic hypersensitivity and contact allergic reactions may occur in sensitive individuals [46, 50, 100, 103, 108, 113, 116, 151, 156, 190, 222, 230, 240, 267, 304, 305, 315, 325, 352, 353, 358, 415, 417, 426, 429, 465, 487, 508, 521]. Dermatitis, photodermatitis, stomatitis, glossitis, gingivitis, perioral dermatitis, perioral leukoderma (simulating vitiligo), oral lesions, cheilitis, eczema, lip edema, irritation, and depigmentation have been noted in case reports after external application of cinnamon (e.g., cinnamon oils in fragrances or cinnamic aldehyde in deodorant) as well as after the use of flavored chewing gums, mints, or toothpastes [3, 12, 20, 27, 31, 40, 47, 50, 51, 52, 59, 91, 98, 100, 107, 110, 112, 135, 136, 137, 142, 156, 157, 158, 159, 163, 164, 167, 170, 178, 187, 188, 206, 237, 242, 243, 246, 247, 269, 285, 286, 297, 317, 323, 330, 339, 340, 346, 349, 350, 362, 369, 370, 380, 381, 403, 409, 429, 432, 447, 440, 451, 452, 456, 459, 460, 461, 484, 493, 503, 515, 519, 522, 539, 549, 551]. One case of allergic contact dermatitis has been reported as a result of airborne exposure to cinnamon [9]. Cinnamaldehyde may provoke orofacial granulomatosis, urticaria, dermatitis, and stomatitis [116, 240, 266, 373, 397]. Squamous cell carcinoma of the tongue and "speckled" lesions have been reported following exposure to cinnamon-flavored gum [84, 525]. A case report of a burn from cinnamon oil has been reported [462]. A case of a 68-year-old Caucasian female with type 2 diabetes mellitus who experienced an acute exacerbation of her rosacea 2 weeks after self-initiating cinnamon oil pills to lower her blood sugar levels has been reported [54]. In a clinical trial using 1 g of cinnamon in capsule form daily, one adverse event (rash) was reported by an individual who withdrew from the study [95].
- Cinnamal may be added to soaps and douches as a fragrance and may cause anogenital dermatitis [521]. Based on retrospective report, the incidence of allergic reactions to cinnamic aldehyde has decreased over time [351].
- **Gastrointestinal**: Nausea and abdominal pain have been reported with cinnamon use [375, 432].
- **Hematologic**: Cinnamon bark caused a significant decrease in platelet counts in normal rats after long-term use [367].

- **Hepatic**: In animal study of the essential oil of *C. cassia* stem bark, coumarin was isolated [87]. Coumarin may cause hepatotoxicity. Due to the potential for high levels of coumarin, European health agencies have recently warned against the consumption of large amounts of *C. aromaticum* (cassia) [299].
- **Neurologic**: Based on secondary sources, cinnamon oil has been used recreationally by children and adolescents to induce an altered state of consciousness.
- **Pulmonary/respiratory**: Asthma and other chronic respiratory symptoms were seen in spice-factory workers [500, 501, 560].
- **Renal**: Based on secondary information, supplementation with cinnamon may increase risk of hyperoxaluria; however, this was not found to be the case based on human evidence [484].

Precautions/Warnings/Contraindications

- Avoid in patients with a known allergy/hypersensitivity to cinnamon, its constituents, members of the Lauraceae family, or Balsam of Peru [3, 12, 37, 53, 91, 100, 116, 126, 127, 133, 150, 151, 156, 172, 173, 222, 240, 266, 298, 303, 325, 328, 329, 341, 358, 359, 373, 422, 433, 434, 493, 519, 525].
- Avoid large amounts of cinnamon (more than those found in foods) due to possible abortifacient effects, based on secondary sources.
- Avoid large amounts of *C. aromaticum* (cassia) (more than those found in foods) due to the potential for high levels of coumarin [299].
- Use cautiously in patients taking drugs metabolized by cytochrome P450 (1A2 and 2E1, specifically), as cinnamon may alter agents metabolized by these enzymes based on in-vitro studies [87, 419, 527].
- Use cautiously in patients using anticoagulant or antiplatelet agents, as cinnamon may decrease platelet counts and increase the risk of bleeding, based on animal study [206, 367].
- Use cautiously in patients with diabetes or those taking antidiabetic medications, as based on in-vitro and animal evidence, cinnamon has demonstrated lowering of blood glucose levels and acted as an insulin mimetic [6, 18, 29, 41, 56, 95, 121, 185, 200, 208, 223, 225, 228, 229, 252, 271, 309, 344, 367, 390–392, 405, 460, 461, 474, 481, 558]. Human data, however, have demonstrated conflicting results [13, 22, 33, 121, 242, 378, 476, 510]. Theoretically, concurrent use of cinnamon with blood sugar-lowering agents may have additive effects and may increase the risk of hypoglycemia.
- Use cautiously in patients with autoimmune diseases or who use immunosuppressants, as cinnamon has been found to have immunomodulatory effects in animal and in-vitro studies [244, 342, 343, 444].
- Use cautiously in patients with liver damage or who are using hepatotoxic drugs, as in animal study of the essential oil of *C. cassia* stem bark, coumarin was isolated [87]. Coumarin may cause hepatotoxicity.
- Use cautiously in patients using antiarrhythmic agents. as cinnamon has demonstrated antiarrhythmic properties in animal studies [77, 466, 471, 473, 543, 553]. The effects of cinnamon with antiarrhythmic agents are not well understood.
- Use cautiously in patients using antilipemics, as based on animal evidence, cinnamon decreased serum total cholesterol and triglyceride concentrations and

markedly increased HDL cholesterol levels [273, 467]. In animals, cinnamate produced higher HDL cholesterol levels and lower atherogenic index compared with lovastatin, and inhibited HMG-CoA reductase activity [273]. In in-vivo evidence in hamsters, a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels and inhibited CD36 and MTTP [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats [420].

Pregnancy and Lactation

- Not suggested due to lack of sufficient data.
- Based on secondary sources, consumption of large amounts of cinnamon (more than those found in foods) may have abortifacient effects.
- The effect of cinnamon and eugenol on human spermatozoa motility in vitro has been studied [46]. The volatile oils studied revealed a potent spermicidal action, whereas the fixed oils were devoid of action on spermatozoa.
- Fish exposed to ethanol and cinnamaldehyde had greater adverse effects on fetal development when combined, compared with their individual effects [173].
- There is a lack of information for cinnamon in the National Library of Medicine's Drugs and Lactation Database (LACT-MED).

INTERACTIONS

Cinnamon-Drug Interactions

- **Alcohol**: Fish exposed to ethanol and cinnamaldehyde had greater adverse effects on fetal development when combined, compared with their individual effects [167].
- **Alzheimer's agents**: In laboratory study, chloroform extracts of *C. cassia* showed a marginal neuronal cell protection from direct betaA(1-42) insult [232]. Theoretically, concurrent use of cinnamon with Alzheimer's agents may have beneficial, additive effects.
- **Analgesics**: Based on animal evidence, an ethanolic extract of *C. zeylanicum* was shown to possess an antinociceptive effect [21]. Theoretically, concurrent use of cinnamon with analgesic agents may have additive effects.
- Antiarrhythmics: Cinnamon has demonstrated antiarrhythmic properties in animal studies [77, 466, 471, 473, 543, 553]. The effects of cinnamon with antiarrhythmic agents are not well understood.
- Antibiotics: In vitro, cinnamon has demonstrated antibacterial properties [109, 145, 201, 202, 369, 441, 459, 544]. A synergistic antimicrobial effect was noted when chlorhexidine was used in combination with essential oils of cinnamon, tea tree (*Melaleuca alternifola*), manuka (*Leptospermum scoparium*), *L. morrisonii*, arnica, eucalyptus, and grapefruit against biofilm and planktonic cultures of *Streptococcus mutans* and *Lactobacillus plantarum* [138]. Theoretically, concurrent use of cinnamon with antibiotic agents may have additive effects. Based on secondary sources, concomitant use of cinnamon with tetracyclines may slow

the absorption and reduce blood levels of tetracycline antibiotics. This effect may be due to adsorption of tetracycline by cinnamon.

- Anticoagulants and antiplatelets: Based on animal evidence, cinnamon bark, cinnamaldehyde, and two other *Cinnamomum* species (*C. altissimum* and *C. pubescens*) caused a decrease in platelet counts after long-term use [193, 206, 320, 367]. Cinnamic aldehyde inhibited arachidonic acid (AA) release and thromboxane B2 formation, which may contribute to reduced platelet aggregation [480]. Theoretically, concurrent use of cinnamon with anticoagulants or antiplatelets may increase the risk of bleeding.
- Antidiabetics: Based on in-vitro and animal evidence, cinnamon has demonstrated lowering of blood glucose levels and acted as an insulin mimetic [6, 18, 29, 41, 56, 95, 121, 185, 200, 208, 223, 225, 228, 229, 252, 271, 309, 344, 367, 390–392, 405, 460, 461, 474, 481, 558]. Human data, however, have demonstrated conflicting results [13, 22, 33, 121, 242, 378, 476, 510]. Theoretically, concurrent use of cinnamon with blood sugar-lowering agents may have additive effects and increase the risk of hypoglycemia.
- Antifungals: Cinnamon has demonstrated antifungal effects in vitro [79, 174, 285, 317, 393, 457] and inhibited oral candidiasis in humans [55]. Based on laboratory study, *C. cassia* in combination with amphotericin B displayed additive antifungal effects and was less toxic compared with amphotericin B alone [154]. Theoretically, concurrent use of cinnamon with antifungal agents may have additive effects.
- Antihypertensives: *C. migao* oil reduced systolic and diastolic arterial blood pressure in animal study [473]. Based on animal study, whole cinnamon and aqueous extracts have been found to reduce systolic blood pressure (SBP) elevations as well as a genetic component of elevated blood pressure [386]. Human study also demonstrated a reduction in SBP upon treatment with Cinnulin PF[®] [558]. Theoretically, concurrent use of cinnamon with antihypertensive agents may have additive effects and increase the risk of hypotension.
- Anti-inflammatories: Based on in-vitro evidence, cinnamon bark may exert anti-inflammatory properties [256, 402]. Theoretically, concurrent use of cinnamon with anti-inflammatory agents may have additive effects.
- Antilipemics: Based on animal evidence, *C. zeylanicum* significantly decreased serum total cholesterol and triglyceride concentrations and markedly increased HDL cholesterol levels [273, 467]. In animal study, cinnamate, a phenolic compound in cinnamon bark, produced higher HDL cholesterol levels and lower atherogenic index compared with lovastatin [273]. Cinnamate inhibited HMG-CoA reductase activity [273]. In in-vivo evidence in hamsters, a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels and inhibited CD36 and MTTP [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats when included in the diet at about fivefold the normal human intake level [420]. Theoretically, concurrent use of cinnamon with antilipemic agents may have additive effects.
- Antineoplastics: Based on in-vitro and animal evidence, cinnamon has exerted antitumor and antigenotoxic effects [8, 171, 216, 355]. Theoretically,

concurrent use of cinnamon with antineoplastic agents may have additive effects.

- Antiobesity agents: In a clinical trial studying the effects of Cinnulin PF[®], compared with placebo, the treatment group noted increases in lean mass (+1.1%: from 53.7 ± 11.8 kg [pre] to 54.3 ± 11.8 kg [post], p < .002) [558].
- Antiretrovirals: Based on clinical study, *C. cassia* bark extract inhibited virus-induced cytopathogenicity in MT-4 cells infected with HIV [385]. Theoretically, concurrent use of cinnamon with antiretroviral agents may have additive effects.
- Antispasmodics: Based on secondary sources, cinnamon may have antispasmodic
 effects. Theoretically, concurrent use of cinnamon with antispasmodic agents may
 have additive effects.
- Aspirin: Aspirin (acetylsalicylic acid) has been shown to reduce contact urticaria reactions caused by cinnamic aldehyde and cinnamic acid due to inhibitory effects of aspirin on prostaglandin bioformation [271].
- Cytochrome P450-metabolized agents: Based on in-vitro evidence, cinnamon or its constituents may interact with hepatic microsomal cytochrome P450 [87, 419, 469, 527]. Cinnamon bark was found to inhibit aminopyrine N-demethylation in rat liver microsomes. The component inhibiting drug oxidations catalyzed by CYP1A2 and CYP2E1 was isolated from Cinnamomi cortex and was identified as o-methoxycinnamaldehyde (OMCA) [176]. *C. burmannii* bark inhibited CYP3A4 and 2D6 via erythromycin N-demethylation and dextromethorphan O-demethylation activities in human liver microsomes [468]. Theoretically, cinnamon may alter the levels of drugs metabolized by cytochrome P450, specifically 1A2 and 2E1.
- **Dexamethasone**: Based on animal study, dexamethasone inhibited nonimmunologic contact urticaria reactions to cinnamic acid and cinnamic aldehyde [263].
- **Drugs that affect GABA**: In animal study, cinnamon may exert an anxiolytic effect via regulation of the serotonergic and GABAergic systems [542]. Theoretically, concurrent use with other drugs that affect GABA (gamma-amino butyric acid) may have additive effects.
- **Estrogens**: Cinnamomi cortex as a component of the unkei-to combination product stimulated estradiol secretion in laboratory study [472]. Theoretically, concurrent use of cinnamon with estrogens may have additive effects.
- **Hepatotoxics**: In animal study of the essential oil of *C. cassia* stem bark, coumarin was isolated [87]. Coumarin may cause hepatotoxicity. Due to the potential for high levels of coumarin, European health agencies have recently warned against the consumption of large amounts of *C. aromaticum* (cassia) [299]. Theoretically, concurrent use of cinnamon with hepatotoxic agents may increase the risk of liver damage.
- Immunosuppressants: Based on in-vitro and animal evidence, cinnamon demonstrated immunomodulatory effects [244, 342, 343, 444]. Theoretically, cinnamon may alter the effects of immunosuppressants.
- **Indomethacin**: Based on animal study, indomethacin inhibited nonimmunologic contact urticaria reactions to cinnamic acid and cinnamic aldehyde [263].
- **Sympathomimetics**: In animal study, TRPA1 agonists, such as cinnamaldehyde, have been shown to activate the sensory nerves and induce adrenaline secretion

via the central nervous system [204]. Theoretically, concurrent use with sympathomimetics may have additive effects.

- **Terfenadine**: Based on human study, terfenadine did not inhibit contact urticaria reactions caused by cinnamic aldehyde [260].
- **Tetracyclines**: Based on secondary sources, concomitant use of cinnamon with tetracyclines may slow the absorption and reduce blood levels of tetracycline antibiotics. This effect may be due to adsorption of tetracycline by cinnamon.

Cinnamon-Herb-Supplement Interactions

- Alzheimer's herbs: In laboratory study, chloroform extracts of *C. cassia* showed a marginal neuronal cell protection from direct betaA(1–42) insult [232]. Theoretically, concurrent use of cinnamon with Alzheimer's herbs may have beneficial, additive effects.
- **Analgesics**: Based on animal evidence, an ethanolic extract of *C. zeylanicum* was shown to possess an antinociceptive effect against both acetic acid-induced writhing and hot plate-induced thermal stimulation in mice [21]. Theoretically, concurrent use of cinnamon with analgesic agents may have additive effects.
- **Antiarrhythmics**: Cinnamon has demonstrated antiarrhythmic properties in animal studies [77, 466, 471, 473, 543, 553]. The effects of cinnamon with antiarrhythmic agents are not well understood.
- Antibacterials: In vitro, cinnamon has demonstrated antibacterial properties [145, 201, 202, 369, 441, 459, 544]. Theoretically, concurrent use of cinnamon with other antibacterial agents may have additive effects. Based on secondary sources, concomitant use of cinnamon with tetracyclines may slow the absorption and reduce blood levels of tetracycline antibiotics. This effect may be due to adsorption of tetracycline by cinnamon.
- Anticoagulants and antiplatelets: Based on animal evidence, cinnamon bark, cinnamaldehyde, and two other *Cinnamomum* species (*C. altissimum* and *C. pubescens*) caused a decrease in platelet counts after long-term use [193, 206, 320, 367]. Cinnamic aldehyde inhibited AA release and thromboxane B2 formation, which may contribute to reduced platelet aggregation [480]. Theoretically, concurrent use of cinnamon with anticoagulants/antiplatelets may increase the risk of bleeding.
- Antifungals: Cinnamon has demonstrated antifungal properties in vitro [79, 174, 285, 317, 393, 457] and inhibited oral candidiasis in humans [55]. Based on laboratory study, *C. cassia* in combination with amphotericin B displayed additive antifungal effects and was less toxic compared with amphotericin B alone [154]. Theoretically, concurrent use of cinnamon with antifungal agents may have additive effects.
- Anti-inflammatory herbs: Based on in-vitro evidence, cinnamon bark exerted anti-inflammatory properties [256, 402]. Theoretically, concurrent use of cinnamon with anti-inflammatory agents may have additive effects.
- Antilipemics: Based on animal evidence, *C. zeylanicum* significantly decreased serum total cholesterol and triglyceride concentrations and markedly increased HDL cholesterol levels [273, 467]. In animal study, cinnamate, a phenolic compound in cinnamon bark, produced higher HDL cholesterol levels and lower

atherogenic index compared with lovastatin [273]. Cinnamate inhibited HMG-CoA reductase activity [273]. In in-vivo evidence in hamsters, a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels and inhibited CD36 and MTTP [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats when included in the diet at about fivefold the normal human intake level [420]. Theoretically, concurrent use of cinnamon with antilipemic agents may have additive effects.

- Antineoplastics: Based on in-vitro and animal evidence, cinnamon has exerted antitumor and antigenotoxic effects [8, 171, 216, 355]. Theoretically, concurrent use of cinnamon with antineoplastic agents may have additive effects.
- Antiobesity herbs and supplements: In a clinical trial studying the effects of Cinnulin PF®, compared with the placebo, the treatment group noted increases in lean mass (+1.1%: from 53.7 \pm 11.8 kg [pre] to 54.3 \pm 11.8 kg [post], p < .002) [558].
- Antioxidants: Cinnamon bark has been shown to contain very high concentrations of antioxidants [115]. Several animal and in-vitro studies demonstrate the antioxidant effects of the essential oil obtained from the bark of *C. zeylanicum* and its main components [83, 175, 210, 236, 272, 278, 455, 491]. Etheric, methanolic, and aqueous cinnamon extracts have also inhibited oxidative processes in vitro [106, 187, 248, 275, 286, 308, 318, 451]. In human study, a dried aqueous extract of cinnamon (Cinnulin PF®), increased the antioxidant status [411]. Theoretically, concurrent use of cinnamon with antioxidants may have additive effects.
- Antispasmodics: Based on secondary sources, cinnamon may have antispasmodic
 effects. Theoretically, concurrent use of cinnamon with antispasmodic agents may
 have additive effects.
- Antivirals: Based on clinical study, *C. cassia* bark extract may be effective against HIV-1 and HIV-2 replication in terms of inhibition of virus-induced cytopathogenicity in MT-4 cells infected with HIV [385]. Theoretically, concurrent use of cinnamon with antiviral agents may have additive effects.
- **Artemisia**: In laboratory study, when *C. camphora* was mixed with *Artemisia princeps* Pamp. (1:1 mixture), a synergistic insecticidal effect was noted [291].
- Cytochrome P450-metabolized herbs and supplements: Based on in-vitro studies, cinnamon or its constituents may interact with hepatic microsomal cytochrome P450 [87, 419, 469, 527]. Cinnamon bark was found to inhibit aminopyrine N-demethylation in rat liver microsomes. The component inhibiting drug oxidations catalyzed by CYP1A2 and CYP2E1 was isolated from Cinnamomi cortex and was identified as OMCA [176]. *C. burmannii* bark inhibited CYP3A4 and 2D6 via erythromycin N-demethylation and dextromethorphan O-demethylation activities in human liver microsomes [468]. Theoretically, cinnamon may alter the levels of drugs metabolized by cytochrome P450, specifically 1A2 and 2E1.
- Clove: Based on a review, synergistic antibacterial effects have been observed between cinnamaldehyde and eugenol, a constituent of clove [47].

- **Ephedra**: Components of ephedra reportedly interact with cinnamon; however, details of this interaction are not well documented [416].
- **Hepatotoxic herbs**: In animal study of the essential oil of *C. cassia* stem bark, coumarin was isolated [87]. Coumarin may cause hepatotoxicity. Due to the potential for high levels of coumarin, European health agencies have recently warned against the consumption of large amounts of *C. aromaticum* (cassia) [299]. Theoretically, concurrent use of cinnamon with hepatotoxic herbs may increase the risk of liver damage.
- **Hypoglycemics**: Based on in-vitro and animal evidence, cinnamon has demonstrated lowering of blood glucose levels and acted as an insulin mimetic [6, 18, 29, 41, 56, 95, 121, 185, 200, 208, 223, 225, 228, 229, 252, 271, 309, 344, 367, 390–392, 405, 460, 461, 474, 481, 558]. Human data, however, have demonstrated conflicting results [13, 22, 33, 121, 242, 378, 476, 510]. Theoretically, concurrent use of cinnamon with blood sugar-lowering agents may have additive effects and increase the risk of hypoglycemia.
- **Hypotensives**: *C. migao* oil reduced systolic and diastolic arterial blood pressure in animal study [473]. Human study also demonstrated a reduction in SBP upon treatment with Cinnulin PF[®] [558]. Based on animal study, whole cinnamon and aqueous extracts have been found to reduce SBP elevations as well as a genetic component of elevated blood pressure [386]. Theoretically, concurrent use of cinnamon with antihypertensive agents may have additive effects and increase the risk of hypotension.
- **Immunosuppressants**: Based on in-vitro and animal evidence, cinnamon has demonstrated immunomodulatory effects [244, 342, 343, 444]. Theoretically, cinnamon may alter the effects of immunosuppressants.
- **Insect repellants**: In a clinical trial, (E)-cinnamaldehyde and cinnamyl alcohol appeared to be effective against *Aedes aegypti* (L.) female mosquitoes [65].
- Neurologic herbs and supplements: In animal study, cinnamon may exert an anxiolytic effect via regulation of the serotonergic and GABAergic systems [542]. Theoretically, concurrent use with other herbs or supplements that affect GABA may have additive effects.
- **Phytoestrogens**: Cinnamomi cortex as a component of the unkei-to combination product stimulated estradiol secretion in laboratory study [472]. Theoretically, concurrent use of cinnamon with estrogens may have additive effects.
- **Sympathomimetics**: In animal study, TRPA1 agonists, such as cinnamaldehyde, have been shown to activate the sensory nerves and induce adrenaline secretion via the central nervous system [204]. Theoretically, concurrent use with sympathomimetics may have additive effects.
- **Vitamin E**: In laboratory study, pretreatment with vitamin E markedly prevented cinnamaldehyde-mediated apoptosis [534].

Cinnamon-Food Interactions

- Carrots: In laboratory study, a low concentration of cinnamaldehyde enhanced the taste of carrot broth [506].
- **Foods containing clove**: Based on a review, synergistic antibacterial effects have been observed between cinnamaldehyde and eugenol, a constituent of clove [47].

• **Foods containing vitamin E**: In laboratory study, pretreatment with vitamin E markedly prevented cinnamaldehyde-mediated apoptosis [534].

Cinnamon-Laboratory Interactions

- **Blood glucose**: Based on in-vitro and animal evidence, cinnamon has demonstrated lowering of blood glucose levels and acted as an insulin mimetic [6, 18, 29, 41, 56, 95, 121, 185, 200, 208, 223, 225, 228, 229, 252, 271, 309, 344, 367, 390–392, 405, 460, 461, 474, 481, 558]. Human data, however, have demonstrated conflicting results [13, 22, 33, 121, 242, 378, 476, 510].
- **Blood pressure**: Based on animal study, *C. migao* oil reduced systolic and diastolic arterial blood pressure [473]. Based on animal study, whole cinnamon and aqueous extracts have been found to reduce SBP elevations as well as a genetic component of elevated blood pressure [386]. Human study also demonstrated a reduction in SBP upon treatment with cinnamon [558].
- Carbon clearance test: A polysaccharide isolated from the bark of *C. cassia* Blume was found to exert reticuloendothelial system-potentiating activity in a carbon clearance test [221].
- **Coagulation panel**: Based on animal evidence, cinnamon bark, cinnamaldehyde, and two other *Cinnamomum* species (*C. altissimum* and *C. pubescens*) caused a decrease in platelet counts after long-term use [193, 206, 320, 367]. Cinnamic aldehyde inhibited AA release and thromboxane B2 formation, which may contribute to reduced platelet aggregation [480].
- **Estrogens**: Cinnamomi cortex as a component of the unkei-to combination product stimulated estradiol secretion in laboratory study [472].
- **Heart rate**: In animal study, various cinnamon species [77, 466, 473, 543, 553], including *C. migao* [471], decreased the heart rate.
- Lipid profile: Based on animal evidence, *C. zeylanicum* significantly decreased serum total cholesterol and triglyceride concentrations and markedly increased HDL cholesterol levels [273, 467]. In animal study, cinnamate, a phenolic compound in cinnamon bark, produced higher HDL cholesterol levels and lower atherogenic index compared with lovastatin [273]. Cinnamate inhibited HMG-CoA reductase activity [273]. In in-vivo evidence in hamsters, a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels and inhibited CD36 and MTTP [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats when included in the diet at about fivefold the normal human intake level [420].
- **Urate levels**: Oral administration of *C. cassia* oil significantly reduced serum and hepatic urate levels in hyperuricemic mice [552]. In normal mice, urate levels in liver, but not in serum, were altered with dose-dependent decrease after *C. cassia* oil treatment.

Cinnamon-Nutrient Depletion

• Glucose: Based on in-vitro and animal evidence, cinnamon has demonstrated lowering of blood glucose levels and acted as an insulin mimetic [6, 18, 29, 41,

- 58, 95, 121, 185, 200, 208, 223, 225, 228, 229, 252, 271, 309, 344, 367, 390–392, 405, 460, 461, 474, 481, 558]. Human data, however, have demonstrated conflicting results [13, 22, 33, 121, 242, 378, 476, 510].
- **Lipids**: Based on animal evidence, *C. zeylanicum* significantly decreased serum total cholesterol and triglyceride concentrations and markedly increased HDL cholesterol levels [273, 467]. In animal study, cinnamate, a phenolic compound in cinnamon bark, produced higher HDL cholesterol levels and lower atherogenic index compared with lovastatin [273]. Cinnamate inhibited HMG-CoA reductase activity [273]. In in-vivo evidence in hamsters, a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels and inhibited CD36 and MTTP [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats when included in the diet at about fivefold the normal human intake level [420].

MECHANISM OF ACTION

Pharmacology

- Constituents: Cinnamon has been shown to contain allylbenzenes and their isomers, the propenylbenzenes [198]. Cinnamon also contains monomeric and oligomeric proanthocyanidins [161, 268], e.g., procyanidin B-2 and procyanidin B-3 [383, 482], and organic compounds *trans*-cinnamaldehyde, alpha-amyl cinnamaldehyde, and *trans*-cinnamic alcohol [124]. Quercetin, kaempferol, luteolin, and pelargonidin have been identified as the major flavonoids [211, 255, 345, 366]. Inorganic constituents of Cinnamomi cortex include potassium, calcium, iron, manganese, and strontium. A feature of the metals profile of Cinnamomi cortex is high manganese content [330].
- Cinnamon species contain volatile oils [4, 147, 282, 489]. At least 94 volatile components are present in cinnamon bark [158]. Fifty-four constituents were identified in the essential oil from cinnamon bark and twigs [450]. The main components of the essential oil obtained from the bark of *C. zeylanicum* are eugenol, cinnamaldehyde, and linalool [83, 146, 169, 196, 445, 457, 458, 502, 503]. Each cinnamon plant part has a different primary constituent: cinnamaldehyde (bark oil), eugenol (leaf oil), and camphor (root-bark oil) [293, 311, 332, 430, 443, 528]. *C. cassia* bark contains cinnamaldehyde, cinnamic acid, cinnamyl alcohol, coumarin, 5,7,3′,4′-tetrahydroxyflavan-3,4-diol, OMCA, lignans, and phenylpropanoids [42, 45, 163, 188, 237, 296, 333, 335]. Other *Cinnamomum* species, *C. wilsonii*, *C. japonicum*, *C. mairei*, and *C. burmannii*, contain low contents of cinnamaldehyde (< 2.00 mg/g) [180].
- The leaves of *C. kotoense* contain isoobtusilactone A, cinnakotolactone, isolinderanolide B, kotomolide A, kotomolide B, isokotomolide A (IKA), secokotomolide A, and secobutanolide [70, 71, 74, 75, 293, 536].
- The leaves of *C. subavenium* contain subamolides A–E, secosubamolide A, as well as 21 known compounds [72, 254, 289].
- Constituents of *C. balansae* leaves include cinbalansan [6]; 1,2-dimethoxy-4-(1-E-propenyl)benzene; 1,2-dimethoxy-4-(1-Z propenyl)benzene; 1,2-dimethoxy-4-(1-Z propenyl)be

- dimethoxy-4-(2-propenyl)benzene; 3,4 dimethoxybenzaldehyde; and E-(3,4-dimethoxyphenyl)-2-propenal [96].
- The stems of *C. tenuifolium* contain tenuifolide A, isotenuifolide A, tenuifolide B, secotenuifolide A, and tenuifolin [288].
- C. insularimontanum contains actinodaphnine [191].
- The fruit of *C. laubatii* contains EBC-23, 24, 25, 72, 73, 75, and 76 [111, 112].
- The volatile oil from C. zeylanicum fruit stalks contains hydrocarbons (44.7%) and oxygenated compounds (52.6%). Twenty-seven compounds constituting approximately 95.98% of the volatile oil have been characterized. (E)-Cinnamyl acetate (36.59%) and (E)-caryophyllene (22.36%) were found to be major compounds [210]. C. zeylanicum buds contain 34 compounds representing approximately 98% of the oil and consist of terpene hydrocarbons (78%) and oxygenated terpenoids (9%). Alpha-bergamotene (27.38%) and alpha-copaene (23.05%) were found to be the major compounds [212]. The steam-distilled oil of C. zeylanicum flowers consists of 23% hydrocarbons and 74% oxygenated compounds. A total of 26 compounds constituting approximately 97% of the oil have been characterized. (E)-Cinnamyl acetate (41.98%), trans-alpha-bergamotene (7.97%), and caryophyllene oxide (7.2%) were found to be major compounds [209]. The essential oil isolated from C. osmophloeum leaves contains six chemotypes: cinnamaldehyde type, cinnamaldehyde/cinnamyl acetate type, cinnamyl acetate type, linalool type, camphor type, and mixed type [79, 82, 132]. The major constituents of C. osmophloeum leaf essential oil are the monoterpenes, 1,8cineole (17.0%) and santolina triene (14.2%), and the sesquiterpenes, spathulenol (15.7%) and caryophyllene oxide (11.2%) [67]. The lignan sesamin has been isolated from C. kanehirae [192].
- Acaricidal effects: Cinnamon displayed caricidal activity against red mites (*Dermanyssus gallinae*) [235]; however, the mechanism of action is not well understood.
- Adrenergic effects: TRPA1 agonists, such as cinnamaldehyde, have been shown to activate the sensory nerves and induce adrenaline secretion via the central nervous system [204].
- **Alzheimer's effects**: Cinnamon extracts have been shown to inhibit tau aggregation and filament formation; a significant portion of the effect attributed to an A-linked proanthocyanidin trimer molecule [377].
- Analgesic effects: An ethanolic extract of *C. zeylanicum* was shown to possess an antinociceptive effect against both acetic acid-induced writhing and hot plate-induced thermal stimulation in mice [21]. However, cinnamaldehyde is a specific TRPA1 (mammalian transient receptor potential [TRP] ion channel) activator, and has been shown to excite a subset of sensory neurons highly enriched in cold-sensitive neurons and to elicit nociceptive behavior in mice [25, 324]. Secondary sources indicate that cinnamon may contain the following potentially bioactive constituents: ascorbic acid, borneol, caffeic acid, camphor, caryophyllene, coumarin, eugenol, linalool, mannitol, myrcene, P-cymene, phenol, thiamin, and zinc. These constituents have been proposed to have analgesic properties, although the exact mechanisms of action for each constituent are unclear.
- **Antibacterial effects**: Extracts of cinnamon, as well as the major components cinnamaldehyde and eugenol, have demonstrated activity against *Campylobacter*

jejuni, E. coli, Listeria monocytogenes, B. subtilis, Salmonella spp., Morganella morganii, Clostridium perferingens, B. cereus, and Staphylococcus aureus in vitro [59, 99, 143–145, 153, 182, 243, 259, 357, 436, 441, 459, 507, 518, 541, 544, 545]. Cinnamaldehyde exhibits bactericidal activity against Listeria monocytogenes. Inhibition of energy generation may be due to the inhibition of glucose uptake or utilization of glucose and effects on membrane permeability [153]. Cinnamaldehyde inhibited the swimming motility of E. coli in laboratory study. Cinnamaldehyde reduced biofilm formation by E. coli ATCC 33456 in part by interfering with its ability to reach the substratum [357]. Cinnamaldehyde has also demonstrated antimicrobial effects against BacPre-targeted ruminal bacteria [136]. Based on review, synergistic antibacterial effects have been observed between cinnamaldehyde and eugenol [47].

- Cinnamon bark oil, the alcoholic extract, and its major components showed antibacterial effects on the major respiratory and gastrointestinal tract pathogens *Haemophilus influenzae*, *Streptococcus pneumoniae*, *Streptococcus pyogenes*, *S. aureus*, *Porphyromonas gingivalis*, *E. coli*, *B. cereus*, and *S. aureus* in vitro [143, 144, 201, 202, 369].
- Cinnamon oil was shown to have antibacterial activity against *B. cereus*, *E. coli*, and *S. aureus* in the following order: *B. cereus* (vegetative) ≫ *S. aureus* approximately *E. coli* ≫ *B. cereus* (spores) [143].
- *C. zeylanicum* tincture did not exhibit antibacterial properties in laboratory study [522].
- Cinnamon has antimicrobial properties that may improve energy or protein use in rumen and may therefore act as an alternative to ruminal modifiers. Cinnamon affected molar proportions of acetate, propionate, and butyrate between Day 2 and Day 6 of fermentation during the adaptation period. The accumulation of peptide nitrogen and the numerical decrease in amino acid nitrogen in cinnamon suggest that peptidolysis was inhibited [58, 310].
- Cinnamaldehyde has not been found to significantly inhibit multidrug resistance proteins MRP1 or MRP2 [532]. Extracts of cinnamon, as well as the major components cinnamaldehyde and eugenol, have demonstrated activity against Campylobacter jejuni, E. coli, Listeria monocytogenes, B. subtilis (ATCC 6633), and Salmonella enterica in vitro [59, 99, 145, 243, 436, 459, 544, 545]. Cinnamon bark oil and its major components showed antibacterial effects on the major respiratory and gastrointestinal tract pathogens Haemophilus influenzae, Streptococcus pneumoniae, Streptococcus pyogenes, S. aureus and Porphyromonas gingivalis, in vitro [201, 202, 369]. Furthermore, cinnamon exhibited significant inhibitory effects both in vitro and in vivo on M. morganii [441]. However, one study found no antibacterial activity in C. zeylanicum tincture using luminescent bacterial biosensors (E. coli strains) [522]. A case report studied an exclusively breastfed infant (4 months of age) with acute diarrhea and who became a chronic carrier of S. enteritidis [409]. The infant was administered ground cinnamon bark in homogenized fruit 3-4 times a day. One month later, stool samples of the infant tested negative for S. enteritidis; repeat tests, 2 and 4 months later, were also negative. In human trial using an alcoholic extract of cinnamon for H. pylori infection, the amount of *H. pylori* colonization measured by the 13C urea breath test served as the outcome measure [354]. The mean urea breath test counts in

- the study and control groups before and after therapy were 22.1 and 23.9 versus 24.4 and 25.9, respectively. The exact mechanism of action remains unclear.
- Cinnamon [300, 371, 380, 486], cinnamon extracts [61, 168, 246, 331, 442], cinnamon essential oils (alone and in combination with other plant oils) [28, 107, 118, 130, 149, 181, 296, 297, 321, 338, 339, 361, 370, 382, 400, 401, 406, 436, 458, 497, 520, 538], and constituents of cinnamon such as cinnamaldehyde (alone and in combination with other plant extracts) [20, 62, 64, 110, 137, 164, 215, 363, 398, 399, 408, 425, 456, 495, 505] and trans-cinnamaldehyde [14, 16, 68, 94, 423, 530] have all been reported to exhibit antimicrobial effects, although a tincture of *C. zeylanicum* was reported as ineffective against *E. coli* [522]. Cinnamaldehyde was found to induce multiple antibiotic resistance (MAR) in *Bacteroides fragilis* [387]. Cinnamaldehyde has also been reported to interfere with quorum sensing [38, 356] and cinnamon essential oil to interact synergistically with clindamycin [440].
- Anticancer/antitumor effects: Cinnamon and its constituents have displayed anticancer and chemopreventive properties in various studies [166, 171, 265, 355, 534]. The antitumor activity of *Cinnamomum* cortex is considered to be based on stimulation of the reticuloendothelial system (RES) and has been shown to be closely related to TNF (tumor necrosis factor) production [171]. A genotoxicity assay (micronucleus test) demonstrated dose-related antigenotoxic effects after urethane was coadministered orally with an aqueous extract of cinnamon to mice [8]. *C. cassia* induced the death of HL-60 cells, demonstrated by reduction in mitochondrial transmembrane potential and increase in caspase-3 activity [355]. Cinnamaldehyde derivative CB403 exerted cytostatic properties through the arrest of cell cycle progression in the G₂/M phase in laboratory study [213]. The bark of *C. cassia* displayed inhibitory effects against matrix metalloproteinase-2 and -9 (MMP-2 and -9) and invasion of SK-Hep1 hepatoma cells [166].
- Cinnamaldehyde is also a potent inducer of apoptosis. It has been shown to transduce the apoptotic signal via reactive oxygen species (ROS) generation, thereby inducing mitochondrial permeability transition (MPT) and cytochrome c release into the cytosol. Thus, the anticancer effects of cinnamaldehyde may result from the induction of ROS-mediated mitochondrial permeability transition and resultant cytochrome c release [216, 534]. Cinnamic aldehyde inhibited leukemia L1210 cells by blocking protein synthesis through trapping sulfhydryl-containing amino acids in the cell [334]. Cinnamaldehyde also upregulated the expression of pro-apoptotic protein (Bax) and downregulated the levels of antiapoptotic proteins such as Bcl-2, the inhibitor of apoptosis protein family (X-linked inhibitor of apoptosis protein [XIAP]), and the cellular inhibitor of apoptosis protein (cIAP-1 and cIAP-2) [534]. Of note, pretreatment with vitamin E markedly prevented cinnamaldehyde-mediated apoptosis, which was associated with the modulation of XIAP, cIAP-1, cIAP-2, Bcl-2, and Bax protein activity [534].
- A strong MMP-9 inhibition was found in the butanol fraction of *C. cassia* [438]. MMP-9 degrades type IV collagen, constituting the major structural component of the basement membrane and the extracellular membrane; the enzymatic activity is found to be elevated in tumor tissues. 2'-Hydroxy-cinnamaldehyde and 2'-benzoyloxycinnamaldehyde isolated from *C. cassia* strongly inhibited in-vitro growth of 29 kinds of human cancer cells and in-vivo growth of SW-620 human tumor xenograft in nude mice. HCA prevented adherence of SW-620 cells to the

culture surface but did not inhibit oncogenic K-Ras processing, implying its antitumor mechanisms at the cellular level [269]. HCT15 and SK-MEL-2 cells were very sensitive to the cinnamaldehyde analogs cinnamic acid, cinnamates, and cinnamyl alcohols [257]. Based on in-vitro evidence, polymeric polyphenols from cinnamon have been shown to block the G_2/M phase of the cell cycle by interacting with phosphorylation/dephosphorylation signaling activities in three myeloid cell lines (Jurkat, Wurzburg, and U937) and a leukemic cell line [427, 428].

- Cinnamon [30, 258], cinnamon extracts and essential oils [446], and constituents of cinnamon [112, 288], including a *Cinnamomum* monoterpenoid [289], cinnamaldehyde [120, 239, 448, 533], isoobtusilactone A [71, 293], kotomolide A [253], subamolides A–C [72] and subamolides D and E [254], isokotomolide A [73], cinnamyl compounds related to 2'-hydroxycinnamaldehyde [454], 2'-hydroxycinnamaldehyde [188], and isoobtusilactone A [74] have all been reported to exhibit anticancer, antitumor, antiproliferative, or antimutagenic effects.
- Antidiabetic effects: Based on human and animal study, cinnamon has been used to control blood sugar either alone or in combination with other essential oils such as fenugreek and oregano [6, 13, 33, 95, 121, 229, 309, 344, 378, 474, 481, 510, 558]. However, results have not been consistent, and various human trials have found that cinnamon did not significantly alter A1C, FBG, or lipid parameters in patients with type 1 or type 2 diabetes [22, 242]. Pharmacological studies have shown that cinnamon may play a possible role in improving glucose and insulin metabolism [367], but that these effects are short lived (i.e., up to 12 hr) once cinnamon feeding is stopped [460, 461].
- In animal and laboratory studies, cinnamon has been shown to potentiate the insulin effect through upregulation of the glucose uptake in cultured adipocytes and to potentiate insulin-regulated glucose utilization by enhancing the insulinsignaling pathway in skeletal muscle [18, 29, 41, 56, 200, 228, 390].
- Cinnamon was highly active in the insulin-dependent utilization of glucose using a rat epididymal adipocyte assay. In animal study, cinnamon prevented insulin resistance in rats fed a high-fructose diet in part by enhancing insulin signaling and possibly via the nitric oxide (NO) pathway in skeletal muscle [391]. A hydroxychalcone from cinnamon functioned as an insulin mimetic in 3T3-LI adipocytes [208]. Furthermore, in-vitro evidence showed that adipocytes exposed to 0.2 mg/ml of cinnamon extract in the absence of insulin showed an approximate twofold increase in glucose uptake relative to controls [405]. Cinnamaldehyde exhibited strong inhibition against aldose reductase [271], an enzyme in carbohydrate metabolism that converts glucose to its sugar alcohol form, sorbitol, using NADPH as the reducing agent. Aqueous extracts of cinnamon significantly lowered the absorption of alanine, an important amino acid for gluconeogenesis, from the rat intestine [252]. Blood glucose-lowering effects within 2 weeks have been shown for C. tamala in alloxan diabetic albino rats [225]. However, another pharmacological study demonstrated that consumption of diets containing C. tamala did not alter diabetes parameters in streptozotocin diabetic mice [476].
- Kannappan et al. showed that cinnamon extract improved glucose metabolism in vivo in fructose-fed rats [223]. Qin et al. showed that cinnamon extract

improved insulin action via increasing glucose uptake in vivo possibly through enhancement of the nitric oxide pathway in skeletal muscle [390, 391]. Based on human evidence, ingestion of 3 g of cinnamon reduced postprandial serum insulin and increased glucagon-like peptide 1 (GLP-1) concentrations without significantly affecting blood glucose, glucose-dependent insulinotropic polypeptide (GIP), ghrelin concentration, satiety, or gastric emptying rate (GER) [185]. Additional study has shown that addition of cinnamon to rice pudding significantly delayed gastric emptying and lowered the postprandial glucose response, with no effects on satiety in healthy subjects [184].

- Antifungal effects: Based on human and in-vitro study, cinnamon oil, cortex cinnamon solution, *trans*-cinnamaldehyde, cinnamaldehyde, and the essential oils have been found to have inhibitory effects against several fungi, including *Candida*, *Coriolus versicolor*, *Laetiporus sulphureus*, *Eurotium* spp., *Aspergillus* spp., and *Penicillium* [55, 79, 85, 165, 174, 285, 294, 317, 335, 374, 457, 504, 541]. High concentrations of cinnamon, however, have been shown to stimulate mycelial growth of *A. flavus* [301]. *C. zeylanicum* has shown potent in-vitro activity against fluconazole-resistant and -susceptible *Candida* isolates [393]. Cinnamaldehyde and *trans*-cinnamaldehyde have also displayed strong activity against various types of fungi [515]. Colonies formed by heat-stressed cells of *S. cereviseae* showed a reduction in size, and heated *Rhodotorula rubra* cells demonstrated a slight increase in sensitivity to cinnamon oil [92].
- **Anthelmintic effects**: Ceylon cinnamon reportedly has anthelmintic effects; however, these properties are not well documented [60].
- Anti-inflammatory effects: Cinnamon bark showed anti-inflammatory properties in vitro [256, 402] and in vivo in the carbon clearance test [221]. Eugenol and cinnamaldehyde were found to inhibit cyclooxygenase-2 (COX-2) in vitro in a rapid semi-homogeneous COX-2 enzymatic assay [195]. Cinnamaldehyde has been shown to inhibit 5-lipoxygenase, a key enzyme involved in the biosynthesis of leukotrienes [384]. This effect was more potent than that of piperine, capsaicin, and allyl sulfide, but less potent than that of quercetin, eugenol, and curcumin. C. massoiae cortex extract inhibited IgE-dependent histamine release [199]. Extracts obtained from C. osmophloeum leaf essential oil and from the twigs of C. osmophloeum Kaneh. have shown in-vitro anti-inflammatory activity [67, 132, 496]. Cinnamaldehyde has been demonstrated to possess anti-inflammatory properties [66, 274]; to alleviate neuropathic and inflammatory pain via antagonism of TRPA1 [123]; to suppress TNF-induced singling pathways [283], to activate nuclear factor (NF)-kappaB and interferon regulatory factor 3 (IRF3) induced by lipopolysaccharides (LPS), a toll-like receptor (TLR4) agonist, leading to the decreased expression of target genes such as COX-2 and interferon (IFN)beta [540]; to reduce expression of inflammatory cytokines (in combination with other plant extract) [281]; and to inhibit age-related activated NF-kappaB upregulation of NF-kappaB targeting genes, inflammatory inducible nitric oxide (iNOS), and COX-2 via three signal transduction pathways (NIK/IKK, ERK, and p38 MAPK) [231]. Cinnamon essential oils [287] and extracts [396] have also been documented as having anti-inflammatory action.
- Components of Cinnamomi ramulus have demonstrated an anti-inflammatory effect via inhibition of the expression of iNOS and COX-2 and by suppressing

nitric oxide (NO) production in both the peripheral and the central nervous systems [197]. Cinnamon bark extract has been suggested as possibly protecting the liver from acute alcohol-induced steatosis through the inhibition of MyD88 expression [224]. Research has also suggested that cinnamon polyphenol extract may affect immune responses by regulating inflammatory dynamics and glucose transporter (GLUT) gene expression [57].

- Secondary sources indicate that cinnamon may contain the following potentially bioactive constituents: (–)-epicatechin, 1,8-cineole, alpha-pinene, alphaterpineol, ascorbic acid, beta-pinene, beta-sitosterol, borneol, caffeic acid, caryophyllene, caryophyllene oxide, cinnamaldehyde, cinnamic acid, copper, coumarin, delta-3-carene, eugenol, eugenyl acetate, isoeugenol, limonene, linalool, mannitol, oleic acid, oligomeric proanthocyanidins (OPCs), salicylates, and stigmaterol. These constituents have been proposed to have anti-inflammatory properties, although the exact mechanisms of action for each constituent are unclear.
- Antilipemic effects: Based on animal evidence, *C. zeylanicum* significantly decreased serum total cholesterol and triglyceride concentrations and markedly increased HDL cholesterol levels [273, 467]. In animal study, cinnamate, a phenolic compound in cinnamon bark, produced higher HDL cholesterol levels and lower atherogenic index compared with lovastatin. Cinnamate inhibited HMG-CoA reductase activity [273]. In-vivo evidence in hamsters has shown that a water extract of cinnamon (Cinnulin PF®) inhibited the postprandial overproduction of apoB48-containing lipoproteins and serum triglyceride levels [389]. However, these results were in contrast to another study that did not show any cholesterol-lowering effect in serum and liver cholesterol levels of rats [420]. Further in-vitro evidence from this study showed that cinnamon extract treatment inhibited CD36 and MTTP [389].
- Research has suggested that postprandial hypertriglycerides and overproduction of apoB48 may be acutely inhibited by cinnamon extract via improved insulin sensitivity of intestinal enterocytes and regulation of MTTP and SREBP1c levels [392]. Cinnamaldehyde has been observed to reduce serum total cholesterol and triglyceride levels in rats [467].
- Antimutagenic effects: The antimutagenic activity of cinnamon and its constituents have been reported in various laboratory studies [8, 102, 178, 207, 217, 218, 447]. *C. cassia* exerted significant antimutagenic effects against benzo[a]pyrene (B[a]P) and cyclophosphamide in mice pretreated with the plant extract, as was observed in the Ames test, bone marrow chromosomal aberration assays, and micronucleus tests [447]. In one laboratory study, *trans*-cinnamaldeyde showed antimutagenic activity against select *Salmonella typhimurium* tester strains TA1535 (hisG46 uvrB rfa) and TA100 (TA1535/pKM101) [102]; however, other evidence revealed no mutagenic effects [284]. *C. cassia* pretreatment decreased liver cytochrome P450 content but increased glutathione content and the activity of glutathione-dependent antioxidant enzymes glutathione S-transferase, glutathione reductase, and glutathione peroxidase. These findings might demonstrate that the antimutagenic potential of *C. cassia* may be attributed to its modulatory effect on xenobiotic bioactivation and detoxification processes. α,β-unsaturated carbonyl compounds in

- (E)-2-cinnamaldehyde were found to induce oxidative purine modifications (formamidopyrimidine DNA glycosylase [FPG]-sensitive sites) in mammalian cells, in addition to direct DNA breakage [207, 364]. Cinnamyl anthranilate enhanced SA7 transformation; cinnamaldehyde produced some evidence of enhanced activity [178].
- The antimutagenic effects of cinnamaldehyde have been evaluated in various studies [364, 365]. Cinnamaldehyde suppressed the umuC-dependent mutagenesis induced by 4-nitroquinoline 1-oxide (4-NOO), furvlfuramide, or captan, but was less effective against the umuC-independent mutagenesis by alkylating agents such as N-methyl-N'-nitro-N-nitrosoguanidine and ethylmethanesulfonate [364]. Cinnamaldehyde did not affect prophage induction or tif-mediated filamentous growth, suggesting it does not prevent the induction of the SOS functions [364]. An increase was observed in the survival of 4-NOO-treated WP2s cells after exposure to cinnamaldehyde, suggesting the promotion of some DNA repair system by cinnamaldehyde [364]. An enhancement in survival was also observed in uvr B, polA, recF, or umuC mutants and less in lexA or recBC mutants; it was not observed in recA mutants [364]. Cinnamaldehyde may enhance an error-free recombinational repair system by acting on recA-enzyme activity [364]. Cinnamaldehyde displayed antimutagenic effects against mutations induced by UV-mimic mutagens but not those induced by -methyl-N'-nitro-Nnitrosoguanidine or ethyl methanesulfonate, and may therefore act by interfering with an inducible error-prone DNA repair pathway [365].
- Antioxidant effects: Cinnamon and cinnamon bark have been shown to contain very high concentrations of antioxidants [34, 115, 300]. Several animal in-vitro studies have demonstrated the antioxidant effects of the essential oil obtained from the bark of C. zeylanicum and its main components [18, 83, 175, 210, 236, 272, 278, 455, 491]. In animal study, cinnamate suppressed lipid peroxidation by enhancing hepatic antioxidant enzyme activities [273]. It elevated catalase and glutathione peroxidase activity and reduced thiobarbituric acid-reactive substances [273]. Cinnamon extracts [88, 119, 336], essential oils [418, 458, 513, 523], bark [383], and cinnamaldehyde [66, 159] have all been identified as possessing antioxidant properties. Cinnamon has also been found to synergistically increase the antioxidant capacity of some teas [48], protect against peroxynitirite damage [186], and scavenge NO (nitric oxide) [494]. In addition, constituents of cinnamon, such as cinnamic aldehyde and methyl-1-cinnamoyl-5-oxo-2pyrrolidine-carboxylate, have been identified as potent nuclear factor-erythroid 2 (Nrf2)-activators (involved in cellular antioxidant response) [529]. Furthermore, cinnamon extracts have been observed to significantly increase levels of reduced glutathione and the activities of glutathione reductase, glutathione S-transferase, glutathione peroxidase, catalase, and superoxide dismutase in the liver of rats [233]. In other research, cooking and storage has been found to reduce the antioxidant capacity of cinnamon extract [86].
- In one study, cinnamon showed higher antioxidant activity and was a better superoxide radical scavenger than anise, ginger, licorice, mint, nutmeg, and vanilla [340].
- In various studies, etheric, methanolic, and aqueous cinnamon extracts also inhibited oxidative processes in vitro [106, 187, 248, 275, 286, 308, 318, 451].

- A randomized controlled trial (RCT) examined the effects of a dried aqueous extract of cinnamon (Cinnulin PF[®]) on the antioxidant status of overweight or obese individuals with impaired fasting glucose [411]. Plasma antioxidant status and plasma thiol (SH) increased, while plasma malondialdehyde levels decreased in subjects receiving the cinnamon extract. A positive correlation was also observed between malondialdehyde and plasma glucose (r = 0.74, p = .014).
- Ethanol extracts of dry bark of *C. cassia* (96.30%) exhibited a greater inhibition of FeCl₂–ascorbic acid-induced lipid peroxidation of rat liver homogenate in vitro than that by alpha-tocopherol (93.74%) [286]. A highly positive linear relationship was obtained between total equivalent antioxidant capacity (TEAC) values and total phenolic content; the authors conclude that phenolic compounds in *C. zeylanicum* and *C. cassia* bark may contribute significantly to their antioxidant capacity [443].
- Suganthi et al. showed that a spice mixture containing 1.0 g/100 g of cinnamon bark increased levels of peroxidation indices such as thiobarbituric acid-reactive substances (TBARS) and concentration of lipid hydroperoxides in tissues [470].
- Secondary sources indicate that cinnamon may contain the following potentially bioactive constituents: (–)-epicatechin, acetyl eugenol, ascorbic acid, beta carotene, caffeic acid, campesterol, camphene, eugenol, gamma terpinene, isoeugenol, lauric acid, linalyl acetate, manganese, mannitol, methyl eugenol, myrcene, myristic acid, OPCs, P-coumaric acid, palmitic acid, phenol, proanthocyanidins, riboflavin, stigmasterol, terpinen-4-ol, terpinolene, and vanillin. These constituents have been proposed to have antioxidant properties, although the exact mechanisms of action for each constituent are unclear.
- Antipyretic effects: The active compounds of guizhi tang, cinnamaldehyde, and cinnamic acid have been found to have antipyretic action by inducing EP3 prostaglandin receptors [280].
- Antiseptic effects: Cinnamaldehyde has been shown to be an effective periodontal disinfectant, significantly reducing levels of oral endotoxin in rats [292]. Secondary sources indicate that cinnamon may contain the following potentially bioactive constituents: 1,8-cineole, alpha-pinene, alpha-terpineol, ascorbic acid, benzaldehyde, benzyl alcohol, beta-pinene, caffeic acid, camphor, chlorine, citronella, eugenol, furfural, geraniol, hexanol, iodine, limonene, linalool, methyl eugenol, nerol, OMCA, oxalic acid, P-coumaric acid, phenol, proanthocyanidins, sabinene, safrole, sulfur, terpinen-4-ol, and zinc. These constituents have been proposed to have antiseptic properties, although the exact mechanisms of action for each constituent are unclear.
- Antispasmodic effects: Secondary sources indicate that cinnamon may contain the following potentially bioactive constituents: 1,8-cineole, alpha-pinene, alphaterpinene, ascorbic acid, benzaldehyde, benzyl benzoate, beta-pinene, borneol, bornyl acetate, caffeic acid, camphor, caryophyllene, cinnamaldehyde, cinnamic acid, eugenol, eugenyl acetate, farnesol, geraniol, limonene, linalool, linalyl acetate, mannitol, myrcene, niacin, P-coumaric acid, potassium, and terpinen-4-ol. These constituents have been proposed to have antispasmodic properties, although the exact mechanisms of action for each constituent are unclear.
- Antiviral effects: C. cassia bark extract has been highly effective against HIV-1 and HIV-2 replication in terms of inhibition of virus induced cytopathogenicity

in MT-4 cells infected with HIV [385]. Cinnamaldehyde derived from cinnamon bark has shown an inhibitory effect on the growth of influenza A/PR/8 virus in vitro and in vivo [179]. Cinnamon extract have been shown to prevent HIV-1 infection in vitro, an effect attributed to the extract's flavonoid content [139]. Cinnzeylanine, a constituent of cinnamon, has also been shown to inhibit herpes simplex virus proliferation in vitro [368]. Another constituent, trans-cinnamaldehyde, has been shown to have a similar inhibitory effect on influenza A/PR/8 viral growth [179]. Secondary sources indicate that cinnamon may contain the following potentially bioactive constituents: (–)-epicatechin, alpha-pinene, ascorbic acid, beta-sitosterol, bornyl acetate, caffeic acid, chlorine, cinnamaldehyde, eugenol, geranial, iodine, lauric acid, limonene, linalool, OPCs, P-cymene, phenol, proanthocyanidins, stigmasterol, and vanillin. These constituents have been proposed to have antiviral properties, although the exact mechanisms of action for each constituent are unclear.

- Anxiolytic effects: As indicated by research in mice, cinnamon may exert an anxiolytic effect via regulation of the serotonergic and GABAergic systems [542].
- Cardiovascular effects: C. cassia bark has been shown to affect the blood and cardiovascular system [77]. It has been shown to reduce blood pressure in animal studies either alone or in combination with other essential oils such as fenugreek and oregano [386, 481]. C. cassia increased the level of atrial natriuretic factor (ANF) in the plasma of mice [553]; ANF acts to reduce the water, sodium, and adipose loads on the circulatory system, thereby reducing blood pressure. Another human trial also demonstrated lowered SBP upon treatment with a watersoluble cinnamon extract (-3.8%: from 133 ± 14 mmHg [pre] to 128 ± 18 mmHg [post], p < .001) [558]. In experimental arrhythmia, C. migao reduced the incidence of ventricular fibrillation caused by chloroform in mice and the ventricular tachycardia induced by adrenalin in rabbits, delayed the onset time of this arrhythmia, increased the arrhythmic doses of strophantin-K in guinea pigs, reduced the incidence of some arrhythmias caused by barium chloride in rats, and slowed down their heart rate [471]. C. migao oil reduced systolic and diastolic arterial blood pressure, slowed down the heart rate, decreased carbon monoxide levels, and reduced left ventricular pressure in anesthetized open-chest cats after i.d. application [473]. Cinnamophilin, a thromboxane A(2) antagonist isolated from C. philippinense, inhibited sodium inward current, calcium inward current, and transient outward current in rat cardiac tissue and converted episodes of ischemia-reperfusion arrhythmia to normal sinus rhythm [466]. Cinnamophilin dose-dependently inhibited human platelet-rich plasma (PRP) aggregation induced by AA, collagen, and U-46619 [543].
- Cell cycle effects: Cinnamaldehyde has been shown to promote more cells in G_0/G_1 phase into S phase [551].
- Coagulation effects: Based on animal evidence, cinnamon bark, cinnamaldehyde, and two other *Cinnamomum* species (*C. altissimum* and *C. pubescens*) caused a decrease in platelet counts after long-term use [193, 206, 320, 367]. Cinnamic aldehyde inhibited AA release and thromboxane B2 formation, which may contribute to reduced platelet aggregation [480].

- **Cytochrome P450 effects**: 5'-hydroxy-5-hydroxymethyl-4",5"-methylenedioxy-1,2,3,4-dibenzo-1,3,5-cycloheptatriene, a constituent of cinnamon extract, has been shown to inhibit CYP3A4 in vitro [469].
- Enzymatic effects: ATPases are a class of enzymes that catalyze the decomposition of adenosine triphosphate (ATP) into adenosine diphosphate (ADP) and a free phosphate ion. Water extracts of cinnamon inhibited the activity of rat liver Na+/K+ ATPase and Cu²+ ATPase but, as did cinnamaldehyde and eugenol, stimulated rat mitochondrial F₀F₁ATPase, reduced mitochondrial membrane potential, inhibited NADH oxidase or Complex I of the respiratory chain, and had no effect on succinate dehydrogenase activity [502, 503]. These effects resulted in a decrease in ATP level; defects in proton and ion transports, leading to electrolyte imbalance; and derangements in mitochondrial function. Furthermore, cinnamon water extract most potently inhibited the in-vitro activity of the rat jejunal Na+-/K+-ATPase, the in-vitro Na+-/K+-ATPase activity in a crude kidney homogenate, and the activity of an isolated dog kidney Na+-/K+-ATPase. The alcoholic extract of cinnamon, compared with the aqueous extract, had a stronger inhibitory action on the jejunal enzyme, as did cinnamaldehyde. Eugenol is the major inhibitory component in both alcoholic and aqueous extracts [252].
- Cinnamon extracts [350], essential oils [313], and constituents, including *trans*-cinnamaldehyde thiosemicarbazone [556], and cinnamic acid [249] have all been identified as inhibiting tyrosinase. Cinnamon essential oils have also been identified as inhibiting xanthine oxidase [516]. In addition, cinnamon extracts have been found to inhibit PKA [337] and elastase [276].
- **Gastrointestinal effects**: Chinese cinnamon (the stem bark of *C. cassia*) prevented serotonin-induced ulcerogenesis and inhibited gastric ulcers in rats after oral administration [483]. In a pharmacological study, Chinese cinnamon inhibited the secretion of gastric acid to a small extent but promoted gastric blood flow [10].
- Cinnamon reportedly contains lipase, a digestive enzyme [177]. In poorly documented research, however, Cassia nomame extracts appear to have lipase-inhibiting activity [322].
- In animal study, cinnamic aldehyde, eugenol, and eugenol acetate exhibited relaxant effects on tracheal and ileal smooth muscles [403].
- TRPA1, of which cinnamaldehyde is an agonist, has been shown to act as a sensor molecule for enterochromaffin cells and may regulate gastrointestinal function [360]. Cinnamaldehyde has been shown to induce intestinal HCO₃(–) secretion, likely via direct activation of nicotinic receptors on epithelial cells [36]. In other gastrointestinal research, cinnamon extract has been shown to have antidiarrheal effects [395].
- **Hormonal effects**: Cinnamomi cortex as a component of the unkei-to combination product was found to stimulate estradiol secretion in laboratory study [472].
- **Hypouricemic effects**: Oral administration of *C. cassia* oil significantly reduced serum and hepatic urate levels in hyperuricemic mice [552]. In normal mice, urate levels in liver, but not in serum, were altered with a dose-dependent decrease after *C. cassia* oil treatment.
- Immunomodulatory effects: In vitro, an extract of *C. cassia* markedly stimulated human lymphocytes to proliferate [444]. Cinnamaldehyde derivatives inhibited the lymphoproliferation and induced a T-cell differentiation through the

blockade of early steps in signaling pathways leading to cell growth [244]. *C. cassia* has shown anticomplement action and inhibited the complement-dependent allergic reaction [342]. In rat nephrotoxic serum (NTS) nephritis, *C. cassia* clearly inhibited the excretion of protein into the urine and the increase in peripheral leukocyte counts [343]. Cinnamomi cortex as a component of the unkei-to combination product was found to stimulate the secretion of cytokines (interleukin [IL]-1, IL-6, and IL-8) and the hypothalamus–pituitary axis in laboratory study [472].

- Research has suggested that cinnamon polyphenol extract may affect immune responses by regulating inflammatory response and GLUT gene expression [57]. In another work, sodium benzoate in combination with a cinnamon metabolite ameliorated disease processes associated with experimental allergic encephalomyelitis, an animal model for multiple sclerosis [40].
- Insecticidal effects: Cinnamon essential oils have been shown to serve as effective larvicides against mosquitoes [80] and various harmful flies [449]. Larvicidal tests demonstrated that the leaf essential oils of cinnamaldehyde type, cinnamaldehyde/cinnamyl acetate type, and cinnamyl alcohol had an excellent inhibitory effect against the fourth-instar larvae of *Aedes aegypti* (yellow fever mosquito) [65, 82]. The alcohol extract of *C. camphora* demonstrated effects on *Aphidius gifuensis* and *Diaeretiella rapae* [554]. Results of the 24-hr mosquito larvicidal assays also showed that the effective constituents in leaf essential oils were cinnamaldehyde, eugenol, anethole, and cinnamyl acetate. Cinnamaldehyde exhibited the strongest mosquito larvicidal activity.
- Cinnamon oils and cinnamaldehyde have both been shown to have insecticidal action against Sitophilus oryzae [270]. In addition, cinnamon essential oils displayed insecticidal action against Solenopsis invicta [81] and repellant action against Resseliella oculiperda [509].
- **Metabolic effects**: Cinnamon (*C. zeylanicum*) extracts stimulated 3T3-L1 preadipocytes [479]. Induction of adipocyte formation by cinnamtannin B1 extract gave similar effects related to insulin activity in adipogenesis [479].
- In clinical study, cinnamon increased lean mass [558]. The exact mechanism of action, however, is not well understood.
- **Nematicidal effects**: Cinnamon essential oils have been observed as having nematicidal effects [247].
- Neurologic effects: Cinnamon and various constituents displayed neuroprotective effects in animal and laboratory study [232, 372, 452, 453]. A water extract from the bark of *C. cassia* significantly protected against glutamate-induced cell death and also inhibited glutamate-induced ⁴⁵Ca²⁺ influx using cultured rat cerebellar granule cells [452]. The authors suggest that *C. cassia* bark may have a protective effect on glutamate-induced neuronal death through the inhibition of Ca²⁺ influx. In a model of cytotoxic brain edema in ischemic injury, cinnamon polyphenol extract reduced oxygen–glucose deprivation-induced cell swelling and caused a decline in inner mitochondrial membrane potential in vitro; researchers suggested that cinnamon polyphenol extract may exert its protective effects through mitochondrial permeability transition inhibition [372]. Keishibukuryo-gan, a herbal combination product that contains cinnamon, displayed neuroprotective effects against NO donor-induced neuronal death in cultured

cerebellar granule cells [453]. In laboratory study, chloroform extracts of *C. cassia* showed a marginal neuronal cell protection from direct betaA(1-42) insult, a major cause of Alzheimer's disease pathology [232].

- In animal study, cinnamaldehyde reduced the amplitude of nerve action potential in sciatic nerves [312]; this effect was almost completely reversible.
- Cinnamaldehyde is a known agonist of TRPA1, a receptor implicated in nociception (as well as other functions), and has been used as such in various experiments [11, 19, 49, 122, 123, 160, 250, 326, 346, 347, 360, 546, 547]. As indicated by research in mice, cinnamon may exert an anxiolytic effect via regulation of the serotonergic and GABAergic systems [542]. Cinnamon extracts have been shown to inhibit tau aggregation and filament formation; a significant portion of the effect is attributed to an A-linked proanthocyanidin trimer molecule [377].
- **Phototoxic effects**: Alpha-amyl cinnamic aldehyde, cinnamic alcohol, cinnamic aldehyde, and alpha-amyl cinnamic aldehyde have all been identified as increasing phototoxicity [381].
- **Pigmentation effects**: Cinnamic acid has been shown to reduce melanin production, likely via inhibition of tyrosinase [249, 313].
- Psychiatric effects: Cinnamon has been found to contain allylbenzenes and their isomers, the propenylbenzenes, which have been speculated to be potential metabolic precursors of amphetamines, which may be responsible, in part, for potential mood-enhancing effects [198]. Humans may be exposed to these precursors during baking and cooking; however, the authors note that the biotransformation, pharmacodynamics, and pharmacokinetics of these aromatic allylbenzene compounds are not well understood in human clinical or laboratory studies.
- **Reproductive effects**: Cinnamomi cortex as a component of the unkei-to combination product stimulated the ovulatory process [472].
- Wound-healing effects: In animal study, *C. zeylanicum* bark extract enhanced the wound breaking strength in the case of incision wound, the rate of wound contraction, and the period of epithelization in the case of excision wound. The granulation tissue weight, its breaking strength, and its hydroxyproline content (a nonessential amino acid) were also increased by the extract in the dead space wound [219].

Pharmacodynamics/Kinetics

- **Absorption**: A pharmacokinetic study was performed for measuring the absorption of orally administered procyanidin B-2 and procyanidin B-3 isolated from Cinnamomi cortex (the bark of *C. cassia*) in rat plasma [482]. Intestinal absorption of cinnamaldehyde in anesthetized dogs administered intradermally occurred very early and was long lasting [170].
- A broad range (in terms of molecular mass) of protein–cinnamaldehyde adducts was detected (as formed in a time- and concentration-dependent manner) in skin treated with cinnamaldehyde and cinnamic alcohol but not with alpha-amyl cinnamaldehyde [124].
- The pharmacokinetics of cinnamic acid was compared following oral administration of a decoction of ramulus Cinnamomi (7.4 g/kg; containing cinnamyl alcohol 7.62 × 10⁻⁵ mol/kg and cinnamaldehyde 1.77 × 10⁻⁵ mol/kg) and pure

- cinnamic acid $(7.62 \times 10^{-5} \text{ mol/kg})$ in rats [78]. Results showed that the areas under the plasma concentration AUC(0-t) and AUC(0-infinity) of cinnamic acid were higher in the decoction group than those in the pure group, and that the bioavailability of cinnamic acid from the decoction was higher than that from pure cinnamic acid.
- **Bioavailability**: A study of the pharmacokinetics and relative bioavailability of a combination of radix *Angelicae sinensis* and cortex Cinnamomi revealed that the combination significantly improved the relative bioavailability of ferulic acid to 226.75% [537].
- Excretion: In animal study, constituents of cinnamon, cinnamyl alcohol, and cinnamic aldehyde (the compound which gives cinnamon its odor and flavor) were found to be excreted in the urine [105]. Rats given cinnamic aldehyde excreted two mercapturic acids in the urine, N-acetyl-S-(1-phenyl-3-hydroxypropyl)cysteine (major) and N-acetyl-S-(1-phenyl-2-carboxy ethyl)cysteine (minor) [105]. Cinnamic aldehyde may be an intermediate in mercapturic acid formation of cinnamyl alcohol [104, 105].
- **Metabolism**: The metabolism of OMCA (intragastrically) was studied in rats [421]. The major metabolic pathway (approx. two thirds of the dose) was oxidation to the corresponding cinnamic and phenylpropionic acids (C6–C3 acids), which were largely excreted as glycine conjugates. Intermediate amounts (approx. 10% of the dose) of the O-demethylated C6–C3 acids were excreted. Urinary excretion of metabolites was rapid (91% in 24 hr and 98% in 48 hr) [421].
- Based on in-vitro studies, cinnamon or its constituents may inhibit hepatic microsomal cytochrome P450 [87, 419, 527]. Cinnamon bark was found to inhibit aminopyrine N-demethylation in rat liver microsomes. The component inhibiting drug oxidations catalyzed by CYP1A2 and CYP2E1 was isolated from Cinnamomi cortex and was identified as OMCA [176].
- Minimum inhibitory concentrations (MIC): The MIC of cinnamaldehyde against *C. versicolor* and *L. sulphureus* was 50 and 75 ppm, respectively [515]. OMCA displayed a strong inhibitory effect on the growth of dermatophytoses species, including *Microsporum canis* (MIC 3.12–6.25 mcg/ml); no effect was observed at concentrations as high as 50 mcg/ml [335].

HISTORY

• Cinnamon has been mentioned in historical documents as a well-known spice in the New World and Europe [152]. According to Herodotus (5th century BC): "Arabia is the only country which produces frankincense, myrrh, cassia, and cinnamon," [492]. According to secondary sources, Zakariya al-Qazwini first made mention around the year 1270 that cinnamon grew in Sri Lanka in "Athar al-bilad wa-akhbar al-'ibad" ("Monument of Places and History of God's Bondsmen"). Indonesian traders would sail loads of cinnamon from a group of islands in eastern Indonesia, called the Moluccas, to East Africa. There, traders would transport the cinnamon to markets in Rome. Arab traders would also import the spice overland to Egypt, where Venetian traders would then procure the spice for trade in Europe. Upon the rise of the Ottoman Empire, European traders were obliged to seek out alternate trade routes.

- According to secondary sources, in the early 1500s, the Portuguese established their own monopoly on the cinnamon trade by establishing a stronghold in Sri Lanka. The Salagamas caste of Sri Lankans was traditionally associated with the highly skilled occupation of peeling cinnamon. The Portuguese used the Salagamas in the cinnamon trade. The Salagamas were also to provide cinnamon as a tax. The Dutch managed to dislodge the Portuguese monopoly by forming an alliance with the Kingdom of Maha Nuvara (Senkadagalapura) (i.e., Kandy) in central Sri Lanka. By the mid-1600s, the Portuguese were ousted, and the Dutch East India Company took control of the cinnamon trade. In the late 1700s, however, the British took control of the island and its spice trade. Sri Lanka gained independence from Britain in the late 1940s.
- Beyond Sri Lanka, cinnamon is commonly cultivated in tropical and subtropical regions such as India, Java, Sumatra, the West Indies, Brazil, Vietnam, and Madagascar. Cinnamon is a major product of the Seychelles, an archipelago located east of mainland Africa [2]. Secondary sources cite that in 2006, 90% of the world's cinnamon was produced in Sri Lanka, followed by China, India, and Vietnam.

Explanation of Columns Heads in Table 3

1	2	3	4	5	6	7	8	9	10
Condition	Study design	First author, year	N	Statistically significant?	Quality of study 0-2 = poor 3-4 = good 5 = excellent	Magnitude of benefit	Absolute risk re- duction	Number needed to treat	Comments

Condition

• Refers to the medical condition or disease targeted by a therapy.

Study Design

Common types include:

- Randomized controlled trial (RCT): An experimental trial in which participants are assigned randomly to receive either an intervention being tested or placebo. Note that Natural Standard defines RCTs as being placebo-controlled, while studies using active controls are classified as equivalence trials (see below). In RCTs, participants and researchers are often blinded (i.e., unaware of group assignments), although unblinded and quasi-blinded RCTs are also often performed. True random allocation to trial arms, proper blinding, and sufficient sample size are the basis for an adequate RCT.
- Equivalence trial: An RCT which compares two active agents. Equivalence trials often compare new treatments to usual (standard) care, and may not include a placebo arm.
- Before and after comparison: A study that reports only the change in outcome in each group of a study and does not report between-group comparisons. This is a common error in studies that claim to be RCTs.

TABLE 3. Evidence table

Condition treated (primary or secondary outcome)	Evidence/study type	First author, year	N (participants)	Statistically significant results?	Quality of study: 0-2 = poor 3-4 = good 5 = excellent	Magnitude of benefit (how strong is the effect?)	Absolute risk reduction	Number of patients needed to treat for one outcome	Comments
Antioxidant	RCT	Roussel, 2009	22	Yes	Ø	Medium	Ą	₹ Z	Study was conducted in overweight or obese individuals with impaired fasting glucose. No apparent between-crouns analysis
Bacterial	Systematic	Dugoua,	1 trial	ΑΝ	N A	Small	NA	¥ A	One study was a case report (one patient) Chronic salmonellosis
Candidiasis	Systematic	Dugoua, 2007	1 trial	ΑΝ	N A	Small	NA	A A	One study was a pilot study.
Candidiasis	Pilot study	Quale, 1996	2	S S	N A	₹ Z	NA	Ą	Pilot study, unblinded, small sample size.
Candidiasis	RCT	Cao, 1993	61	Yes	-	Large	Y Y	Y V	Statistically significant ($\rho < .05$), but study design and methods poorly described.
Diabetes	Meta-analysis	Baker, 2008	5 trials; 282 partici- pants	^o Z	A A	∀ Z	Y Y	Y V	Patients with type 1 or type 2 diabetes.
Diabetes	Systematic review	Dugoua, 2007	3 trials	AN	N A	Mixed	Ϋ́	Ą	Patients with type 2 diabetes.
Diabetes	Systematic review	Kleefstra, 2007	5 human trials	Ϋ́ V	A A	∀ Z	Y Y	∀ Z	Authors conclude that cinnamon is not effective for improvements in diversing control
Diabetes	Systematic review	Pham, 2007	3 trials	Y Z	Y V	A A	N	A V	Particular of the properties o

One trial not randomized, one trial investigated adolescents with type 1 diabetes. Other three were RCTs. FBG level reduction in two of three trials.	Women with polycystic ovary syndrome.	Adolescents with type 1 diabetes.	Patients with type 2 diabetes.	Patients with diabetes type 2.	Reduced fasting plasma glucose concentration.	Small sample size, limited	collective, inadequate description of blinding, 1500 mg of cinnamon dailv.	Patients with type 2 diabetes single blind.	Unblinded, no information on standardization of dosing. 1, 3. or 6 o of cinnamon dally.	Unblinded, 1 g daily cinnamon capsules for 90 days.
₹ Z	Ϋ́	₹ Z	ΑN	Ν		Υ		Υ V	Z Z	∀ Z
٧ ٧	N A	N A	Ą	ΑN		Ą		N A	Š Š	Υ V
¥ Z	Medium	N A	Ą	Small		ΑΝ		Y Y	Small	Small
∀ Z	2	4	က	ო		2		-	-	ო
Y Z	Yes	<u>8</u>	N _o	Yes		N _O		N _O	Yes	Yes
5 trials	15	72	77	79		25		09	09	109
Nahas, 2009	Wang, 2007	Altschuler, 2007	Blevins, 2007	Mang, 2006		Vanschoonbeek,	2006	Suppapitiporn, 2006	Khan, 2003	Crawford, 2009
Systematic review	RCT	RCT	RCT	RCT		RCT		RCT	RCT	RCT
Diabetes	Diabetes (insulin potentiation)	Diabetes	Diabetes	Diabetes		Diabetes		Diabetes	Diabetes	Diabetes

(Continued on next page)

TABLE 3. Evidence table (Continued)

Condition treated (primary or secondary outcome)	Evidence/study type	First author, year	N (participants)	Statistically significant results?	Quality of study: 0-2 = poor 3-4 = good 5 = excellent	Magnitude of benefit (how strong is the effect?)	Absolute risk reduction	Number of patients needed to treat for one outcome	Comments
Diabetes	RCT	Solomon, 2007		Yes	-	Small	¥.	A N	5 g vegi-capsulated micronized cinnamon spice (obtained from the C. cassia plant; Everythingcinnamon.com, Essex, UK). No blinding, randomization method unclear.
H. pylori infection Systematic review	Systematic review	Dugoua, 2007	1 trial	Y Y	Y V	None	Υ Y	Y Y	Included one unblinded pilot study (Nir et al., 2000).
H. pylori infection Systematic review	Systematic review	Martin, 2003 1 tria	1 trial	Ϋ́	Ą Z	۷ Z	N A	N A	Included controlled trial (Nir et al., 2000).
H. pylori infection Controlled trial	Controlled trial	Nir, 2000	23	No	Y V	Ϋ́	Υ	N A	Pilot study, unblinded; 40 mg
Insect repellant	Controlled clinical trial	Chang, 2006 4	4	Mixed	₹ Z	Mixed	N A	Y Y	Repellency of trans-cinnamaldehyde, cinnamyl alcohol, and cinnamon oil cream against femal Aedes aegypti
Metabolic syndrome (coronary heart disease)	RCT	Ziegenfuss, 2006	22	Yes	ဇ	Small	V	V	tested using unietent bloassays. Cinnulin PF $^{\oplus}$, 250 mg twice daily.

- Case series: A description of a group of patients with a condition, treatment, or outcome (e.g., 20 patients with migraine headache underwent acupuncture and 17 reported feeling better afterwards). Case series are considered weak evidence of efficacy.
- Case—control study: A study in which patients with a certain outcome are selected and compared with similar patients (without the outcome) to see whether certain risk factors/predictors are more common in patients with that outcome. This study design is not common in the CAM literature.
- Cohort study: A study which assembles a group of patients with certain baseline characteristics (e.g., use of a drug) and follows them forward in time for outcomes. This study design is not common in the CAM literature.
- Meta-analysis: A pooling of multiple trials to increase statistical power (often used to pool data from a number of RCTs with small sample sizes, none which demonstrates significance alone, but in aggregate, can achieve significance). Multiple difficulties are encountered when designing/reviewing these analyses; in particular, outcomes measures or therapies may differ from study to study, hindering direct comparison.
- Review: An author's description of his or her opinion based on personal, nonsystematic review of the evidence.
- Systematic review: A review conducted according to prespecified criteria in an attempt to limit bias from the investigators. Systematic reviews often include a meta-analysis of data from the included studies.

First Author, Year

• Identifies the study being described in a row of the table.

N

• The total number of subjects included in a study (treatment group plus placebo group). Some studies recruit a larger number of subjects initially, but do not use them all because they do not meet the study's entry criteria. In this case, it is the second, smaller number that qualifies as N. N includes all subjects that are part of a study at the start date, even if they drop out, are lost to follow-up, or are deemed unsuitable for analysis by the authors. Trials with a large number of dropouts who are not included in the analysis are considered to be weaker evidence of efficacy. For systematic reviews, the number of studies included is reported. For meta-analyses, the number of total subjects included in the analysis or the number of studies may be reported.

Statistically Significant?

• Results are noted as being statistically significant if a study's authors report statistical significance or if quantitative evidence of significance is present (such as *p* values). P = pending verification.

Quality of Study

• A numerical score between 0 and 5 is assigned as a rough measure of study design/reporting quality (0 being weakest and 5 being strongest). This number is

TABLE 4. Jadad score calculation.

Item	Score
Was the study described as randomized (this includes words such as randomly, random, and randomization)?	0/1
Was the method used to generate the sequence of randomization described and appropriate (table of random numbers, computer-generated, etc)?	0/1
Was the study described as double-blind?	0/1
Was the method of double-blinding described and appropriate (identical placebo, active placebo, dummy, etc.)?	0/1
Was there a description of withdrawals and dropouts?	0/1
Deduct one point if the method used to generate the sequence of randomization was described and it was inappropriate (patients were allocated alternately, or according to date of birth, hospital number, etc.).	0/-1
Deduct one point if the study was described as double-blind but the method of blinding was inappropriate (e.g., comparison of tablet vs. injection with no double-dummy).	0/-1

based on a well-established, validated scale developed by Jadad et al. [206]. This calculation does not account for all study elements that may be used to assess quality (other aspects of study design/reporting are addressed in the "Evidence Discussion" section).

• A Jadad score is calculated using the seven items presented in Table 4. The first five items are indications of good quality, and each counts as one point toward an overall quality score. The final two items indicate poor quality, and a point is subtracted for each if its criteria are met. The range of possible scores is 0 to 5.

Magnitude of Benefit

- This summarizes how strong a benefit is: small, medium, large, or none. If results are not statistically significant, "NA" for "not applicable" is entered. In order to be consistent in defining small, medium, and large benefits across different studies and reviews, Natural Standard defines the magnitude of benefit in terms of the standard deviation (SD) of the outcome measure. Specifically, the benefit is considered:
 - large if >1 SD,
 - medium if 0.5–0.9 SD,
 - small if 0.2–0.4 SD
- In many cases, studies do not report the standard deviation of change of the outcome measure. However, the change in the SD of the outcome measure (also known as effect size) can be calculated, and is derived by subtracting the mean (or mean difference) in the placebo/control group from the mean (or mean difference) in the treatment group, and dividing that quantity by the pooled SD (Effect size = [Mean treatment Mean placebo]/SD_p).

Absolute Risk Reduction

• This describes the difference between the percentage of people in the control/placebo group experiencing a specific outcome (control event rate) and the percentage of people in the experimental/therapy group experiencing that same outcome (experimental event rate). Mathematically, absolute risk

reduction (ARR) equals experimental event rate minus control event rate. ARR is better able to discriminate between large and small treatment effects than relative risk reduction (RRR), a calculation that is often cited in studies ([Control event rate – Experimental event rate]/Control event rate). Many studies do not include adequate data to calculate the ARR, in which cases "NA" is entered into this column. P = pending verification.

Number Needed to Treat

• This is the number of patients who would need to use the therapy under investigation, for the period of time described in the study, in order for one person to experience the specified benefit. It is calculated by dividing the AAR into 1 (1/ARR). P = pending verification.

Comments

• When appropriate, this brief section may comment on design flaws (inadequately described subjects, lack of blinding, brief follow-up, no intention to treat, etc.), notable study design elements (crossover, etc.), dosing, and/or specifics of study group/subgroups (age, gender, etc.). More detailed description of studies is found in the "Evidence Discussion" section.

EVIDENCE DISCUSSION

Allergic Rhinitis

- **Summary**: Cinnamon has demonstrated antiallergic properties in in-vitro study [342]. Based on one randomized, double-blind, placebo-controlled trial, a combination product including *C. zeylanicum*, *Malpighia glabra*, and *Bidens pilosa* has demonstrated reduced allergic nasal symptoms in patients with allergic rhinitis [93]. More well-designed trials are needed before a firm conclusion can be made.
- Evidence (combination study not included in Table 3): Corren et al. conducted a randomized controlled crossover trial to examine the effects of a combination product including C. zeylanicum, Malpighia glabra, and Bidens pilosa for the reduction in nasal symptoms in patients with seasonal allergic rhinitis [93]. Twenty subjects were randomized to receive the combination botanical product (CBP) including C. zeylanicum, Malpighia glabra, and Bidens pilosa (two tablets three times per day), 10 mg of loratadine once a day in the morning, or a placebo. The outcome was a nasal symptom score (NSS). The researchers assessed nasal lavage fluid for tryptase, prostaglandin D2, leukotriene E4 concentrations, and inflammatory cells. The CBP reduced NSS compared with placebo (p = .034). CBP prevented the increase in prostaglandin D2 release; the placebo and loratadine did not have this effect. Tryptase or leukotriene E4 release or inflammatory cell infiltration was not affected by the treatments. No adverse or toxic effects were reported. One of the major limitations of the study is that it employs a combination product. It is therefore difficult to deduce the effects of cinnamon on allergic nasal symptoms alone. It is also unclear as to who sponsored the study.

Angina

- **Summary**: The use of cinnamon for bacterial angina has been reviewed [512]. However, well-designed trials are needed before a firm conclusion can be made.
- Meta-analysis (meta-analysis of combination studies, not included in Table 3): Wang et al. conducted a meta-analysis to evaluate the safety and efficacy of compound salvia pellet (CSP; consisting of Salvia miltiorrhiza, Panax notoginseng, and C. camphora) compared with nitrates for the treatment of chronic stable angina [512]. A search was performed of MEDLINE, EMBASE, BA, Chinese Biomedical Database (CBM), and Chinese Cochrane Centre Controlled Trials Register (1994–2004). Regardless of language and publication status, 27 RCTs (n = 3,722) were isolated. The quality of the methodology of the trials, assessed by the Jadad scale, was determined to be suboptimal. One trial, however, reached a score of three points (quality = good). Pooled results indicated that, compared with nitrates, CSP treatment improved angina symptoms (RR = 1.13, 95% confidence interval [CI]: 1.07-1.20) and improved electrocardiogram (ECG) results (RR = 1.39, 95% CI: 1.28–1.50). Compared with nitrates, patients treated with CSP evinced a decreased percentage of adverse events (2.4% vs. 29.7%). The authors conclude that CSP has a significant, positive effect on angina symptoms and ECG results and has few adverse events. A limitation of the meta-analysis is that it analyzes results from trials studying a combination product; thus, the positive effect of cinnamon alone in the treatment of angina is unclear. The quality of the methodology of the various studies was suboptimal. The authors assert that outcome measures should be expanded to include other endpoints, including mortality and quality of life.

Antioxidant

- Summary: Cinnamon has been suggested as an antioxidant in various studies and reviews [106, 115, 175, 187, 236, 272, 275, 286, 308, 318, 366, 455, 491] due to the high antioxidant content of cinnamon bark [115]. Based on one randomized, double-blind, placebo-controlled trial, a dried aqueous extract of cinnamon (Cinnulin PF®) has been shown to improve the antioxidant status of overweight or obese individuals with impaired fasting glucose [411]. More well-designed trials are needed before a firm conclusion can be made.
- Evidence: Roussel et al. conducted an RCT to examine the effects of a dried aqueous extract of cinnamon (Cinnulin PF®) on the antioxidant status of overweight or obese individuals with impaired fasting glucose [411]. Twenty-two subjects with body mass index (BMI) ranging from 25 to 45 with impaired fasting blood glucose (FBG) received capsules containing 250 mg of an aqueous extract of cinnamon (Cinnulin PF®) or placebo two times per day for 12 weeks. Primary outcome measures included plasma malondialdehyde (MDA) concentrations, plasma antioxidant status (ferric reducing antioxidant power [FRAP] assay), erythrocyte Cu–Zn superoxide (Cu–Zn SOD) activity, and erythrocyte glutathione peroxidase (GPx) activity. Plasma antioxidant status and plasma SH increased, while plasma MDA levels decreased in subjects receiving the cinnamon extract. A positive correlation was also observed between MDA and plasma glucose (r = 0.74, p = .014). A lack of discussion regarding randomization or

blinding methods limits the usefulness of this study. Furthermore, there is a lack of apparent between-groups analyses and a lack of detailed information about study withdrawals. Integrity Nutraceuticals International (Spring Hill, TN, USA) also partially funded the study and supplied the placebo and cinnamon extract capsules used in the study.

Bacterial Infection

- Summary: Preliminary study suggests that cinnamon may treat bacterial infections, including chronic salmonellosis. The use of cinnamon for bacterial enteric infections has been reviewed [410]. However, well-designed trials are needed before a firm conclusion can be made.
- Systematic reviews: Dugoua et al. conducted a systematic review to examine the usefulness of common (*C. verum*, *C. zeylanicum*) and cassia (*C. aromaticum*) cinnamon in various medical conditions [121]. The authors searched nine electronic databases. One pharmacological study on antioxidant activity and seven clinical studies, including chronic salmonellosis (one case study), were found [409]. The authors conclude that common cinnamon showed weak-to-very weak evidence of efficacy in treating chronic salmonellosis.
- Studies of lesser design quality (not included in the Table 3): Rosti et al. report on an exclusively breastfed infant (4 months of age) with acute diarrhea [409]. After consuming raw seafood, the mother developed emesis and diarrhea. *S. enteritidis* was identified in the stool of both the mother and the infant; symptoms resolved after a few days. Stool samples from the mother tested negative for *Salmonella* spp. The infant became a chronic carrier of *S. enteritidis*. Upon the recommendation of a natural remedy expert, the mother fed the infant ground cinnamon bark in homogenized fruit 3–4 times a day. One month later, stool samples of the infant tested negative for *S. enteritidis*; repeat tests 2 and 3 months later were also negative.

Candidiasis

- **Summary**: Cinnamon has been found to have activity against fluconazole-resistant and -susceptible *Candida* isolates in in-vitro studies [393]. Preliminary human studies have yielded mixed results [55, 393]. Further clinical trials may be necessary to determine the usefulness of cinnamon for the treatment of mucosal candidiasis.
- **Systematic reviews**: Dugoua et al. conducted a systematic review to examine the usefulness of common (*C. verum*, *C. zeylanicum*) and cassia (*C. aromaticum*) cinnamon in various medical conditions [121]. The authors searched nine electronic databases. One pharmacological study on antioxidant activity and seven clinical studies, including one study on oral candidiasis in HIV, were found [393]. The authors conclude that common cinnamon showed weak-to-very weak evidence of efficacy in treating oral candidiasis in HIV patients.
- Evidence: Quale et al. conducted a pilot study in five patients with HIV infection and oral candidiasis to investigate the activity of cinnamon (*C. zeylanicum*) against fluconazole-resistant and -susceptible *Candida* isolates [393]. All included patients had pseudomembranous candida infection confirmed by culture.

Patients were given eight lozenges of cinnamon candy no. 1 daily. The commercially available extract was administered for 1 week. No adverse effects were reported. No toxic effects were reported. There were no dropouts. No interactions were reported. Improvement in oral candidiasis served as the outcome measure. Three of the five patients showed improvement in their oral candidiasis (no further details given). Limitations of this study include that it was neither randomized nor blinded and the sample size was very small.

• Cao et al. conducted an RCT to examine the effectiveness of a cortex cinnamon solution in the prevention and treatment of oral candidiasis in hospitalized patients who were likely to receive prolonged and aggressive treatment with antibiotics due to single- or multiple-organ failure [55]. Subjects were randomized to two groups: treatment (N = 34) or placebo (N = 27). The results (no outbreaks vs. 21 outbreaks in placebo group) were statistically significant (p < .05). In-vitro laboratory tests suggested that the cinnamon solution was efficacious in suppressing cell growth. In the discussion, however, the authors indicated that cinnamon solution was used to treat oral candidiasis infection in subjects in the placebo group. It was unclear with regard to whether the use of cinnamon solution was implemented during or after the study, which would have important implications in the validity of the study and interpretation of the results. Although the results were statistically significant, the study design and statistical methods were poorly and insufficiently described and information pertaining to the duration of the study was lacking.

Diabetes

- Summary: Based on human and animal study, cinnamon has been used to control blood sugar [6, 13, 33, 69, 121, 229, 309, 378, 474, 510]. However, results have not been consistent, and various human trial have found that cinnamon did not significantly alter A1C, FBG, or lipid parameters in patients with type 1 or type 2 diabetes [22, 242]. More research on the proposed health benefits of cinnamon supplementation is warranted.
- **Preclinical studies**: The insulin-sensitizing effect of cinnamon was established in in-vitro cell line studies with adipocytes as well as in in-vivo animal studies [29, 41, 200, 208, 228, 252, 271, 367]. The first published in-vivo study on cinnamon supplementation in humans reported a substantial reduction in fasting serum glucose concentration and improvement in blood lipid profile in patients suffering from type 2 diabetes [229].
- **Meta-analysis**: Baker et al. conducted a meta-analysis of RCTs of cinnamon to better characterize its impact on glucose and plasma lipids [22]. A literature search (through July 2007) was conducted; studies examining the effects of on A1C, FBG, or lipid parameters were included. Five prospective RCTs (*N* = 282) were identified [13, 33, 229, 309, 510]. Upon meta-analysis, the authors conclude that the use of cinnamon did not significantly alter A1C, FBG, or lipid parameters in patients with type 1 or type 2 diabetes.
- Systematic reviews: Dugoua et al. conducted a systematic review to examine the usefulness of common (*C. verum*, *C. zeylanicum*) and cassia cinnamon (*C. aromaticum*) in various medical conditions [121]. The authors searched

nine electronic databases. One pharmacological study on antioxidant activity and seven clinical studies, including three randomized clinical trials on type 2 diabetes, were found [229, 309, 510]. The authors conclude that two of the three randomized clinical trials on type 2 diabetes provided strong scientific evidence that cassia cinnamon demonstrates a therapeutic effect in reducing FBG by 10.3%–29%; the third clinical trial did not observe this effect. Cassia cinnamon, however, did not have any effect in lowering glycosylated hemoglobin (HbA1c).

- Kleefstra et al. conducted a systematic review to identify published studies evaluating the effects of cinnamon on glycemic control [242]. The authors searched the MEDLINE database using the search terms (alone and in combination): cinnamon, diabetes mellitus, HbA1c, and glucose. Several animal studies and five randomized placebo-controlled trials in humans were found. Beneficial effects of cinnamon on fasting plasma glucose were observed in patients with type 2 diabetes in one placebo-controlled trial. No effects were observed on HbA1c in any of the studies. Cinnamon was not found to have any effect in patients with type 1 diabetes. The authors conclude that based on available evidence, cinnamon does not appear to have any beneficial effects on glycemic control.
- Pham et al. conducted a systematic review to identify published studies evaluating the effectiveness of cinnamon in patients with type 2 diabetes [378]. The authors conducted a literature search, limited to English-language human studies, using MEDLINE (1966–August 2006), EMBASE (1980–August 2006), International Pharmaceutical Abstracts (1970–August 2006), and the Iowa Drug Information Service (1966–August 2006). References from articles and clinical trials were reviewed for additional sources; no abstracts were reviewed. Two prospective, randomized, double-blind, placebo-controlled, peer-reviewed clinical trials and one prospective, placebo-controlled, peer-reviewed clinical trial that evaluated the efficacy of cinnamon supplementation in patients with type 2 diabetes were identified; a total of 164 patients were involved in these trials [229, 309, 510]. In two of the studies, cinnamon was reported to lower blood glucose levels in small patient samples, while one trial showed no significant difference. The authors conclude that cinnamon may have a modest effect in lowering plasma glucose levels in patients with poorly controlled type 2 diabetes.
- Nahas et al. conducted a review of the clinical evidence from human clinical trials that supports the use of integrative medicine interventions for the improvement in glycemic control in individuals with type 2 diabetes mellitus [344]. MEDLINE (January 1966–August 2008) and EMBASE (January 1966–August 2008) were searched using the search term "type 2 diabetes" in combination with each of the following terms for specific therapies: cinnamon, fenugreek, gymnema, green tea, fibre, momordica, chromium, and vanadium. Cinnamon improved FBG. Its effects on HbA(1c), however, were unknown. The authors conclude that further research on cinnamon is warranted.
- Evidence: Wang et al. conducted a randomized, double-blind, placebo-controlled trial to examine the ability of cinnamon extract to reduce insulin resistance in women with polycystic ovary syndrome (PCOS) [514]. Fifteen women with PCOS were randomized to daily oral cinnamon (one capsule containing 333 mg of cinnamon extract, three times daily) or placebo for 8 weeks. Patients with diabetes, hyperprolactinemia, thyroid disorders, and hypertension were

- excluded. Comparisons of post-treatment to baseline insulin sensitivity indices using fasting and 2-hr oral glucose tolerance tests showed significant reductions in insulin resistance in the cinnamon group but not in the placebo group. This was a well-designed study complete with descriptions of randomization, blinding methods, and study withdrawals.
- Altschuler et al. conducted a randomized, double-blind, placebo-controlled trial to determine the effect of cinnamon on glycemic control in adolescents with type 1 diabetes [13]. Seventy-two adolescent type 1 diabetic subjects were treated in an outpatient setting with cinnamon (1 g daily) or an equivalent-appearing placebo for 90 days. No statistically significant differences in final A1C (8.8 vs. 8.7, p = .88), change in A1C (0.3 vs. 0.0, p = .13), total daily insulin intake, or number of hypoglycemic episodes were observed between the cinnamon and the placebo group. The authors conclude that cinnamon is not effective in improving glycemic control in adolescents with type 1 diabetes. This study was well designed, although randomization methods were not described.
- Blevins et al. conducted a randomized, double-blind, placebo-controlled trial to examine the effect of cinnamon on glucose and lipid levels in non-insulindependent type 2 diabetes [33]. Seventy-seven individuals with type 2 diabetes were enrolled; 17 were excluded. Therefore, 60 patients were randomized to receive 500 mg of cinnamon (*C. cassia*) or placebo (wheat flour) two times daily for 3 months. Fasting glucose, cholesterol (total, low-density lipoprotein [LDL], and HDL), triglyceride, and insulin levels were measured at each visit; no significant changes were observed for any measure at any time point (from baseline to 3 months or from baseline to 1 and 2 months). A lack of discussion regarding randomization or blinding methods limits the usefulness of this study.
- Mang et al. conducted a randomized, double-blind, placebo-controlled trial to determine whether an aqueous purified cinnamon extract improves glycosylated HbA1c, fasting plasma glucose, total cholesterol, LDL, HDL, and triglyceride concentrations in patients with type 2 diabetes [309]. Seventy-nine patients with type 2 diabetes, not on insulin therapy but treated with oral antidiabetics or diet, received a cinnamon extract (equivalent to 3 g of cinnamon powder per day) or a placebo capsule three times a day for 4 months. There was a greater reduction in the cinnamon group (10.3%) than in the placebo group (3.4%) when pre- and post-intervention fasting plasma glucose levels were compared. No significant effects were observed for HbA1c or lipid profiles. Observed decreases in plasma glucose correlated with baseline concentrations; therefore, individuals with higher initial plasma glucose levels may have received more benefit from cinnamon supplementation. No adverse effects were observed. The authors conclude that cinnamon extract may have a moderate effect in reducing fasting plasma glucose concentrations in diabetic patients with poor glycemic control. A lack of discussion regarding randomization or blinding methods limits the usefulness of this study.
- Vanschoonbeek et al. conducted a randomized, placebo-controlled trial of 25 postmenopausal patients to investigate the effects of cinnamon supplementation on insulin sensitivity or glucose tolerance and blood lipid profile in patients with type 2 diabetes [510]. Postmenopausal women diagnosed with type 2 diabetes were included. Exclusion criteria were impaired liver or renal function,

cardiovascular disease, and exogenous insulin therapy. All subjects were using either oral blood glucose-lowering agents or diet only. The subjects received either 1,500 mg of cinnamon (C. cassia) or 1,500 mg of a placebo daily. The cinnamon was consumed for 6 weeks. One capsule (500 mg) was to be ingested at each meal. No information is given concerning standardization of the drug. No allergies or adverse effects were reported. No toxic effects were observed. No dropouts were mentioned. No interactions were observed. Outcome measures were whole-body insulin sensitivity or oral glucose tolerance after 2 and 6 weeks of supplementation. In addition, glycosylated HbA1c and blood lipid profiles were determined. During the intervention period, there were no interactions for plasma HbA1c, fasting glucose, insulin concentrations, or fasting blood lipid concentrations (p > .05). Limitations of the study include inadequate description of blinding or randomization and withdrawals, as well as a small sample size and a limited patient collective, which may have allowed for the introduction of bias.

- Suppapitiporn et al. conducted a randomized, single-blind, placebo-controlled trial to investigate the antidiabetic effect of cinnamon cassia powder in type 2 diabetic patients [474]. Sixty type 2 diabetic patients were randomized to receive either 1.5 g daily of cinnamon cassia powder or placebo for 12 weeks in combination with current treatments (metformin or sulfonylureal) for both groups. Measures of efficacy included HbA1c fasting plasma glucose, lipid profile, blood urea nitrogen (BUN), creatinine, liver function test, and adverse effects. After a 12-week treatment period, HbA1c decreased similarly in both groups. However the proportion of patients achieving HbA1c ≤7% was greater in patients receiving cinnamon compared with those receiving placebo, although this was not statistically significant. No significant between-groups differences were observed for lipid profile or fasting plasma glucose, except for serum glutamic oxaloacetic transaminase (SGOT) activity, from 27.1 (8.75) to 22.1 (5) in the cinnamon group and 24.08 (8.5) to 23.63 (8.88) in the placebo group (*p* = .001). Limitations of the study include inadequate description of blinding or randomization and withdrawals.
- Khan et al. conducted a randomized, placebo-controlled trial of 60 patients (30 men, 30 women) to determine whether cinnamon improves blood glucose and triglyceride, total cholesterol, HDL cholesterol, and LDL cholesterol levels in patients with type 2 diabetes [229]. Selection criteria for the study included the following for patients with type 2 diabetes: age >40 years, not on insulin therapy, not taking medicine for other health conditions, and FBG levels between 7.8 and 22.2 mmol/L (140-400 mg/dL). All subjects were taking sulfonylurea drugs, i.e., glibenclamide; medications did not change during the study. The subjects were randomly divided into six groups. Groups 1, 2, and 3 consumed 1, 3, and 6 g of cinnamon daily, respectively, and Groups 4, 5, and 6 were given placebo capsules corresponding to the number of capsules consumed for the three levels of cinnamon consumed by Groups 12, and 3. The cinnamon was consumed for 40 days followed by a 20-day washout period. No information is given concerning standardization of the drug. No allergies or adverse effects were reported. There were also no problems with compliance or those associated with the consumption of ≤ 6 g of cinnamon per day. No toxic effects were observed. There were no dropouts. No interactions were observed. Outcome measures were the reduction in blood glucose, triglyceride, total cholesterol, HDL cholesterol, and LDL cholesterol

- levels. After 40 days, all three levels of cinnamon reduced the mean fasting serum glucose (18%–29%) and triglyceride (23%–30%), LDL cholesterol (7%–27%), and total cholesterol (12%–26%) levels (p < .05 for each). No significant changes were noted in the placebo groups. Changes in HDL cholesterol were not significant. Limitations of the study include inadequate description of standardization of dosing, blinding, or randomization and withdrawals, as well as failure to conduct an intention-to-treat analysis and lack of dietary standardization.
- Crawford conducted an RCT comparing treatment with cinnamon plus usual care to usual care alone for lowering HbA1c in patients with type 2 diabetes [95]. One hundred nine type 2 diabetics (HbA1c >7.0) were randomized from three primary care clinics on an air force base. Participants were randomly allocated to a usual care (with management changes) group or usual care (with management changes) in addition to 1 g daily of cinnamon capsules for 90 days. One adverse event (rash) was reported by one individual, who withdrew from the study. There were 20 dropouts in total. Cinnamon lowered HbA1c by 0.83% (95% CI: 0.46–1.20). Usual care alone lowered HbA1c by 0.37% (95% CI: 0.15–0.59). The between-groups analysis indicated a statistically significant difference (p < .04). The authors concluded that in addition to usual care, taking cinnamon may be useful in lowering serum HbA1c in type 2 diabetics (HbA1c >7.0). A limitation of this study was that it was not blinded.
- Solomon and Blannin conducted a randomized, controlled crossover trial in lean healthy males (N=7) to assess cinnamon's effects on glucose homeostasis [460]. The doses were as follows: 5 g of a placebo (OGTTcontrol), 5 g of cinnamon (OGTTcin), or 5 g of cinnamon taken 12 hr before the oral glucose tolerance test (OGTTcin12hpre). The 5 g vegi-capsulated micronized cinnamon spice was obtained from the *C. cassia* plant (Everythingcinnamon.com, Essex, UK). Outcome measures were total plasma glucose response and insulin sensitivity. Compared with OGTTcontrol, total plasma glucose responses were found to be lower following OGTTcin and OGTTcin12hpre (p < .05) [–13% and –10% for OGTTcin and OGTTcin12hpre, respectively]. The effect, however, did not reach statistical significance when comparing OGTTcin with OGTTcin12hpre (p > .05). Serum insulin concentration for each OGTT demonstrated a statistically significant difference from baseline (p < .01). There was not, however, a statistically significant main effect of the trial (p > .05). Limitations of the study include that randomization was not discussed and blinding was not apparent.
- Studies of lesser design quality (not included in Table 3): Scheidegger et al. conducted a questionnaire-based study of patients (N = 342) with type 1 diabetes mellitus (T1DM) [424]. In addition to insulin therapy, 48 patients (14%; 13.4% adult, 18.5% pediatric; male = 20, female = 28) used CAM. The most frequently used modalities were cinnamon, homeopathy, magnesium, and "special beverages" (mostly teas). The authors conclude that collaboration between healthcare professionals of various disciplines may provide for optimal patient care.

Eve Disorders

• **Summary**: Preliminary data suggest that a combination herbal eye drop preparation (OphthaCare) may be useful in the treatment of various ophthalmic

- disorders, including conjunctivitis, conjunctival xerosis (dry eye), acute dacryocystitis, degenerative conditions (pterygium or pinguecula), and disorders in postoperative cataract patients [31]. However, well-designed trials are needed before a firm conclusion can be made.
- Evidence (combination study not included in Table 3): Biswas et al. conducted an uncontrolled, prospective, multicenter clinical trial in patients (N = 100) with varying ocular disorders [31]. Diagnosed eye disorders included acute conjunctivitis (bacterial, viral, or allergic), postoperative cataract patients, acute dacryocystitis, conjunctival xerosis (dry eye), and degenerative conditions, such as ptervgium or pinguecula. OphthaCare eye drops were used in the affected eye, two drops four times daily for 15 days. The outcome measures included conjunctival reactions on an ordinal (mild, moderate, or severe) scale. The eye drop preparation contains C. camphora, in addition to Carum coptcium, Terminalia bellirica, Emblica officinalis, Curcuma longa, Ocimum sanctum, Rosa damascena, and Meldespumapum. Acute conjunctivitis improved in 28 of 35 cases (87.5%). Dacryocystitis improved in 15 of 20 cases (88.2%). Degenerative conditions improved in 10 of 15 cases (76.5%). Conjunctival xerosis improved in four of seven cases (66.7%). Postoperative cataracts improved in 19 of 23 cases (95%). No sideeffects were noted. The authors conclude that the product may be safely prescribed. Limitations of this trial include that it used a combination product, the study was not controlled or randomized, and reasons for withdrawal were not discussed. Study methods and analyses of data were suboptimal.

H. pylori infection

- Summary: Based on in-vitro studies, which have shown the effectiveness of cinnamon extracts against *H. pylori* [477, 478], a pilot study was conducted in order to test the activity of an alcoholic extract of cinnamon in a group of patients infected with *H. pylori*. The cinnamon extract, at a concentration of 80 mg daily as a single agent, was ineffective in eradicating *H. pylori*. However, the combination of cinnamon with other antimicrobials, or cinnamon extract at a higher concentration, may prove useful. Further studies are warranted.
- **Systematic reviews**: Dugoua et al. conducted a systematic review to examine the usefulness of common (*C. verum*, *C. zeylanicum*) and cassia cinnamon (*C. aromaticum*) in various medical conditions [121]. The authors searched nine electronic databases. One pharmacological study on antioxidant activity and seven clinical studies, including one study on *H. pylori* infection, were found [354]. The authors conclude that there is a lack of evidence to support the use of cinnamon in *H. pylori* infection eradication.
- Many different plant extracts have been tested for in-vitro antibacterial activity. A review by Martin and Ernst critically evaluated controlled clinical trials of herbal medicines with antibacterial activity [314]. Seven clinical trials met the inclusion criteria. One clinical trial conducted by Nir et al. investigated cinnamon [354].
- **Evidence**: Nir et al. conducted a controlled trial of 23 patients (18 women, 5 men) to test the activity of an alcoholic extract of cinnamon for *H. pylori* infection [354]. Patients were eligible for the study if they had a positive *Camphylobacter* urease

test (CUT) for H. pylori. Patients with a bleeding duodenal ulcer or poor general condition were excluded, as well as pregnant women (or women planning pregnancy); patients using nonsteroidal anti-inflammatory drugs (NSAIDS), steroids, bismuth preparations, alcohol, or illicit drugs; or those having used antibiotics in the preceding months. Fifteen patients (11 women, 4 men) received 40 mg of cinnamon extract; eight patients (7 women, 1 man) received placebo. The extract or the placebo was administered twice daily for 4 weeks. The concentration of the major growth inhibitory component (cinnamaldehyde) was 1.68 mg/ml. The cinnamon extract was well tolerated. Five patients reported minor side-effects. No toxic effects were observed. Seven patients were excluded from the final analysis for the following reasons: negligible count on urea breath test despite presence of bacteria (1 patient), noncompliance (3 patients), and antibiotic treatment (3 patients). No interactions were observed. The amount of H. pylori colonization measured by the ¹³C urea breath test served as the outcome measure. The mean urea breath test counts in the study and control groups before and after therapy were 22.1 and 23.9 versus 24.4 and 25.9, respectively. Results were not statistically significant. This pilot study was neither randomized nor double-blind.

Insect Repellant

- Summary: In laboratory studies, cassia oil (*C. cassia*) sprays reduced dust mites (*Dermatophagoides farinae* and *D. pteronyssinus*) [234]. Preliminary human [65] and laboratory [90] studies suggest that cinnamon may be useful as an insect repellant. However, well-designed trials are needed before a firm conclusion can be made.
- Evidence: Chang et al. conducted a controlled trial comparing the efficacy of (E)-cinnamaldehyde, cinnamyl alcohol, and DEET to repel against Aedes aegypti (L.) female mosquitoes [65]. An "indoor test," a patch bioassay, and a skin bioassay were evaluated (N = 4). In the indoor test, a 5% cassia oil cream was compared with MeiMei® cream (containing citronella and geranium oils) and Repellan S[®] spray (containing 19% DEET). Incremental concentrations of the products were used, and efficacy was measured over time (up to 120 min). The cassia oil cream provided 94, 83, and 61% protection against mosquito bites at 30, 50, and 70 min post application, respectively. MeiMei® cream appeared to be slightly more effective than the cassia oil cream. Repellan S® aerosol continued to provide 91% repellency at 120 min post application. In the patch bioassay tests, at 40 min post application and exposure to the mosquitoes, (E)-cinnamaldehyde (0.153 mg cm⁻²) provided 93% protection and DEET (0.051 mg cm⁻²) provided 89% protection. In the skin bioassay tests, at 30 min post application, (E)-cinnamaldehyde (0.051mg cm⁻²) provided 87% protection against mosquito bites and DEET (0.025 mg cm⁻²) provided 95% protection. In both bioassays, (E)-cinnamaldehyde appeared to be more effective than cinnamyl alcohol. The authors conclude that products containing cassia oil merit further study as potential insect repellents. Limitations of the study include that it was not randomized, it is unclear whether between-groups analyses were conducted in separate trials, and the study population was small (N = 4).

Lung Cancer

- **Summary**: Preliminary study suggests that cinnamon may be useful in the treatment of lung cancer [220]. However, well-designed trials are needed before a firm conclusion can be made.
- Evidence (combination case study not included in Table 3): Kamei et al. conducted a case study pertaining to the treatment of a 77-year-old female with lung cancer with a combination Chinese medicine product Ninjin Yoei To (NYT; Ren-Shen-Yang-Rong-Tang; Kotaro Pharmaceutical Co., Ltd, Osaka, Japan) [220]. In total, 15 g per day of NYT was used for 7 weeks. The dried extract contained ginseng, cinnamon bark, Japanese angelica root, astragalus root, peony root, citrus unshiu peel, rehmannia root, polygala root, atractylodes rhizome, schisanda fruit, poria sclerotium, and glycyrrhiza. Outcome measures were the tumor marker levels (carcinoembryonic antigen [CEA] and carbohydrate antigen 19–9 [CA19–9]), yin-yang, and xu-shi scores. CEA and CA19-9 before the treatment were 14.6 ng/ml and 55.0 U/ml, respectively. At the end of the seventh week, the CEA and CA19-9 scores decreased to 11.3 ng/ml and 39.2 U/ml, respectively. Yang and shi scores were based on the following: (a) a return to a natural facial complexion from pale, (b) a return to a reddish tint on the tip of tongue, (c) easily palpable pulse, (d) disappearance of a feeling of cold in limbs, (e) a change in eyes from dull to shining, (f) marked increase in abdominal tension, (g) improvement in skin color, and (h) decreased spontaneous sweating. Before treatment, yin-yang and xu-shi scores were -5 and -12.5, respectively, and +12.5 and +22.5 at the end of the 7 weeks. The patient's cough disappeared, her appetite recovered, and an increase of 6 kg in body weight was observed. Limitations of this study are that it was a case study, thus findings are not generalizable, and that a combination product was used.

Metabolic Syndrome (Coronary Heart Disease)

- **Summary**: Preliminary study suggests that cinnamon may be useful in the treatment of features of metabolic syndrome in prediabetic subjects [558]. However, well-designed trials are needed before a firm conclusion can be made.
- Evidence: Ziegenfuss et al. conducted a randomized, placebo-controlled, double-blind clinical trial with two parallel groups to verify the effects of a water-soluble cinnamon extract (Cinnulin PF®) on FBG, SBP, and body composition (i.e., characteristics of metabolic syndrome) in patients with prediabetes [558]. Subjects (N=22) were randomly assigned to treatment with Cinnulin PF® (two capsules of 250 mg each, twice daily) or a placebo for 12 weeks. Primary outcomes included changes in FBG, SBP, and body composition. Compared with the placebo group, the treatment group noted decreases in FBG (-8.4%: from 116.3 \pm 12.8 mg/dL [pre] to 106.5 ± 20.1 mg/dL [post], p < .01) and SBP (-3.8%: from 133 \pm 14 mmHg [pre] to 128 ± 18 mmHg [post], p < .001), as well as increases in lean mass (+1.1%: from 53.7 ± 11.8 kg [pre] to 54.3 ± 11.8 kg [post], p < .002). The authors conclude that Cinnulin PF® may reduce FBG and SBP and improve body composition in individuals with metabolic syndrome. The authors continue that cinnamon may reduce risk factors correlated with diabetes and cardiovascular disease.

BRANDS USED IN CLINICAL TRIALS/THIRD-PARTY TESTING

• Cinnulin PF® [411, 558].

Declaration of interest: The authors report no conflict of interest. The authors alone are responsible for the content and writing of this monograph.

ABOUT THE AUTHORS

Catherine Ulbricht, PharmD, Massachusetts General Hospital, Boston, MA, USA. Erica Seamon, PharmD, Natural Standard Research Collaboration, Somerville, MA, USA. Regina C. Windsor, MPH, Natural Standard Research Collaboration, Somerville, MA, USA. Nicole Armbruester, PhD, Analyze & Realize, Berlin, Germany. J. Kathryn Bryan, BA, Natural Standard Research Collaboration, Somerville, MA, USA. Dawn Costa, BA, BS, Natural Standard Research Collaboration, Somerville, MA, USA, Nicole Giese, MS, Natural Standard Research Collaboration, Somerville, MA, USA. Joerg Gruenwald, PhD, Phytopharm Research, Berlin, Germany. Ramon Iovin, PhD, Natural Standard Research Collaboration, Somerville, MA, USA. Richard Isaac, BA, Natural Standard Research Collaboration, Somerville, MA, USA. Jill M. Grimes Serrano, PhD, Natural Standard Research Collaboration, Somerville, MA, USA. Shaina Tanguay-Colucci, BS, Natural Standard Research Collaboration, Somerville, MA, USA. Wendy Weissner, BA, Natural Standard Research Collaboration, Somerville, MA, USA. Heeja Yoon, PharmD, Drake University, Des Moines, IA, USA. Jie Zhang, MPH, PhD, University of Alabama at Birmingham, Birmingham, AL, USA.

REFERENCES

- [1] WORKER'S compensation acts: permanence of cinnamon sensitivity. J Am Med Assoc. 1952;149(17):1596.
- [2] Seychelles. Backgr. Notes Ser. 1989;1–7.
- [3] Cinnamon linked to oral sores, irritation. J Am Dent Assoc. 1995;126(9):1214.
- [4] Application of gas-liquid chromatography to the analysis of essential oils. Part XVIII: Determination of safrole in oils of cinnamon leaf, *Litsea cubeba*, and nutmeg. Report by the Analytical Methods Committee. Analyst 2002;127(3):428–429.
- [5] NTP toxicology and carcinogenesis studies of trans-cinnamaldehyde (CAS No. 14371–10-9) in F344/N rats and B6C3F1 mice (feed studies). Natl Toxicol Program Tech Rep Ser. 2004;(514):1–281.
- [6] [Cinnamon to control blood sugar]. MMW Fortschr Med. 2004;146(48):12.
- [7] Is it true that cinnamon can lower blood sugar in people with diabetes? Mayo Clin Health Lett. 2007;25(4):8.
- [8] Abraham SK, Singh SP, Kesavan PC. In vivo antigenotoxic effects of dietary agents and beverages co-administered with urethane: assessment of the role of glutathione Stransferase activity. Mutat Res. 1998;413(2):103–110.
- [9] Ackermann L, Aalto-Korte K, Jolanki R, Alanko K. Occupational allergic contact dermatitis from cinnamon including one case from airborne exposure. Contact Dermatitis 2009;60(2):96–99.
- [10] Akira T, Tanaka S, Tabata M. Pharmacological studies on the antiulcerogenic activity of Chinese cinnamon. Planta Med. 1986;52(6):440–443.
- [11] Albin KC, Carstens MI, Carstens E. Modulation of oral heat and cold pain by irritant chemicals. Chem Senses 2008;33(1):3–15.

- [12] Allen CM, Blozis GG. Oral mucosal reactions to cinnamon-flavored chewing gum. J Am Dent Assoc. 1988;116(6):664–667.
- [13] Altschuler JA, Casella SJ, MacKenzie TA, Curtis KM. The effect of cinnamon on A1C among adolescents with type 1 diabetes. Diabetes Care 2007;30(4):813–816.
- [14] Amalaradjou MA, Hoagland TA, Venkitanarayanan K. Inactivation of *Enterobacter sakazakii* in reconstituted infant formula by trans-cinnamaldehyde. Int J Food Microbiol. 2009;129(2):146–149.
- [15] Ammon HP. [Cinnamon in type 2 diabetics]. Med Monatsschr Pharm. 2008;31(5):179–183.
- [16] Ananda, Baskaran S, Kazmer GW, Hinckley L, Andrew SM, Venkitanarayanan K. Antibacterial effect of plant-derived antimicrobials on major bacterial mastitis pathogens in vitro. J Dairy Sci. 2009;92(4):1423–1429.
- [17] Anderson RA. Chromium and polyphenols from cinnamon improve insulin sensitivity. Proc Nutr Soc. 2008;67(1):48–53.
- [18] Anderson RA, Broadhurst CL, Polansky MM, Schmidt WF, Khan A, Flanagan VP, Schoene NW, Graves DJ. Isolation and characterization of polyphenol type-A polymers from cinnamon with insulin-like biological activity. J Agric Food Chem. 2004;52(1):65–70.
- [19] Andrade EL, Luiz AP, Ferreira J, Calixto JB. Pronociceptive response elicited by TRPA1 receptor activation in mice. Neuroscience 2008;152(2):511–520.
- [20] Arfa AB, Preziosi-Belloy L, Chalier P, Gontard N. Antimicrobial paper based on a soy protein isolate or modified starch coating including carvacrol and cinnamaldehyde. J Agric Food Chem. 2007;55(6):2155–2162.
- [21] Atta AH, Alkofahi A. Anti-nociceptive and anti-inflammatory effects of some Jordanian medicinal plant extracts. J Ethnopharmacol 1998;60(2):117–124.
- [22] Baker WL, Gutierrez-Williams G, White CM, Kluger J, Coleman CI. Effect of cinnamon on glucose control and lipid parameters. Diabetes Care 2008;31(1):41–43.
- [23] Balachandran, B, Sivaramkrishnan VM. Induction of tumours by Indian dietary constituents. Indian J Cancer 1995;32(3):104–109.
- [24] Ballabh B, Chaurasia OP, Ahmed Z, Singh SB. Traditional medicinal plants of cold desert Ladakh-used against kidney and urinary disorders. J Ethnopharmacol. 2008;118(2):331–339.
- [25] Bandell M, Story GM, Hwang SW, Viswanath V, Eid SR, Petrus MJ, Earley TJ, Patapoutian A. Noxious cold ion channel TRPA1 is activated by pungent compounds and bradykinin. Neuron 2004;41(6):849–857.
- [26] Barceloux DG. Cinnamon (Cinnamonum species). Dis Mon. 2009;55(6):327–335.
- [27] Bartine H, Tantaoui-Elaraki A. Growth and toxigenesis of *Aspergillus flavus* isolates on selected spices. J Environ Pathol Toxicol Oncol. 1997;16(1):61–65.
- [28] Becerril R, Gomez-Lus R, Goni P, Lopez P, Nerin C. Combination of analytical and microbiological techniques to study the antimicrobial activity of a new active food packaging containing cinnamon or oregano against *E. coli* and *S. aureus*. Anal Bioanal Chem. 2007;388(5–6):1003–1011.
- [29] Berrio LF, Polansky MM, Anderson RA. Insulin activity: stimulatory effects of cinnamon and brewer's yeast as influenced by albumin. Horm Res. 1992;37(6):225– 229.
- [30] Bhattacharjee S, Rana T, Sengupta A. Inhibition of lipid peroxidation and enhancement of GST activity by cardamom and cinnamon during chemically induced colon carcinogenesis in Swiss albino mice. Asian Pac J Cancer Prev. 2007;8(4):578–582.
- [31] Biswas NR, Gupta SK, Das GK, Kumar N, Mongre PK, Haldar D, Beri S. Evaluation of Ophthacare eye drops a herbal formulation in the management of various ophthalmic disorders. Phytother Res. 2001;15(7):618–620.
- [32] Blank DM, Mattes RD. Sugar and spice: similarities and sensory attributes. Nurs Res. 1990;39(5):290–293.
- [33] Blevins SM, Leyva MJ, Brown J, Wright J, Scofield RH, Aston CE. Effect of cinnamon on glucose and lipid levels in non insulin-dependent type 2 diabetes. Diabetes Care 2007;30(9):2236–2237.

- [34] Blomhoff, R. [Antioxidants and oxidative stress]. Tidsskr Nor Laegeforen. 2004;124(12):1643–1645.
- [35] Blumenthal, Busse WR, Goldberg A, et al. Cinnamon bark. In: Blumenthal, Busse WR, Goldberg A, et al. (Eds.), *The complete German Commission E monographs: therapeutic guide to herbal medicines*, pp. 110–111. Austin, TX: American Botanical Council, 1998.
- [36] Boudry, G, Perrier C. Thyme and cinnamon extracts induce anion secretion in piglet small intestine via cholinergic pathways. J Physiol Pharmacol. 2008;59(3):543–552.
- [37] Bousquet PJ, Guillot B, Guilhou JJ, Raison-Peyron N. A stomatitis due to artificial cinnamon-flavored chewing gum. Arch. Dermatol. 2005;141(11):1466–1467.
- [38] Brackman G, Defoirdt T, Miyamoto C, Bossier P, Van Calenbergh S, Nelis H, Coenye T. Cinnamaldehyde and cinnamaldehyde derivatives reduce virulence in *Vibrio* spp. by decreasing the DNA-binding activity of the quorum sensing response regulator LuxR. BMC Microbiol. 2008;8:149.
- [39] Bradley R, Oberg EB, Calabrese C, Standish LJ. Algorithm for complementary and alternative medicine practice and research in type 2 diabetes. J Altern Complement Med. 2007;13(1):159–175.
- [40] Brahmachari, S, Pahan K. Sodium benzoate, a food additive and a metabolite of cinnamon, modifies T cells at multiple steps and inhibits adoptive transfer of experimental allergic encephalomyelitis. J Immunol. 2007;179(1):275–283.
- [41] Broadhurst CL, Polansky MM, Anderson RA. Insulin-like biological activity of culinary and medicinal plant aqueous extracts in vitro. J Agric Food Chem 2000;48(3):849–852.
- [42] Brophy GC, Mohandas J, Slaytor M, Sternhell S, Watson TR, Wilson LA. Novel lignans from a *Cinnamomum* sp. from Bougainville. Tetrahedron Lett. 1969;59:5159– 5162.
- [43] Browder LW, Williamson JH. The effects of cinnamon on xanthine dehydrogenase, aldehyde oxidase, and pyridoxal oxidase activity during development in *Drosophila melanogaster*. Dev Biol 1976;53(2):241–249.
- [44] Buch JG, Dikshit RK, Mansuri SM. Effect of certain volatile oils on ejaculated human spermatozoa. Indian J Med Res. 1988;87:361–363.
- [45] Buchalter, L. Identification of polymeric 5,7,3',4'-tetrahydroxyflavan-3,4-diol from tannin extract of powdered cassia bark, Saigon, cinnamon USP, *Cinnamomum loureirii* Nees, family Lauraceae. J Pharm Sci. 1971;60(1):144–145.
- [46] Buckley DA, Basketter DA, Smith Pease CK, Rycroft RJ, White IR, McFadden JP. Simultaneous sensitivity to fragrances. Br J Dermatol. 2006;154(5):885–888.
- [47] Burt S. Essential oils: their antibacterial properties and potential applications in foods a review. Int J Food Microbiol. 2004;94(3):223–253.
- [48] Buyukbalci A, El SN. Determination of in vitro antidiabetic effects, antioxidant activities and phenol contents of some herbal teas. Plant Foods Hum Nutr. 2008;63(1):27–33.
- [49] Cahusac PM, Noyce R. A pharmacological study of slowly adapting mechanoreceptors responsive to cold thermal stimulation. Neuroscience 2007;148(2):489–500.
- [50] Calnan CD. Cinnamon dermatitis from an ointment. Contact Dermatitis 1976;2(3):167–170.
- [51] Calsamiglia S, Busquet M, Cardozo PW, Castillejos L, Ferret A. Invited review: essential oils as modifiers of rumen microbial fermentation. J Dairy Sci. 2007;90(6):2580–2595.
- [52] Calucci L, Pinzino C, Zandomeneghi M, Capocchi A, Ghiringhelli S, Saviozzi F, Tozzi S, Galleschi L. Effects of gamma-irradiation on the free radical and antioxidant contents in nine aromatic herbs and spices. J Agric Food Chem 2003;51(4):927–934.
- [53] Camarasa JM. First epidemiological study of contact dermatitis in Spain 1977. Spanish Contact Dermatitis Research Group. Acta Derm Venereol Suppl (Stockh). 1979;59(85):33-37.
- [54] Campbell TM, Neems R, Moore J. Severe exacerbation of rosacea induced by cinnamon supplements. J Drugs Dermatol. 2008;7(6):586–587.
- [55] Cao GY. [Prevention and treatment of oral candidiasis with cortex cinnamon solution]. Zhonghua Hu Li Za Zhi. 1993;28(12):711–712.

- [56] Cao H, Polansky MM, Anderson RA. Cinnamon extract and polyphenols affect the expression of tristetraprolin, insulin receptor, and glucose transporter 4 in mouse 3T3-L1 adipocytes. Arch Biochem Biophys. 2007;459(2):214–222.
- [57] Cao H, Urban JF, Jr., Anderson RA. Cinnamon polyphenol extract affects immune responses by regulating anti- and proinflammatory and glucose transporter gene expression in mouse macrophages. J Nutr. 2008;138(5):833–840.
- [58] Cardozo PW, Calsamiglia S, Ferret A, Kamel, C. Effects of natural plant extracts on ruminal protein degradation and fermentation profiles in continuous culture. J Anim Sci. 2004;82(11):3230–3236.
- [59] Cava R, Nowak E, Taboada A, Marin-Iniesta F. Antimicrobial activity of clove and cinnamon essential oils against *Listeria monocytogenes* in pasteurized milk. J Food Prot. 2007;70(12):2757–2763.
- [60] Cavier, R, Debelmas. [Anthelmintic properties of the essences of Ceylon cinnamon and of clove.]. Therapie 1950;5(3):140–143.
- [61] Cervenka L, Peskova I, Foltynova E, Pejchalova M, Brozkova I, Vytrasova J. Inhibitory effects of some spice and herb extracts against *Arcobacter butzleri*, *A. cryaerophilus*, and *A. skirrowii*. Curr Microbiol. 2006;53(5):435–439.
- [62] Cervenka L, Peskova I, Pejchalova M, Vytrasova J. Inhibition of Arcobacter butzleri, Arcobacter cryaerophilus, and Arcobacter skirrowii by plant oil aromatics. J Food Prot. 2008;71(1):165–169.
- [63] Chaigneau, M, Muraz B. [Decontamination of some spices by ethylene oxide. Development of 2-chloroethanol and ethylene glycol during the preservation]. Ann Pharm Fr. 1993;51(1):47–53.
- [64] Chang CW, Chang WL, Chang ST, Cheng SS. Antibacterial activities of plant essential oils against *Legionella pneumophila*. Water Res. 2008;42(1–2):278–286.
- [65] Chang KS, Tak JH, Kim SI, Lee WJ, Ahn YJ. Repellency of *Cinnamomum cassia* bark compounds and cream containing cassia oil to *Aedes aegypti* (Diptera: Culicidae) under laboratory and indoor conditions. Pest Manag Sci. 2006;62(11):1032–1038.
- [66] Chao LK, Hua KF, Hsu HY, Cheng SS, Lin IF, Chen CJ, Chen ST, Chang ST. Cinnamaldehyde inhibits pro-inflammatory cytokines secretion from monocytes/macrophages through suppression of intracellular signaling. Food Chem Toxicol. 2008;46(1):220–231.
- [67] Chao LK, Hua KF, Hsu HY, Cheng SS, Liu JY, Chang ST. Study on the antiinflammatory activity of essential oil from leaves of *Cinnamomum osmophloeum*. J Agric Food Chem 2005;53(18):7274–7278.
- [68] Charles AS, Baskaran SA, Murcott C, Schreiber D, Hoagland T, Venkitanarayanan K. Reduction of *Escherichia coli* O157:H7 in cattle drinking-water by trans-cinnamaldehyde. Foodborne Pathog Dis. 2008;5(6):763–771.
- [69] Chase CK, McQueen CE. Cinnamon in diabetes mellitus. Am J Health Syst Pharm. 2007;64(10):1033–1035.
- [70] Chen CH, Lo WL, Liu YC, Chen CY. Chemical and cytotoxic constituents from the leaves of *Cinnamomum kotoense*. J Nat Prod. 2006;69(6):927–933.
- [71] Chen CY, Chen CH, Lo YC, Wu BN, Wang HM, Lo WL, Yen CM, Lin RJ. Anticancer activity of isoobtusilactone A from *Cinnamomum kotoense*: involvement of apoptosis, cell-cycle dysregulation, mitochondria regulation, and reactive oxygen species. J Nat Prod. 2008;71(6):933–940.
- [72] Chen CY, Chen CH, Wong CH, Liu YW, Lin YS, Wang YD, Hsui YR. Cytotoxic constituents of the stems of Cinnamomum subavenium. J Nat Prod. 2007;70(1):103–106.
- [73] Chen CY, Hsu YL, Chen YY, Hung JY, Huang MS, Kuo PL. Isokotomolide A, a new butanolide extracted from the leaves of *Cinnamomum kotoense*, arrests cell cycle progression and induces apoptosis through the induction of p53/p21 and the initiation of mitochondrial system in human non-small cell lung cancer A549 cells. Eur J Pharmacol. 2007;574(2–3):94–102.
- [74] Chen CY, Liu TZ, Chen CH, Wu CC, Cheng JT, Yiin SJ, Shih MK, Wu MJ, Chern CL. Isoobtusilactone A-induced apoptosis in human hepatoma Hep G2 cells is

- mediated via increased NADPH oxidase-derived reactive oxygen species (ROS) production and the mitochondria-associated apoptotic mechanisms. Food Chem Toxicol. 2007;45(7):1268–1276.
- [75] Chen FC, Peng CF, Tsai IL, Chen IS. Antitubercular constituents from the stem wood of Cinnamomum kotoense. J Nat Prod. 2005;68(9):1318–1323.
- [76] Chen JF, Chen GX, Li XJ, Quan SM, Zeng YY, Hu YJ, Huang KE, Liu XL. [Effect of overall alkali of Tongbiling on CD69 expression activated mouse T lymphocytes]. Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi. 2003;19(1):93–94.
- [77] Chen, Y. [Pharmacological studies of *Cinnamomum cassia* bark. Part I. Effects on the blood and cardiovascular system]. Zhong Yao Tong Bao. 1981;6(5):32–34.
- [78] Chen Y, Ma Y, Ma W. Pharmacokinetics and bioavailability of cinnamic acid after oral administration of ramulus Cinnamomi in rats. Eur J Drug Metab Pharmacokinet. 2009;34(1):51–56.
- [79] Cheng SS, Liu JY, Hsui YR, Chang ST. Chemical polymorphism and antifungal activity of essential oils from leaves of different provenances of indigenous cinnamon (*Cinnamomum osmophloeum*). Bioresour Technol. 2006;97(2):306–312.
- [80] Cheng SS, Liu JY, Huang CG, Hsui YR, Chen WJ, Chang ST. Insecticidal activities of leaf essential oils from *Cinnamomum osmophloeum* against three mosquito species. Bioresour Technol. 2009;100(1):457–464.
- [81] Cheng SS, Liu JY, Lin CY, Hsui YR, Lu MC, Wu WJ, Chang ST. Terminating red imported fire ants using *Cinnamomum osmophloeum* leaf essential oil. Bioresour Technol. 2008;99(4):889–893.
- [82] Cheng SS, Liu JY, Tsai KH, Chen WJ, Chang ST. Chemical composition and mosquito larvicidal activity of essential oils from leaves of different *Cinnamomum osmophloeum* provenances. J Agric Food Chem 2004;52(14):4395–4400.
- [83] Chericoni S, Prieto JM, Iacopini P, Cioni P, Morelli I. In vitro activity of the essential oil of *Cinnamomum zeylanicum* and eugenol in peroxynitrite-induced oxidative processes. J Agric Food Chem 2005;53(12):4762–4765.
- [84] Chi AC, Ravenel MC. AAOMP case challenge: a "speckled" lesion. J Contemp Dent Pract. 2005;6(1):168–172.
- [85] Chipley JR, Uraih N. Inhibition of Aspergillus growth and aflatoxin release by derivatives of benzoic acid. Appl Environ Microbiol. 1980;40(2):352–357.
- [86] Chohan M, Forster-Wilkins G, Opara EI. Determination of the antioxidant capacity of culinary herbs subjected to various cooking and storage processes using the ABTS(*+) radical cation assay. Plant Foods Hum Nutr. 2008;63(2):47–52.
- [87] Choi J, Lee KT, Ka H, Jung WT, Jung HJ, Park HJ. Constituents of the essential oil of the *Cinnamomum cassia* stem bark and the biological properties. Arch Pharm Res. 2001;24(5):418–423.
- [88] Chua MT, Tung YT, Chang ST. Antioxidant activities of ethanolic extracts from the twigs of *Cinnamomum osmophloeum*. Bioresour Technol. 2008;99(6):1918–1925.
- [89] Clark L, Shivik J. Aerosolized essential oils and individual natural product compounds as brown treesnake repellents. Pest Manag Sci. 2002;58(8):775–783.
- [90] Cloyd RA, Galle CL, Keith SR, Kalscheur NA, Kemp KE. Effect of commercially available plant-derived essential oil products on arthropod pests. J Econ Entomol. 2009;102(4):1567–1579.
- [91] Cohen DM, Bhattacharyya I. Cinnamon-induced oral erythema multiforme-like sensitivity reaction. J Am Dent Assoc 2000;131(7):929–934.
- [92] Conner DE, Beuchat LR. Sensitivity of heat-stressed yeasts to essential oils of plants. Appl Environ Microbiol. 1984;47(2):229–233.
- [93] Corren J, Lemay M, Lin Y, Rozga L, Randolph RK. Clinical and biochemical effects of a combination botanical product (ClearGuard) for allergy: a pilot randomized double-blind placebo-controlled trial. Nutr J. 2008;7:20.
- [94] Cox SD, Markham JL. Susceptibility and intrinsic tolerance of *Pseudomonas aeruginosa* to selected plant volatile compounds. J Appl Microbiol. 2007;103(4):930–936.

- [95] Crawford P. Effectiveness of cinnamon for lowering hemoglobin A1C in patients with type 2 diabetes: a randomized, controlled trial. J Am Board Fam Med 2009;22(5):507–512.
- [96] Cuong NM, Taylor WC, Sung VT. A new cyclobutane lignan from Cinnamomum balansae. Nat Prod Lett. 2001;15(5):331–338.
- [97] Danneman PJ, Booman KA, Dorsky J, Kohrman KA, Rothenstein AS, Sedlak RI, Steltenkamp RJ, Thompson GR. Cinnamic aldehyde: a survey of consumer patch-test sensitization. Food Chem Toxicol. 1983;21(6):721–725.
- [98] Dannemann K, Hecker W, Haberland H, Herbst A, Galler A, Schafer T, Brahler E, Kiess W, Kapellen TM. Use of complementary and alternative medicine in children with type 1 diabetes mellitus prevalence, patterns of use, and costs. Pediatr Diabetes 2008;9(3 Pt 1):228–235.
- [99] De M, De AK, Banerjee AB. Antimicrobial screening of some Indian spices. Phytother Res. 1999;13(7):616–618.
- [100] De Benito V, Alzaga R. Occupational allergic contact dermatitis from cassia (Chinese cinnamon) as a flavouring agent in coffee. Contact Dermatitis 1999;40(3):165.
- [101] De Luis DA, Aller R, Romero E. [Cinnamon as possible treatment of diabetes mellitus type 2]. Med Clin (Barc.) 2008;131(7):279.
- [102] De Silva HV, Shankel DM. Effects of the antimutagen cinnamaldehyde on reversion and survival of selected *Salmonella* tester strains. Mutat Res. 1987;187(1):11–19.
- [103] Decapite TJ, Anderson BE. Allergic contact dermatitis from cinnamic aldehyde found in an industrial odour-masking agent. Contact Dermatitis 2004;51(5-6):312-313.
- [104] Delbressine LP, Klippert PJ, Reuvers JT, Seutter-Berlage F. Identification of two sulphur containing urinary metabolites of cinnamic aldehyde in the rat [proceedings]. Br J Pharmacol. 1980;68(1):165P.
- [105] Delbressine LP, Klippert PJ, Reuvers JT, Seuttler-Berlage F. Isolation and identification of mercapturic acids of cinnamic aldehyde and cinnamyl alcohol from urine of female rats. Arch Toxicol. 1981;49(1):57–64.
- [106] Dhuley JN. Anti-oxidant effects of cinnamon (Cinnamomum verum) bark and greater cardamom (Amomum subulatum) seeds in rats fed high-fat diet. Indian J Exp Biol. 1999;37(3):238–242.
- [107] Di Pasqua R, Betts G, Hoskins N, Edwards M, Ercolini D, Mauriello G. Membrane toxicity of antimicrobial compounds from essential oils. J Agric Food Chem. 2007;55(12):4863–4870.
- [108] Diba VC, Statham BN. Contact urticaria from cinnamal leading to anaphylaxis. Contact Dermatitis 2003;48(2):119.
- [109] Didry N, Dubreuil L, Pinkas M. [Antibacterial activity of thymol, carvacrol and cinnamaldehyde alone or in combination]. Pharmazie 1993;48(4):301–304.
- [110] Domadia P, Swarup S, Bhunia A, Sivaraman J, Dasgupta D. Inhibition of bacterial cell division protein FtsZ by cinnamaldehyde. Biochem Pharmacol. 2007;74(6):831–840.
- [111] Dong L, Gordon VA, Grange RL, Johns J, Parsons PG, Porzelle A, Reddell P, Schill H, Williams CM. Structure and absolute stereochemistry of the anticancer agent EBC-23 from the Australian rainforest. J Am Chem Soc. 2008;130(46):15262–15263.
- [112] Dong L, Schill H, Grange RL, Porzelle A, Johns JP, Parsons PG, Gordon VA, Reddell PW, Williams CM. Anticancer agents from the Australian tropical rainforest: Spiroacetals EBC-23, 24, 25, 72, 73, 75 and 76. Chemistry 2009 Oct 26;15(42):11307–11318.
- [113] Dooms-Goossens A, Dubelloy R, Degreef H. Contact and systemic contact-type dermatitis to spices. Dermatol Clin 1990;8(1):89–93.
- [114] Double KL, Rowe DB, Hayes M, Chan DK, Blackie J, Corbett A, Joffe R, Fung VS, Morris J, Halliday GM. Identifying the pattern of olfactory deficits in Parkinson disease using the brief smell identification test. Arch Neurol. 2003;60(4):545–549.
- [115] Dragland S, Senoo H, Wake K, Holte K, Blomhoff R. Several culinary and medicinal herbs are important sources of dietary antioxidants. J Nutr. 2003;133(5):1286–1290.
- [116] Drake TE, Maibach HI. Allergic contact dermatitis and stomatitis caused by a cinnamic aldehyde-flavored toothpaste. Arch Dermatol 1976;112(2):202–203.

- [117] Du ZQ, Zhou ZM, Xiong YL, Zhao X, Li JH, Wu YH. [Pharmacodynamical research of Jingu Tongxiao granule]. Zhongguo Zhong Yao Za Zhi. 2004;29(8):796–799, 818.
- [118] Duan J, Zhao Y. Antimicrobial efficiency of essential oil and freeze-thaw treatments against *Escherichia coli* O157:H7 and *Salmonella enterica* Ser. Enteritidis in strawberry juice. J Food Sci. 2009;74(3):M131–M137.
- [119] Dudonne S, Vitrac X, Coutiere P, Woillez M, Merillon JM. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. J Agric Food Chem. 2009.
- [120] Duessel S, Heuertz RM, Ezekiel UR. Growth inhibition of human colon cancer cells by plant compounds. Clin Lab Sci. 2008;21(3):151–157.
- [121] Dugoua JJ, Seely D, Perri D, Cooley K, Forelli T, Mills E, Koren G. From type 2 diabetes to antioxidant activity: a systematic review of the safety and efficacy of common and cassia cinnamon bark. Can J Physiol Pharmacol. 2007;85(9):837–847.
- [122] Dunham JP, Kelly S, Donaldson LF. Inflammation reduces mechanical thresholds in a population of transient receptor potential channel A1-expressing nociceptors in the rat. Eur J Neurosci. 2008;27(12):3151–3160.
- [123] Eid SR, Crown ED, Moore EL, Liang HA, Choong KC, Dima S, Henze DA, Kane SA, Urban MO. HC-030031, a TRPA1 selective antagonist, attenuates inflammatory- and neuropathy-induced mechanical hypersensitivity. Mol Pain 2008;4:48.
- [124] Elahi EN, Wright Z, Hinselwood D, Hotchkiss SA, Basketter DA, Pease CK. Protein binding and metabolism influence the relative skin sensitization potential of cinnamic compounds. Chem Res Toxicol. 2004;17(3):301–310.
- [125] Elshafie AE, Al Rashdi TA, Al Bahry SN, Bakheit CS. Fungi and aflatoxins associated with spices in the Sultanate of Oman. Mycopathologia 2002;155(3):155–160.
- [126] Endo H, Rees TD. Clinical features of cinnamon-induced contact stomatitis. Compend Contin Educ Dent. 2006;27(7):403–409.
- [127] Endo H, Rees TD. Cinnamon products as a possible etiologic factor in orofacial granulo-matosis. Med Oral Patol Oral Cir Bucal. 2007;12(6):E440–E444.
- [128] Epstein FW. Contact dermatitis due to cinnamon. Ohio Med 1950;46(7):659.
- [129] European Scientific Cooperative on Phytotherapy (ESCOP). Cinnamomi cortex. ESCOP Monographs, 2nd ed., Exeter: ESCOP, 2003, pp. 92–97.
- [130] Fabio A, Cermelli C, Fabio G, Nicoletti P, Quaglio P. Screening of the antibacterial effects of a variety of essential oils on microorganisms responsible for respiratory infections. Phytother Res. 2007;21(4):374–377.
- [131] Fairman KA, Curtiss FR. Call for comparative effectiveness research: lowering A1C with sitagliptin, saxagliptin, or cinnamon. J Manag Care Pharm. 2009;15(8):696–700.
- [132] Fang SH, Rao YK, Tzeng YM. Inhibitory effects of flavonol glycosides from Cinnamomum osmophloeum on inflammatory mediators in LPS/IFN-gamma-activated murine macrophages. Bioorg Med Chem. 2005;13(7):2381–2388.
- [133] Farkas J. Perioral dermatitis from marjoram, bay leaf and cinnamon. Contact Dermatitis 1981;7(2):121.
- [134] Feng GP, Zhang SD, Yi NY. [Effects of *Rehmannia glutinosa*, *Plastrum testudinis*, *Aconitum carmichaeli* and *Cinnamomum cassia* on the beta-adrenergic receptors of hyperthyroid rat kidneys]. Zhong Xi Yi Jie He Za Zhi. 1986;6(10):606–608, 582.
- [135] Fergurson J, Sharma S. Cinnamic aldehyde test concentrations. Contact Dermatitis 1984;10(3):191–192.
- [136] Ferme D, Banjac M, Calsamiglia S, Busquet M, Kamel C, Avgustin G. The effects of plant extracts on microbial community structure in a rumen-simulating continuous-culture system as revealed by molecular profiling. Folia Microbiol. (Praha) 2004;49(2):151–155.
- [137] Ferme D, Malnersic M, Lipoglavsek L, Kamel C, Avgustin G. Effect of sodium monensin and cinnamaldehyde on the growth and phenotypic characteristics of *Prevotella bryantii* and *Prevotella ruminicola*. Folia Microbiol. (Praha) 2008;53(3):204–208.
- [138] Filoche SK, Soma K, Sissons CH. Antimicrobial effects of essential oils in combination with chlorhexidine digluconate. Oral Microbiol Immunol. 2005;20(4):221–225.

- [139] Fink RC, Roschek B, Alberte RS. HIV type-1 entry inhibitors with a new mode of action. Antivir Chem Chemother. 2009;19(6):243–255.
- [140] Fisher AA, Dooms-Goossens A. The effect of perfume "ageing" on the allergenicity of individual perfume ingredients. Contact Dermatitis 1976;2(3):155–159.
- [141] Foti C, Bonamonte D, Conserva A, Stingeni L, Lisi P, Lionetti N, Rigano L, Angelini G. Allergic and photoallergic contact dermatitis from ketoprofen: evaluation of cross-reactivities by a combination of photopatch testing and computerized conformational analysis. Curr Pharm Des. 2008;14(27):2833–2839.
- [142] Fowles J, Mitchell J, McGrath H. Assessment of cancer risk from ethylene oxide residues in spices imported into New Zealand. Food Chem Toxicol. 2001;39(11):1055–1062.
- [143] Friedman M, Buick R, Elliott CT. Antibacterial activities of naturally occurring compounds against antibiotic-resistant *Bacillus cereus* vegetative cells and spores, *Escherichia coli*, and *Staphylococcus aureus*. J Food Prot. 2004;67(8):1774–1778.
- [144] Friedman M, Henika PR, Levin CE, Mandrell RE. Antibacterial activities of plant essential oils and their components against *Escherichia coli* O157:H7 and *Salmonella enterica* in apple juice. J Agric Food Chem. 2004;52(19):6042–6048.
- [145] Friedman M, Henika PR, Mandrell RE. Bactericidal activities of plant essential oils and some of their isolated constituents against *Campylobacter jejuni*, *Escherichia coli*, *Listeria monocytogenes*, and *Salmonella enterica*. J Food Prot. 2002;65(10):1545–1560.
- [146] Friedman M, Kozukue N, Harden LA. Cinnamaldehyde content in foods determined by gas chromatography-mass spectrometry. J Agric Food Chem. 2000;48(11):5702–5709.
- [147] Fujita, S. [Miscellaneous contributions to the essential oils of plants from various territories: XLVII. On the components of essential oils of *Cinnamomum sieboldii* Meisn]. Yakugaku Zasshi 1986;106(1):17–21.
- [148] Fundaro A, Cassone MC. [Action of essential oils of chamomile, cinnamon, absinthium, mace and origanum on operant conditioning behavior of the rat]. Boll Soc Ital Biol Sper. 1980;56(22):2375–2380.
- [149] Fuselli SR, Garcia De La Rosa SB, Gende LB, Eguaras MJ, Fritz R. [Inhibition of *Paeni-bacillus* larvae employing a mixture of essential oils and thymol]. Rev Argent Microbiol. 2006;38(2):89–92.
- [150] Futrell JM, Rietschel RL. Spice allergy evaluated by results of patch tests. Cutis 1993;52(5):288–290.
- [151] Garcia-Abujeta JL, de Larramendi CH, Berna JP, Palomino EM. Mud bath dermatitis due to cinnamon oil. Contact Dermatitis 2005;52(4):234.
- [152] Gardeta, P. [Consequences of the discovery of America on nutrition: the introduction of new foods in Europe]. Rev Med Chil. 1999;127(1):101–109.
- [153] Gill AO, Holley RA. Mechanisms of bactericidal action of cinnamaldehyde against *Listeria monocytogenes* and of eugenol against *L. monocytogenes* and *Lactobacillus sakei*. Appl Environ Microbiol. 2004;70(10):5750–5755.
- [154] Giordani R, Regli P, Kaloustian J, Portugal H. Potentiation of antifungal activity of amphotericin B by essential oil from *Cinnamomum cassia*. Phytother Res. 2006;20(1):58–61.
- [155] Girardin P, Vigan M, Humbert P, Aubin F. Cross-reactions in patch testing with ketoprofen, fragrance mix and cinnamic derivatives. Contact Dermatitis 2006;55(2):126–128.
- [156] Goh CL, Ng SK. Bullous contact allergy from cinnamon. Derm Beruf Umwelt. 1988;36(6):186–187.
- [157] Gollhausen R, Kligman AM. Human assay for identifying substances which induce non-allergic contact urticaria: the NICU-test. Contact Dermatitis 1985;13(2):98–106.
- [158] Gong F, Liang YZ, Fung YS. Analysis of volatile components from cortex Cinnamomi with hyphenated chromatography and chemometric resolution. J Pharm Biomed Anal. 2004;34(5):1029–1047.
- [159] Gowder SJ, Devaraj H. Effect of the food flavour cinnamaldehyde on the antioxidant status of rat kidney. Basic Clin Pharmacol Toxicol. 2006;99(5):379–382.
- [160] Gratzke C, Streng T, Waldkirch E, Sigl K, Stief C, Andersson KE, Hedlund P. Transient receptor potential A1 (TRPA1) activity in the human urethra evidence for a functional role for TRPA1 in the outflow region. Eur Urol. 2009;55(3):696–704.

- [161] Gu L, Kelm MA, Hammerstone JF, Zhang Z, Beecher G, Holden J, Haytowitz D, Prior RL. Liquid chromatographic/electrospray ionization mass spectrometric studies of proanthocyanidins in foods. J Mass Spectrom. 2003;38(12):1272–1280.
- [162] Guin JD, Meyer BN, Drake RD, Haffley P. The effect of quenching agents on contact urticaria caused by cinnamic aldehyde. J Am Acad Dermatol. 1984;10(1):45–51.
- [163] Guo JY, Huo HR, Zhao BS, Liu HB, Li LF, Guo SY, Jiang TL. Effect of 3-phenyl-2-propene-1-ol on PGE2 release from rat cerebral microvascular endothelial cells stimulated by IL-1beta. Am J Chin Med. 2006;34(4):685–693.
- [164] Gutierrez L, Escudero A, Batlle R, Nerin C. Effect of mixed antimicrobial agents and flavors in active packaging films. J Agric Food Chem. 2009;57(18):8564–8571.
- [165] Guynot ME, Ramos AJ, Seto L, Purroy P, Sanchis V, Marin S. Antifungal activity of volatile compounds generated by essential oils against fungi commonly causing deterioration of bakery products. J Appl Microbiol. 2003;94(5):893–899.
- [166] Ha KT, Kim JK, Kang SK, Kim DW, Lee YC, Kim HM, Kim CH. Inhibitory effect of Sihoga-Yonggol-Moryo-Tang on matrix metalloproteinase-2 and -9 activities and invasiveness potential of hepatocellular carcinoma. Pharmacol Res. 2004;50(3):279–285.
- [167] Haasch ML, Ford AW. Combined effects of ethanol and cinnamaldehyde in the Japanese medaka embryo-larval assay (MELA). Mar Environ Res. 2004;58(2–5):175–179.
- [168] Hafez HM, Hauck R. Efficacy of a herbal product against *Histomonas meleagridis* after experimental infection of turkey poults. Arch Anim Nutr. 2006;60(5):436–442.
- [169] Harada M, Fujii Y, Kamiya J. Pharmacological studies on chinese cinnamon: III. Electroencephalographic studies of cinnamaldehyde in the rabbit. Chem Pharm Bull. (Tokyo) 1976;24(8):1784–1788.
- [170] Harada M, Hirayama Y, Yamazaki R. Pharmacological studies on Chinese cinnamon: V. Catecholamine releasing effect of cinnamaldehyde in dogs. J Pharmacobiodyn. 1982;5(8):539–546.
- [171] Haranaka K, Satomi N, Sakurai A, Haranaka R, Okada N, Kobayashi M. Antitumor activities and tumor necrosis factor producibility of traditional Chinese medicines and crude drugs. Cancer Immunol Immunother. 1985;20(1):1–5.
- [172] Haring JI. Case #5. Cinnamon-induced stomatitis. RDH 1993;13(5):12, 50.
- [173] Hartmann K, Hunzelmann N. Allergic contact dermatitis from cinnamon as an odourneutralizing agent in shoe insoles. Contact Dermatitis 2004;50(4):253–254.
- [174] Hasan HA. Alternaria mycotoxins in black rot lesion of tomato fruit: conditions and regulation of their production. Acta Microbiol Immunol Hung. 1996;43(2–3):125–133.
- [175] Hasani-Ranjbar S, Larijani B, Abdollahi M. A systematic review of the potential herbal sources of future drugs effective in oxidant-related diseases. Inflamm Allergy Drug Targets. 2009;8(1):2–10.
- [176] Hasegawa A, Yoshino M, Nakamura H, Ishii I, Watanabe T, Kiuchi M, Ishikawa T, Ohmori S, Kitada M. Identification of inhibitory component in cinnamon–O-methoxycinnamaldehyde inhibits CYP1A2 and CYP2E1. Drug Metab Pharmacokinet. 2002;17(3):229–236.
- [177] Haslbeck F, Senser F, Grosch W. [Detection of low lipase activities in food]. Z Lebensm Unters Forsch. 1985;181(4):271–275.
- [178] Hatch GG, Anderson TM, Lubet RA, Kouri RE, Putman DL, Cameron JW, Nims RW, Most B, Spalding JW, Tennant RW. Chemical enhancement of SA7 virus transformation of hamster embryo cells: evaluation by interlaboratory testing of diverse chemicals. Environ Mutagen. 1986;8(4):515–531.
- [179] Hayashi K, Imanishi N, Kashiwayama Y, Kawano A, Terasawa K, Shimada Y, Ochiai H. Inhibitory effect of cinnamaldehyde, derived from Cinnamomi cortex, on the growth of influenza A/PR/8 virus in vitro and in vivo. Antiviral Res. 2007;74(1):1–8.
- [180] He ZD, Qiao CF, Han QB, Cheng CL, Xu HX, Jiang RW, But PP, Shaw PC. Authentication and quantitative analysis on the chemical profile of cassia bark (cortex Cinnamomi) by high-pressure liquid chromatography. J Agric Food Chem. 2005;53(7):2424–2428.

- [181] Hernandez-Herrero LA, Giner MJ, Valero M. Effective chemical control of psychrotrophic Bacillus cereus EPSO-35AS and INRA TZ415 spore outgrowth in carrot broth. Food Microbiol. 2008;25(5):714–721.
- [182] Hersch-Martinez P, Leanos-Miranda BE, Solorzano-Santos F. Antibacterial effects of commercial essential oils over locally prevalent pathogenic strains in Mexico. Fitoterapia 2005;76(5):453–457.
- [183] Hitokoto H, Morozumi S, Wauke T, Sakai S, Ueno I. Inhibitory effects of condiments and herbal drugs on the growth and toxin production of toxigenic fungi. Mycopathologia 1979;66(3):161–167.
- [184] Hlebowicz J, Darwiche G, Bjorgell O, Almer LO. Effect of cinnamon on postprandial blood glucose, gastric emptying, and satiety in healthy subjects. Am J Clin Nutr. 2007;85(6):1552–1556.
- [185] Hlebowicz J, Hlebowicz A, Lindstedt S, Bjorgell O, Hoglund P, Holst JJ, Darwiche G, Almer LO. Effects of 1 and 3 g cinnamon on gastric emptying, satiety, and postprandial blood glucose, insulin, glucose-dependent insulinotropic polypeptide, glucagon-like peptide 1, and ghrelin concentrations in healthy subjects. Am J Clin Nutr. 2009;89(3):815–821.
- [186] Ho SC, Tsai TH, Tsai PJ, Lin CC. Protective capacities of certain spices against peroxynitrite-mediated biomolecular damage. Food Chem Toxicol. 2008;46(3):920–928.
- [187] Hong CH, Hur SK, Oh OJ, Kim SS, Nam KA, Lee SK. Evaluation of natural products on inhibition of inducible cyclooxygenase (COX-2) and nitric oxide synthase (iNOS) in cultured mouse macrophage cells. J Ethnopharmacol. 2002;83(1–2):153–159.
- [188] Hong SH, Kim J, Kim JM, Lee SY, Shin DS, Son KH, Han DC, Sung YK, Kwon BM. Apoptosis induction of 2'-hydroxycinnamaldehyde as a proteasome inhibitor is associated with ER stress and mitochondrial perturbation in cancer cells. Biochem Pharmacol. 2007;74(4):557–565.
- [189] Hoskins JA. The occurrence, metabolism and toxicity of cinnamic acid and related compounds. J Appl Toxicol. 1984;4(6):283–292.
- [190] Hoskyn J, Guin JD. Contact allergy to cinnamal in a patient with oral lichen planus. Contact Dermatitis 2005;52(3):160–161.
- [191] Hsieh TJ, Liu TZ, Lu FJ, Hsieh PY, Chen CH. Actinodaphnine induces apoptosis through increased nitric oxide, reactive oxygen species and down-regulation of NF-kappaB signaling in human hepatoma Mahlavu cells. Food Chem Toxicol. 2006;44(3):344–354.
- [192] Hsieh TJ, Lu LH, Su CC. NMR spectroscopic, mass spectroscopic, X-ray crystallographic, and theoretical studies of molecular mechanics of natural products: farformolide B and sesamin. Biophys Chem. 2005;114(1):13–20.
- [193] Huang J, Wang S, Luo X, Xie Y, Shi X. Cinnamaldehyde reduction of platelet aggregation and thrombosis in rodents. Thromb Res. 2007;119(3):337–342.
- [194] Hurlimann AF, Wuthrich B. [Eating cinnamon in cinnamaldehyde allergy]. Hautarzt 1995;46(9):660-661.
- [195] Huss U, Ringbom T, Perera P, Bohlin L, Vasange M. Screening of ubiquitous plant constituents for COX-2 inhibition with a scintillation proximity based assay. J Nat Prod. 2002;65(11):1517–1521.
- [196] Hussain RA, Kim J, Hu TW, Pezzuto JM, Soejarto DD, Kinghorn AD. Isolation of a highly sweet constituent from Cinnamomum osmophloeum leaves. Planta Med 1986;(5):403–404.
- [197] Hwang SH, Choi YG, Jeong MY, Hong YM, Lee JH, Lim S. Microarray analysis of gene expression profile by treatment of Cinnamomi ramulus in lipopolysaccharide-stimulated BV-2 cells. Gene 2009;443(1–2):83–90.
- [198] Idle JR. Christmas gingerbread (Lebkuchen) and Christmas cheer review of the potential role of mood elevating amphetamine-like compounds formed in vivo and in furno. Prague Med Rep. 2005;106(1):27–38.
- [199] Ikawati Z, Wahyuono S, Maeyama K. Screening of several Indonesian medicinal plants for their inhibitory effect on histamine release from RBL-2H3 cells. J Ethnopharmacol 2001;75(2–3):249–256.

- [200] Imparl-Radosevich J, Deas S, Polansky MM, Baedke DA, Ingebritsen TS, Anderson RA, Graves DJ. Regulation of PTP-1 and insulin receptor kinase by fractions from cinnamon: implications for cinnamon regulation of insulin signalling. Horm Res. 1998;50(3):177–182.
- [201] Inouye S, Takizawa T, Yamaguchi H. Antibacterial activity of essential oils and their major constituents against respiratory tract pathogens by gaseous contact. J Antimicrob Chemother. 2001;47(5):565–573.
- [202] Inouye S, Yamaguchi H, Takizawa T. Screening of the antibacterial effects of a variety of essential oils on respiratory tract pathogens, using a modified dilution assay method. J Infect Chemother. 2001;7(4):251–254.
- [203] Ishidate M, Jr., Sofuni T, Yoshikawa K, Hayashi M, Nohmi T, Sawada M, Matsuoka A. Primary mutagenicity screening of food additives currently used in Japan. Food Chem Toxicol. 1984;22(8):623–636.
- [204] Iwasaki Y, Tanabe M, Kobata K, Watanabe T. TRPA1 agonists allyl isothiocyanate and cinnamaldehyde – induce adrenaline secretion. Biosci Biotechnol Biochem. 2008;72(10):2608–2614.
- [205] Jadad AR, Moore RA, Carroll D, Jenkinson C, Reynolds DJ, Gavaghan DJ, McQuay HJ. Assessing the quality of reports of randomized clinical trials: is blinding necessary? Control Clin Trials 1996;17(1):1–12.
- [206] Jantan I, Rafi IA, Jalil J. Platelet-activating factor (PAF) receptor-binding antagonist activity of Malaysian medicinal plants. Phytomedicine 2005;12(1-2):88-92.
- [207] Janzowski C, Glaab V, Mueller C, Straesser U, Kamp HG, Eisenbrand G. Alpha, beta-unsaturated carbonyl compounds: induction of oxidative DNA damage in mammalian cells. Mutagenesis 2003;18(5):465–470.
- [208] Jarvill-Taylor KJ, Anderson RA, Graves DJ. A hydroxychalcone derived from cinnamon functions as a mimetic for insulin in 3T3-L1 adipocytes. J Am Coll Nutr. 2001;20(4):327–336.
- [209] Jayaprakasha GK, Jagan Mohan, Rao L, Sakariah KK. Chemical composition of the flower oil of Cinnamomum zeylanicum Blume. J Agric Food Chem. 2000;48(9):4294–4295.
- [210] Jayaprakasha GK, Jagan Mohan, Rao L, Sakariah KK. Volatile constituents from Cinnamomum zeylanicum fruit stalks and their antioxidant activities. J Agric Food Chem. 2003;51(15):4344–4348.
- [211] Jayaprakasha GK, Ohnishi-Kameyama M, Ono H, Yoshida M, Jaganmohan, Rao L. Phenolic constituents in the fruits of *Cinnamomum zeylanicum* and their antioxidant activity. J Agric Food Chem. 2006;54(5):1672–1679.
- [212] Jayaprakasha GK, Rao LJ, Sakariah KK. Chemical composition of volatile oil from Cinnamomum zeylanicum buds. Z Naturforsch. [C.] 2002;57(11–12):990–993.
- [213] Jeong HW, Han DC, Son KH, Han MY, Lim JS, Ha JH, Lee CW, Kim HM, Kim HC, Kwon BM. Antitumor effect of the cinnamaldehyde derivative CB403 through the arrest of cell cycle progression in the G2/M phase. Biochem Pharmacol. 2003;65(8):1343–1350.
- [214] Josimovic L, Cudina I. Spectrophotometric analysis of irradiated spices. Int J Rad Appl Instrum. [A] 1987;38(4):269–274.
- [215] Juneja VK, Friedman M. Carvacrol, cinnamaldehyde, oregano oil, and thymol inhibit Clostridium perfringens spore germination and outgrowth in ground turkey during chilling. J Food Prot. 2007;70(1):218–222.
- [216] Ka H, Park HJ, Jung HJ, Choi JW, Cho KS, Ha J, Lee KT. Cinnamaldehyde induces apoptosis by ROS-mediated mitochondrial permeability transition in human promyelocytic leukemia HL-60 cells. Cancer Lett. 2003;196(2):143–152.
- [217] Kada T, Inoue T, Ohta T, Shirasu Y. Antimutagens and their modes of action. Basic Life Sci. 1986;39:181–196.
- [218] Kada T, Shirasu Y, Ikekawa N, Nomoto M. Detection of natural bio-antimutagens and in vivo and in vitro analysis of their action. Prog Clin Biol Res. 1986;209A:385–393.
- [219] Kamath JV, Rana AC, Chowdhury AR. Pro-healing effect of *Cinnamomum zeylanicum* bark. Phytother Res. 2003;17(8):970–972.
- [220] Kamei T, Kumano H, Iwata K, Nariai Y, Matsumoto T. The effect of a traditional Chinese prescription for a case of lung carcinoma. J Altern Complement Med. 2000;6(6):557–559.

- [221] Kanari M, Tomoda M, Gonda R, Shimizu N, Kimura M, Kawaguchi M, Kawabe C. A reticuloendothelial system-activating arabinoxylan from the bark of *Cinnamomum cassia*. Chem Pharm Bull. (Tokyo) 1989;37(12):3191–3194.
- [222] Kanerva L, Estlander T, Jolanki R. Occupational allergic contact dermatitis from spices. Contact Dermatitis 1996;35(3):157–162.
- [223] Kannappan S, Jayaraman T, Rajasekar P, Ravichandran MK, Anuradha CV. Cinnamon bark extract improves glucose metabolism and lipid profile in the fructose-fed rat. Singapore Med J. 2006;47(10):858–863.
- [224] Kanuri G, Weber S, Volynets V, Spruss A, Bischoff SC, Bergheim I. Cinnamon extract protects against acute alcohol-induced liver steatosis in mice. J Nutr. 2009;139(3):482–487.
- [225] Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. J Ethnopharmacol. 2003;84(1):105–108.
- [226] Kasamaki A, Urasawa S. Transforming potency of flavoring agents in Chinese hamster cells. J Toxicol Sci. 1985;10(3):177–185.
- [227] Kern AB. Contact dermatitis from cinnamon. Arch Dermatol. 1960;81:599–600.
- [228] Khan A, Bryden NA, Polansky MM, Anderson RA. Insulin potentiating factor and chromium content of selected foods and spices. Biol Trace Elem Res 1990;24(3):183–188.
- [229] Khan A, Safdar M, Ali Khan MM, Khattak KN, Anderson RA. Cinnamon improves glucose and lipids of people with type 2 diabetes. Diabetes Care 2003;26(12):3215–3218.
- [230] Kiec-Swierczynska M, Krecisz B, Swierczynska-Machura D. [Allergy to cosmetics: I. Fragrances]. Med Pract. 2004;55(2):203–206.
- [231] Kim DH, Kim CH, Kim MS, Kim JY, Jung KJ, Chung JH, An WG, Lee JW, Yu BP, Chung HY. Suppression of age-related inflammatory NF-kappaB activation by cinnamaldehyde. Biogerontology 2007;8(5):545–554.
- [232] Kim DS, Kim JY, Han YS. Alzheimer's disease drug discovery from herbs: neuroprotectivity from beta-amyloid (1–42) insult. J Altern Complement Med. 2007;13(3):333–340.
- [233] Kim SH, Hyun SH, Choung SY. Antioxidative effects of Cinnamomi cassiae and *Rhodiola rosea* extracts in liver of diabetic mice. Biofactors 2006;26(3):209–219.
- [234] Kim SI, Kim HK, Koh YY, Clark JM, Ahn YJ. Toxicity of spray and fumigant products containing cassia oil to *Dermatophagoides farinae* and *Dermatophagoides pteronyssinus* (Acari: Pyroglyphidae). Pest Manag Sci. 2006;62(8):768–774.
- [235] Kim SI, Yi JH, Tak JH, Ahn YJ. Acaricidal activity of plant essential oils against *Dermanyssus gallinae* (Acari: Dermanyssidae). Vet Parasitol. 2004;120(4):297–304.
- [236] Kim SJ, Han D, Moon KD, Rhee JS. Measurement of superoxide dismutase-like activity of natural antioxidants. Biosci Biotechnol Biochem. 1995;59(5):822–826.
- [237] Kim W, Khil LY, Clark R, Bok SH, Kim EE, Lee S, Jun HS, Yoon JW. Naphthalenemethyl ester derivative of dihydroxyhydrocinnamic acid, a component of cinnamon, increases glucose disposal by enhancing translocation of glucose transporter 4. Diabetologia 2006;49(10):2437–2448.
- [238] Kind F, Scherer K, Bircher AJ. Allergic contact stomatitis to cinnamon in chewing gum mistaken as facial angioedema. Allergy 2009.
- [239] King AA, Shaughnessy DT, Mure K, Leszczynska J, Ward WO, Umbach DM, Xu Z, Ducharme D, Taylor JA, Demarini DM, Klein CB. Antimutagenicity of cinnamaldehyde and vanillin in human cells: global gene expression and possible role of DNA damage and repair. Mutat Res. 2007;616(1–2):60–69.
- [240] Kirton V. Contact urticaria and cinnamic aldehyde. Contact Dermatitis 1978;4(6):374–375.
- [241] Kirton V, Wilkinson DS. Sensitivity to cinnamic aldehyde in a toothpaste: 2. Further studies. Contact Dermatitis 1975;1(2):77–80.
- [242] Kleefstra N, Logtenberg SJ, Houweling ST, Verhoeven S, Bilo HJ. [Cinnamon: not suitable for the treatment of diabetes mellitus]. Ned Tijdschr Geneeskd. 2007;151(51):2833–2837.
- [243] Knight KP, McKellar RC. Influence of cinnamon and clove essential oils on the D- and z-values of *Escherichia coli* O157:H7 in apple cider. J Food Prot. 2007;70(9):2089–2094.

- [244] Koh WS, Yoon SY, Kwon BM, Jeong TC, Nam KS, Han MY. Cinnamaldehyde inhibits lymphocyte proliferation and modulates T-cell differentiation. Int J Immunopharmacol. 1998;20(11):643–660.
- [245] Koller WD. [The importance of temperature on storage of ground natural spices]. Z Lebensm Unters Forsch. 1976;160(2):143–147.
- [246] Kong B, Wang J, Xiong YL. Antimicrobial activity of several herb and spice extracts in culture medium and in vacuum-packaged pork. J Food Prot. 2007;70(3):641–647.
- [247] Kong JO, Lee SM, Moon YS, Lee SG, Ahn YJ. Nematicidal activity of cassia and cinnamon oil compounds and related compounds toward Bursaphelenchus xylophilus (Nematoda: Parasitaphelenchidae). J Nematol. 2007;39(1):31–36.
- [248] Kong LD, Cai Y, Huang WW, Cheng CH, Tan RX. Inhibition of xanthine oxidase by some Chinese medicinal plants used to treat gout. J Ethnopharmacol. 2000;73(1–2):199–207.
- [249] Kong YH, Jo YO, Cho CW, Son D, Park S, Rho J, Choi SY. Inhibitory effects of cinnamic acid on melanin biosynthesis in skin. Biol Pharm Bull. 2008;31(5):946–948.
- [250] Kosugi M, Nakatsuka T, Fujita T, Kuroda Y, Kumamoto E. Activation of TRPA1 channel facilitates excitatory synaptic transmission in substantia gelatinosa neurons of the adult rat spinal cord. J Neurosci. 2007;27(16):4443–4451.
- [251] Kovalenko S, Belenichev I, Nikitin V, Karpenko A. Search for substances with antioxidant and antiamnestic activities among 2-substituted 4-(3H)-quinazolones. Acta Pol Pharm. 2003;60(4):275–279.
- [252] Kreydiyyeh SI, Usta J, Copti R. Effect of cinnamon, clove and some of their constituents on the Na(+)-K(+)-ATPase activity and alanine absorption in the rat jejunum. Food Chem Toxicol. 2000;38(9):755–762.
- [253] Kuo PL, Chen CY, Tzeng TF, Lin CC, Hsu YL. Involvement of reactive oxygen species/c-Jun NH(2)-terminal kinase pathway in kotomolide A induces apoptosis in human breast cancer cells. Toxicol Appl Pharmacol. 2008;229(2):215–226.
- [254] Kuo SY, Hsieh TJ, Wang YD, Lo WL, Hsui YR, Chen CY. Cytotoxic constituents from the leaves of *Cinnamomum subavenium*. Chem Pharm Bull. (Tokyo) 2008;56(1):97–101.
- [255] Kuo YC, Lu CK, Huang LW, Kuo YH, Chang C, Hsu FL, Lee TH. Inhibitory effects of acylated kaempferol glycosides from the leaves of *Cinnamomum kotoense* on the proliferation of human peripheral blood mononuclear cells. Planta Med. 2005;71(5):412–415.
- [256] Kurokawa M, Kumeda CA, Yamamura J, Kamiyama T, Shiraki K. Antipyretic activity of cinnamyl derivatives and related compounds in influenza virus-infected mice. Eur J Pharmacol. 1998;348(1):45–51.
- [257] Kwon BM, Lee SH, Choi SU, Park SH, Lee CO, Cho YK, Sung ND, Bok SH. Synthesis and in vitro cytotoxicity of cinnamaldehydes to human solid tumor cells. Arch Pharm Res. 1998;21(2):147–152.
- [258] Kwon HK, Jeon WK, Hwang JS, Lee CG, So JS, Park JA, Ko BS, Im SH. Cinnamon extract suppresses tumor progression by modulating angiogenesis and the effector function of CD8+ T cells. Cancer Lett. 2009;278(2):174–182.
- [259] Kwon JA, Yu CB, Park HD. Bacteriocidal effects and inhibition of cell separation of cinnamic aldehyde on *Bacillus cereus*. Lett Appl Microbiol. 2003;37(1):61–65.
- [260] Lahti A. Terfenadine does not inhibit non-immunologic contact urticaria. Contact Dermatitis 1987;16(4):220–223.
- [261] Lahti A, Maibach HI. An animal model for nonimmunologic contact urticaria. Toxicol Appl Pharmacol. 1984;76(2):219–224.
- [262] Lahti A, Maibach HI. Species specificity of nonimmunologic contact urticaria: guinea pig, rat, and mouse. J Am Acad Dermatol. 1985;13(1):66–69.
- [263] Lahti A, McDonald DM, Tammi R, Maibach HI. Pharmacological studies on nonimmunologic contact urticaria in guinea pigs. Arch Dermatol Res. 1986;279(1):44–49.
- [264] Lahti A, Vaananen A, Kokkonen EL, Hannuksela M. Acetylsalicylic acid inhibits nonimmunologic contact urticaria. Contact Dermatitis 1987;16(3):133–135.
- [265] Lai PK, Roy J. Antimicrobial and chemopreventive properties of herbs and spices. Curr Med Chem 2004;11(11):1451–1460.

- [266] Lamey PJ, Lewis MA, Rees TD, Fowler C, Binnie WH, Forsyth A. Sensitivity reaction to the cinnamonal dehyde component of toothpaste. Br Dent J. 1990;168(3):115–118.
- [267] Larsen WG. Perfume dermatitis. J Am Acad Dermatol. 1985;12(1 Pt 1):1–9.
- [268] Lazarus SA, Adamson GE, Hammerstone JF, Schmitz HH. High-performance liquid chromatography/mass spectrometry analysis of proanthocyanidins in foods and beverages. J Agric Food Chem. 1999;47(9):3693–3701.
- [269] Lee CW, Hong DH, Han SB, Park SH, Kim HK, Kwon BM, Kim HM. Inhibition of human tumor growth by 2'-hydroxy- and 2'-benzoyloxycinnamaldehydes. Planta Med. 1999:65(3):263–266.
- [270] Lee EJ, Kim JR, Choi DR, Ahn YJ. Toxicity of cassia and cinnamon oil compounds and cinnamaldehyde-related compounds to *Sitophilus oryzae* (Coleoptera: Curculionidae). J Econ Entomol. 2008;101(6):1960–1966.
- [271] Lee HS. Inhibitory activity of *Cinnamomum cassia* bark-derived component against rat lens aldose reductase. J Pharm Pharm Sci. 2002;5(3):226–230.
- [272] Lee HS, Kim BS, Kim MK. Suppression effect of *Cinnamomum cassia* bark-derived component on nitric oxide synthase. J Agric Food Chem. 2002;50(26):7700–7703.
- [273] Lee JS, Jeon SM, Park EM, Huh TL, Kwon OS, Lee MK, Choi MS. Cinnamate supplementation enhances hepatic lipid metabolism and antioxidant defense systems in high cholesterol-fed rats. J Med Food. 2003;6(3):183–191.
- [274] Lee JY, Kang HS, Park BE, Moon HJ, Sim SS, Kim CJ. Inhibitory effects of Geijigajakyak-Tang on trinitrobenzene sulfonic acid-induced colitis. J Ethnopharmacol. 2009;126:244–251.
- [275] Lee KG, Shibamoto T. Determination of antioxidant potential of volatile extracts isolated from various herbs and spices. J Agric Food Chem. 2002;50(17):4947–4952.
- [276] Lee KK, Kim JH, Cho JJ, Choi JD. Inhibitory effects of 150 plant extracts on elastase activity and their anti-inflammatory effects. Int J Cosmet Sci. 1999;21(2):71–82.
- [277] Lee R, Balick MJ. Sweet wood cinnamon and its importance as a spice and medicine. Explore. (NY) 2005;1(1):61–64.
- [278] Lee SH, Lee SY, Son DJ, Lee H, Yoo HS, Song S, Oh KW, Han DC, Kwon BM, Hong JT. Inhibitory effect of 2'-hydroxycinnamaldehyde on nitric oxide production through inhibition of NF-kappa B activation in RAW 264.7 cells. Biochem Pharmacol. 2005;69(5):791–799.
- [279] Leifer W. Contact dermatitis due to cinnamon: recurrence of dermatitis following oral administration of cinnamon oil. AMA Arch Derm Syphilol. 1951;64(1):52–55.
- [280] Li CH, Zhou J, Huo HR, Kang XL, Jiang TL. [Effect of guizhi tang and its active components on the fever induced by EP3 agonist]. Zhongguo ZhongYao Za Zhi. 2003;28(11):1056–1060.
- [281] Li TJ, Qiu Y, Mao JQ, Yang PY, Rui YC, Chen WS. Protective effects of Guizhi-Fuling capsules on rat brain ischemia/reperfusion injury. J Pharmacol Sci. 2007;105(1):34–40.
- [282] Li XR, Liang YZ, Li XN. [Analysis of essential oil in *Herba ephedrae*-ramulus Cinnamomi by GC-MS and chemometric resolution method]. Yao Xue Xue Bao 2007;42(2):187–191.
- [283] Liao BC, Hsieh CW, Liu YC, Tzeng TT, Sun YW, Wung BS. Cinnamaldehyde inhibits the tumor necrosis factor-alpha-induced expression of cell adhesion molecules in endothelial cells by suppressing NF-kappaB activation: effects upon IkappaB and Nrf2. Toxicol Appl Pharmacol. 2008;229(2):161–171.
- [284] Lijinsky W, Andrews AW. Mutagenicity of vinyl compounds in *Salmonella typhimurium*. Teratog Carcinog Mutagen. 1980;1(3):259–267.
- [285] Lima EO, Gompertz OF, Giesbrecht AM, Paulo MQ. In vitro antifungal activity of essential oils obtained from officinal plants against dermatophytes. Mycoses 1993;36(9–10):333–336.
- [286] Lin CC, Wu SJ, Chang CH, Ng LT. Antioxidant activity of *Cinnamomum cassia*. Phytother Res. 2003;17(7):726–730.
- [287] Lin CT, Chen CJ, Lin TY, Tung JC, Wang SY. Anti-inflammation activity of fruit essential oil from *Cinnamomum insularimontanum* Hayata. Bioresour Technol. 2008;99(18):8783–8787.

- [288] Lin RJ, Cheng MJ, Huang JC, Lo WL, Yeh YT, Yen CM, Lu CM, Chen CY. Cytotoxic compounds from the stems of *Cinnamomum tenuifolium*. J Nat Prod. 2009;72:1816–1824.
- [289] Lin RJ, Lo WL, Wang YD, Chen CY. A novel cytotoxic monoterpenoid from the leaves of *Cinnamomum subavenium*. Nat Prod Res. 2008;22(12):1055–1059.
- [290] Lirussi D, Li J, Prieto JM, Gennari M, Buschiazzo H, Rios JL, Zaidenberg A. Inhibition of *Trypanosoma cruzi* by plant extracts used in Chinese medicine. Fitoterapia 2004;75(7–8):718–723.
- [291] Liu CH, Mishra AK, Tan RX, Tang C, Yang H, Shen YF. Repellent and insecticidal activities of essential oils from *Artemisia princeps* and *Cinnamomum camphora* and their effect on seed germination of wheat and broad bean. Bioresour Technol. 2006;97(15):1969–1973.
- [292] Liu HY, Li WY, Hao TG, Gao HY, Ma Y. [Quantitative study of effects of cinnamaldehyde on levels of endotoxin in root canals in rats' periapical periodontitis model]. Hua Xi Kou Qiang Yi Xue Za Zhi. 2007;25(3):233–236.
- [293] Liu TZ, Cheng JT, Yiin SJ, Chen CY, Chen CH, Wu MJ, Chern CL. Isoobtusilactone A induces both caspase-dependent and -independent apoptosis in Hep G2 cells. Food Chem Toxicol. 2008;46(1):321–327.
- [294] Llewellyn GC, Burkett ML, Eadie T. Potential mold growth, aflatoxin production, and antimycotic activity of selected natural spices and herbs. J Assoc Off Anal Chem. 1981;64(4):955–960.
- [295] Lockwood GB. Phenylpropanoids from a Nigerian sample of *Cinnamomum cassia* [proceedings]. J Pharm Pharmacol. 1979;31(Suppl):8P.
- [296] Lopez P, Sanchez C, Batlle R, Nerin C. Development of flexible antimicrobial films using essential oils as active agents. J Agric Food Chem. 2007;55(21):8814–8824.
- [297] Lopez P, Sanchez C, Batlle R, Nerin C. Vapor-phase activities of cinnamon, thyme, and oregano essential oils and key constituents against foodborne microorganisms. J Agric Food Chem. 2007;55(11):4348–4356.
- [298] Ludera-Zimoch, G. [Case of urticaria with immediate local and generalized reaction to cinnamon oil and benzaldehyde]. Przegl Dermatol. 1981;68(1):67–70.
- [299] Lungarini S, Aureli F, Coni E. Coumarin and cinnamaldehyde in cinnamon marketed in Italy: a natural chemical hazard? Food Addit Contam Part A Chem Anal Control Expo Risk Assess. 2008;25(11):1297–1305.
- [300] Luo H, Lin S, Ren F, Wu L, Chen L, Sun Y. Antioxidant and antimicrobial capacity of Chinese medicinal herb extracts in raw sheep meat. J Food Prot. 2007;70(6):1440–1445.
- [301] Mabrouk SS, El Shayeb NM. Inhibition of aflatoxin formation by some spices. Z Lebensm Unters Forsch. 1980;171(5):344–347.
- [302] MacDonald S, Castle L. A UK retail survey of aflatoxins in herbs and spices and their fate during cooking. Food Addit Contam. 1996;13(1):121–128.
- [303] Magnusson B, Wilkinson DS. Cinnamic aldehyde in toothpaste: 1. Clinical aspects and patch tests. Contact Dermatitis 1975;1(2):70–76.
- [304] Maibach HI. Cheilitis: occult allergy to cinnamic aldehyde. Contact Dermatitis 1986;15(2):106–107.
- [305] Maisey J, Miller K. Assessment of the ability of mice fed on vitamin A supplemented diet to respond to a variety of potential contact sensitizers. Contact Dermatitis 1986;15(1):17–23.
- [306] Majeti VA, Suskind RR. Mechanism of cinnamaldehyde sensitization. Contact Dermatitis 1977;3(1):16–18.
- [307] Malten KE, van Ketel WG, Nater JP, Liem DH. Reactions in selected patients to 22 fragrance materials. Contact Dermatitis 1984;11(1):1–10.
- [308] Mancini-Filho J, Van Koiij A, Mancini DA, Cozzolino FF, Torres RP. Antioxidant activity of cinnamon (*Cinnamomum zeylanicum* Breyne) extracts. Boll Chim Farm. 1998;137(11):443–447.
- [309] Mang B, Wolters M, Schmitt B, Kelb K, Lichtinghagen R, Stichtenoth DO, Hahn A. Effects of a cinnamon extract on plasma glucose, HbA, and serum lipids in diabetes mellitus type 2. Eur J Clin Invest. 2006;36(5):340–344.

- [310] Manzanilla EG, Perez JF, Martin M, Kamel C, Baucells F, Gasa J. Effect of plant extracts and formic acid on the intestinal equilibrium of early-weaned pigs. J Anim Sci. 2004;82(11):3210–3218.
- [311] Maralhas A, Monteiro A, Martins C, Kranendonk M, Laires A, Rueff J, Rodrigues AS. Genotoxicity and endoreduplication inducing activity of the food flavouring eugenol. Mutagenesis 2006;21(3):199–204.
- [312] Margineanu DG, Katona E, Popa J. Kinetics of nerve impulse blocking by protein cross-linking aldehydes: apparent critical thermal points. Biochim Biophys Acta 1981:649(3):581–586.
- [313] Marongiu B, Piras A, Porcedda S, Tuveri E, Sanjust E, Meli M, Sollai F, Zucca P, Rescigno A. Supercritical CO₂ extract of *Cinnamomum zeylanicum*: chemical characterization and antityrosinase activity. J Agric Food Chem. 2007;55(24):10022–10027.
- [314] Martin KW, Ernst E. Herbal medicines for treatment of bacterial infections: a review of controlled clinical trials. J Antimicrob Chemother. 2003;51(2):241–246.
- [315] Marzulli FN, Maibach HI. Effects of vehicles and elicitation concentration in contact dermatitis testing: I. Experimental contact sensitization in humans. Contact Dermatitis 1976;2(6):325–329.
- [316] Marzulli FN, Maibach HI. Further studies of effects of vehicles and elicitation concentration in experimental contact sensitization testing in humans. Contact Dermatitis 1980;6(2):131–133.
- [317] Mastura M, Nor Azah MA, Khozirah S, Mawardi R, Manaf AA. Anticandidal and antidermatophytic activity of *Cinnamomum* species essential oils. Cytobios 1999;98(387):17–23.
- [318] Mathew S, Abraham TE. In vitro antioxidant activity and scavenging effects of Cinnamomum verum leaf extract assayed by different methodologies. Food Chem Toxicol. 2006;44(2):198–206.
- [319] Mathias CG, Chappler RR, Maibach HI. Contact urticaria from cinnamic aldehyde. Arch Dermatol. 1980;116(1):74–76.
- [320] Matsuda H, Matsuda R, Fukuda S, Shiomoto H, Kubo M. Anti-thrombic actions of 70% methanolic extract and cinnamic aldehyde from cinnamomi cortex. Chem Pharm Bull. (Tokyo) 1987;35(3):1275–1280.
- [321] Mayaud L, Carricajo A, Zhiri A, Aubert G. Comparison of bacteriostatic and bactericidal activity of 13 essential oils against strains with varying sensitivity to antibiotics. Lett Appl Microbiol. 2008;47(3):167–173.
- [322] McCarty MF. Nutraceutical resources for diabetes prevention an update. Med Hypotheses 2005;64(1):151–158.
- [323] McCarty MF. Toward prevention of Alzheimers disease potential nutraceutical strategies for suppressing the production of amyloid beta peptides. Med Hypotheses 2006;62(3):139–48.
- [324] McKemy DD. How cold is it? TRPM8 and TRPA1 in the molecular logic of cold sensation. Mol Pain 2005;1(1):16.
- [325] Meding B. Skin symptoms among workers in a spice factory. Contact Dermatitis 1993;29(4):202–205.
- [326] Merrill AW, Cuellar JM, Judd JH, Carstens MI, Carstens E. Effects of TRPA1 agonists mustard oil and cinnamaldehyde on lumbar spinal wide-dynamic range neuronal responses to innocuous and noxious cutaneous stimuli in rats. J Neurophysiol. 2008;99(2):415–425.
- [327] Mi ZG, Liu SX, Cao YQ, Yang XF. [Adjustive effect of yi qi tong lin chongji on the three growth factors in rats prostatic tissues]. Zhongguo Zhong Yao Za Zhi. 2003;28(7):653–655.
- [328] Mihail RC. Oral leukoplakia caused by cinnamon food allergy. J Otolaryngol. 1992;21(5):366–367.
- [329] Miller RL, Gould AR, Bernstein ML. Cinnamon-induced stomatitis venenata: clinical and characteristic histopathologic features. Oral Surg Oral Med Oral Pathol 1992;73(6):708–716.
- [330] Mino Y, Ota N. Inorganic chemical approaches to pharmacognosy: V. X-ray fluorescence spectrometric studies on the inorganic constituents of crude drugs. Chem Pharm Bull. (Tokyo) 1990;38(3):709–713.

- [331] Mishra AK, Mishra A, Kehri HK, Sharma B, Pandey AK. Inhibitory activity of Indian spice plant *Cinnamomum zeylanicum* extracts against *Alternaria solani* and *Curvularia lunata*, the pathogenic dematiaceous moulds. Ann Clin Microbiol Antimicrob. 2009;8:9.
- [332] Miyazawa M, Hashimoto Y, Taniguchi Y, Kubota K. Headspace constituents of the tree remain of *Cinnamomum camphora*. Nat Prod Lett. 2001;15(1):63–69.
- [333] Moon EY, Lee MR, Wang AG, Lee JH, Kim HC, Kim HM, Kim JM, Kwon BM, Yu DY. Delayed occurrence of H-ras12V-induced hepatocellular carcinoma with long-term treatment with cinnamaldehydes. Eur J Pharmacol. 2006;530(3):270–275.
- [334] Moon KH, Pack MY. Cytotoxicity of cinnamic aldehyde on leukemia L1210 cells. Drug Chem Toxicol. 1983;6(6):521–535.
- [335] Morozumi S. Isolation, purification, and antibiotic activity of o-methoxycinnamaldehyde from cinnamon. Appl Environ Microbiol. 1978;36(4):577–583.
- [336] Moselhy SS, Ali HK. Hepatoprotective effect of cinnamon extracts against carbon tetrachloride induced oxidative stress and liver injury in rats. Biol Res. 2009;42(1):93–98.
- [337] Moskaug JO, Borge GI, Fagervoll AM, Paur I, Carlsen H, Blomhoff R. Dietary polyphenols identified as intracellular protein kinase A inhibitors. Eur J Nutr. 2008;47(8):460–469.
- [338] Mosqueda-Melgar J, Raybaudi-Massilia RM, Martin-Belloso O. Combination of highintensity pulsed electric fields with natural antimicrobials to inactivate pathogenic microorganisms and extend the shelf-life of melon and watermelon juices. Food Microbiol. 2008;25(3):479–491.
- [339] Mosqueda-Melgar J, Raybaudi-Massilia RM, Martin-Belloso O. Inactivation of Salmonella enterica Ser. Enteritidis in tomato juice by combining of high-intensity pulsed electric fields with natural antimicrobials. J Food Sci. 2008;73(2):M47– M53.
- [340] Murcia MA, Egea I, Romojaro F, Parras P, Jimenez AM, Martinez-Tome M. Antioxidant evaluation in dessert spices compared with common food additives: influence of irradiation procedure. J Agric Food Chem. 2004;52(7):1872–1881.
- [341] Nadiminti H, Ehrlich A, Udey MC. Oral erosions as a manifestation of allergic contact sensitivity to cinnamon mints. Contact Dermatitis 2005;52(1):46–47.
- [342] Nagai H, Shimazawa T, Matsuura N, Koda A. Immunopharmacological studies of the aqueous extract of *Cinnamomum cassia* (CCAq): I. Anti-allergic action. Jpn J Pharmacol. 1982;32(5):813–822.
- [343] Nagai H, Shimazawa T, Takizawa T, Koda A, Yagi A, Nishioka I. Immunopharmacological studies of the aqueous extract of *Cinnamomum cassia* (CCAq): II. Effect of CCAq on experimental glomerulonephritis. Jpn J Pharmacol. 1982;32(5):823–831.
- [344] Nahas R, Moher M. Complementary and alternative medicine for the treatment of type 2 diabetes. Can Fam Physician 2009;55(6):591–596.
- [345] Nair S, Nagar R, Gupta R. Antioxidant phenolics and flavonoids in common Indian foods. J Assoc Physicians India 1998;46(8):708–710.
- [346] Namer B, Kleggetveit IP, Handwerker H, Schmelz M, Jorum E. Role of TRPM8 and TRPA1 for cold allodynia in patients with cold injury. Pain 2008;139(1):63–72.
- [347] Nassenstein C, Kwong K, Taylor-Clark T, Kollarik M, Macglashan DM, Braun A, Undem BJ. Expression and function of the ion channel TRPA1 in vagal afferent nerves innervating mouse lungs. J Physiol. 2008;586(6):1595–1604.
- [348] Nater JP, De Jong MC, Baar AJ, Bleumink E. Contact urticarial skin responses to cinnamaldehyde. Contact Dermatitis 1977;3(3):151–154.
- [349] Ng TP, Chiam PC, Lee T, Chua HC, Lim L, Kua EH. Curry consumption and cognitive function in the elderly. Am J Epidemiol. 2006;164(9):898–906.
- [350] Ngoc TM, Lee I, Ha, do T, Kim H, Min B, Bae K. Tyrosinase-inhibitory constituents from the twigs of *Cinnamomum cassia*. J Nat Prod. 2009;72(6):1205–1208.
- [351] Nguyen SH, Dang TP, MacPherson C, Maibach H, Maibach HI. Prevalence of patch test results from 1970 to 2002 in a multi-centre population in North America (NACDG). Contact Dermatitis 2008;58(2):101–106.
- [352] Niinimaki A. Delayed-type allergy to spices. Contact Dermatitis 1984;11(1):34–40.

- [353] Niinimaki A. Double-blind placebo-controlled peroral challenges in patients with delayed-type allergy to balsam of Peru. Contact Dermatitis 1995;33(2):78–83.
- [354] Nir Y, Potasman I, Stermer E, Tabak M, Neeman I. Controlled trial of the effect of cinnamon extract on *Helicobacter pylori*. Helicobacter 2000;5(2):94–97.
- [355] Nishida S, Kikuichi S, Yoshioka S, Tsubaki M, Fujii Y, Matsuda H, Kubo M, Irimajiri K. Induction of apoptosis in HL-60 cells treated with medicinal herbs. Am J Chin Med. 2003;31(4):551–562.
- [356] Niu C, Afre S, Gilbert ES. Subinhibitory concentrations of cinnamaldehyde interfere with quorum sensing. Lett Appl Microbiol. 2006;43(5):489–494.
- [357] Niu C, Gilbert ES. Colorimetric method for identifying plant essential oil components that affect biofilm formation and structure. Appl Environ Microbiol. 2004;70(12):6951–6956.
- [358] Nixon R. Cinnamon allergy in a baker. Australas J Dermatol. 1995;36(1):41.
- [359] Noonan V, Kemp S. Cinnamon stomatitis. J Mass Dent Soc. 2007;56(3):43.
- [360] Nozawa K, Kawabata-Shoda E, Doihara H, Kojima R, Okada H, Mochizuki S, Sano Y, Inamura K, Matsushime H, Koizumi T, Yokoyama T, Ito H. TRPA1 regulates gastrointestinal motility through serotonin release from enterochromaffin cells. Proc Natl Acad Sci USA 2009;106(9):3408–3413.
- [361] Nuryastuti T, van der Mei HC, Busscher HJ, Iravati S, Aman AT, Krom BP. Effect of cinnamon oil on icaA expression and biofilm formation by *Staphylococcus epidermidis*. Appl Environ Microbiol. 2009;75:6850–6855.
- [362] O'Keefe JH, Gheewala NM, O'Keefe JO. Dietary strategies for improving postprandial glucose, lipids, inflammation, and cardiovascular health. J Am Coll Cardiol. 2008;51(3):249–255.
- [363] Obaidat MM, Frank JF. Inactivation of *Salmonella* and *Escherichia coli* O157:H7 on sliced and whole tomatoes by allyl isothiocyanate, carvacrol, and cinnamaldehyde in vapor phase. J Food Prot. 2009;72(2):315–324.
- [364] Ohta T, Watanabe K, Moriya M, Shirasu Y, Kada T. Analysis of the antimutagenic effect of cinnamaldehyde on chemically induced mutagenesis in *Escherichia coli*. Mol Gen Genet. 1983;192(3):309–315.
- [365] Ohta T, Watanabe K, Moriya M, Shirasu Y, Kada T. Antimutagenic effects of cinnamaldehyde on chemical mutagenesis in *Escherichia coli*. Mutat Res. 1983;107(2):219–227.
- [366] Okawa M, Kinjo J, Nohara T, Ono M. DPPH (1,1-diphenyl-2-picrylhydrazyl) radical scavenging activity of flavonoids obtained from some medicinal plants. Biol Pharm Bull. 2001;24(10):1202–1205.
- [367] Onderoglu S, Sozer S, Erbil KM, Ortac R, Lermioglu F. The evaluation of long-term effects of cinnamon bark and olive leaf on toxicity induced by streptozotocin administration to rats. J Pharm Pharmacol. 1999;51(11):1305–1312.
- [368] Orihara Y, Hamamoto H, Kasuga H, Shimada T, Kawaguchi Y, Sekimizu K. A silkworm baculovirus model for assessing the therapeutic effects of antiviral compounds: characterization and application to the isolation of antivirals from traditional medicines. J Gen Virol. 2008;89(Pt 1):188–194.
- [369] Osawa K, Matsumoto T, Yasuda H, Kato T, Naito Y, Okuda K. The inhibitory effect of plant extracts on the collagenolytic activity and cytotoxicity of human gingival fibroblasts by *Porphyromonas gingivalis* crude enzyme. Bull Tokyo Dent Coll. 1991;32(1):1–7.
- [370] Oussalah M, Caillet S, Salmieri S, Saucier L, Lacroix M. Antimicrobial effects of alginate-based film containing essential oils for the preservation of whole beef muscle. J Food Prot. 2006;69(10):2364–2369.
- [371] Oussalah M, Caillet S, Salmieri S, Saucier L, Lacroix M. Antimicrobial effects of alginate-based films containing essential oils on *Listeria monocytogenes* and *Salmonella* typhimurium present in bologna and ham. J Food Prot. 2007;70(4):901–908.
- [372] Panickar KS, Polansky MM, Anderson RA. Cinnamon polyphenols attenuate cell swelling and mitochondrial dysfunction following oxygen-glucose deprivation in glial cells. Exp Neurol. 2009;216(2):420–427.
- [373] Patton DW, Ferguson MM, Forsyth A, James J. Oro-facial granulomatosis: a possible allergic basis. Br J Oral Maxillofac Surg. 1985;23(4):235–242.

- [374] Pauli A, Knobloch K. Inhibitory effects of essential oil components on growth of foodcontaminating fungi. Z Lebensm Unters Forsch. 1987;185(1):10–13.
- [375] Perry PA, Dean BS, Krenzelok EP. Cinnamon oil abuse by adolescents. Vet Hum Toxicol. 1990;32(2):162–164.
- [376] Perry TL, Yong VW, Clavier RM, Jones K, Wright JM, Foulks JG, Wall RA. Partial protection from the dopaminergic neurotoxin N-methyl-4-phenyl-1,2,3,6-tetrahydropyridine by four different antioxidants in the mouse. Neurosci Lett. 1985;60(2):109–114.
- [377] Peterson DW, George RC, Scaramozzino F, LaPointe NE, Anderson RA, Graves DJ, Lew J. Cinnamon extract inhibits tau aggregation associated with Alzheimer's disease in vitro. J Alzheimers Dis. 2009;17:585–597.
- [378] Pham AQ, Kourlas H, Pham DQ. Cinnamon supplementation in patients with type 2 diabetes mellitus. Pharmacotherapy 2007;27(4):595–599.
- [379] Pilapil VR. Toxic manifestations of cinnamon oil ingestion in a child. Clin Pediatr. (Phila.) 1989;28(6):276.
- [380] Pina-Perez MC, Silva-Angulo AB, Muguerza-Marquinez B, Aliaga DR, Lopez AM. Syner-gistic effect of high hydrostatic pressure and natural antimicrobials on inactivation kinetics of *Bacillus cereus* in a liquid whole egg and skim milk mixed beverage. Foodborne Pathog Dis. 2009;6(6):649–656.
- [381] Placzek M, Fromel W, Eberlein B, Gilbertz KP, Przybilla B. Evaluation of phototoxic properties of fragrances. Acta Derm Venereol. 2007;87(4):312–316.
- [382] Prabuseenivasan S, Jayakumar M, Ignacimuthu S. In vitro antibacterial activity of some plant essential oils. BMC Complement Altern Med 2006;6:39.
- [383] Prakash D, Suri S, Upadhyay G, Singh BN. Total phenol, antioxidant and free radical scavenging activities of some medicinal plants. Int J Food Sci Nutr. 2007;58(1):18–28.
- [384] Prasad NS, Raghavendra R, Lokesh BR, Naidu KA. Spice phenolics inhibit human PMNL 5-lipoxygenase. Prostaglandins Leukot Essent Fatty Acids 2004;70(6):521–528.
- [385] Premanathan M, Rajendran S, Ramanathan T, Kathiresan K, Nakashima H, Yamamoto N. A survey of some Indian medicinal plants for anti-human immunodeficiency virus (HIV) activity. Indian J Med Res 2000;112:73–77.
- [386] Preuss HG, Echard B, Polansky MM, Anderson R. Whole cinnamon and aqueous extracts ameliorate sucrose-induced blood pressure elevations in spontaneously hypertensive rats. J Am Coll Nutr. 2006;25(2):144–150.
- [387] Pumbwe L, Skilbeck CA, Wexler HM. Induction of multiple antibiotic resistance in *Bacteroides fragilis* by benzene and benzene-derived active compounds of commonly used analgesics, antiseptics and cleaning agents. J Antimicrob Chemother. 2007;60(6):1288–1297.
- [388] Qidwai W, Alim SR, Dhanani RH, Jehangir S, Nasrullah A, Raza A. Use of folk remedies among patients in Karachi, Pakistan. J Ayub Med Coll. Abbottabad 2003;15(2):31–33.
- [389] Qin B, Dawson H, Polansky MM, Anderson RA. Cinnamon extract attenuates TNF-alphainduced intestinal lipoprotein ApoB48 overproduction by regulating inflammatory, insulin, and lipoprotein pathways in enterocytes. Horm Metab Res. 2009;41(7):516–522.
- [390] Qin B, Nagasaki M, Ren M, Bajotto G, Oshida Y, Sato Y. Cinnamon extract (traditional herb) potentiates in vivo insulin-regulated glucose utilization via enhancing insulin signaling in rats. Diabetes Res Clin Pract. 2003;62(3):139–148.
- [391] Qin B, Nagasaki M, Ren M, Bajotto G, Oshida Y, Sato Y. Cinnamon extract prevents the insulin resistance induced by a high-fructose diet. Horm. Metab Res. 2004;36(2):119–125.
- [392] Qin B, Polansky MM, Sato Y, Adeli K, Anderson RA. Cinnamon extract inhibits the postprandial overproduction of apolipoprotein B48-containing lipoproteins in fructose-fed animals. J Nutr Biochem. 2009;20(11):901–908.
- [393] Quale JM, Landman D, Zaman MM, Burney S, Sathe SS. In vitro activity of *Cinnamo-mum zeylanicum* against azole resistant and sensitive *Candida* species and a pilot study of cinnamon for oral candidiasis. Am J Chin Med. 1996;24(2):103–109.
- [394] Raj RK. Screening of indigenous plants for anthelmintic action against human *Ascaris lumbricoides*: Part–II. Indian J Physiol Pharmacol. 1975;19(1): 47–49.

- [395] Rao CV, Vijayakumar M, Sairam K, Kumar V. Antidiarrhoeal activity of the standardised extract of *Cinnamomum tamala* in experimental rats. Nat Med. (Tokyo) 2008;62(4):396–402.
- [396] Rao YK, Fang SH, Tzeng YM. Evaluation of the anti-inflammatory and anti-proliferation tumoral cells activities of *Antrodia camphorata*, *Cordyceps sinensis*, and *Cinnamomum osmophloeum* bark extracts. J Ethnopharmacol. 2007;114(1):78–85.
- [397] Rastogi SC, Johansen JD, Frosch P, Menne T, Bruze M, Lepoittevin JP, Dreier B, Andersen KE, White IR. Deodorants on the European market: quantitative chemical analysis of 21 fragrances. Contact Dermatitis 1998;38(1):29–35.
- [398] Ravishankar S, Zhu L, Law B, Joens L, Friedman M. Plant-derived compounds inactivate antibiotic-resistant *Campylobacter jejuni* strains. J Food Prot. 2008;71(6):1145–1149.
- [399] Ravishankar S, Zhu L, Olsen CW, McHugh TH, Friedman M. Edible apple film wraps containing plant antimicrobials inactivate foodborne pathogens on meat and poultry products. J Food Sci. 2009;74(8):M440–M445.
- [400] Raybaudi-Massilia RM, Mosqueda-Melgar J, Martin-Belloso O. Edible alginate-based coating as carrier of antimicrobials to improve shelf-life and safety of fresh-cut melon. Int J Food Microbiol. 2008;121(3):313–327.
- [401] Raybaudi-Massilia RM, Rojas-Grau MA, Mosqueda-Melgar J, Martin-Belloso O. Comparative study on essential oils incorporated into an alginate-based edible coating to assure the safety and quality of fresh-cut Fuji apples. J Food Prot. 2008;71(6):1150–1161.
- [402] Reddy AM, Seo JH, Ryu SY, Kim YS, Kim YS, Min KR, Kim Y. Cinnamaldehyde and 2-methoxycinnamaldehyde as NF-kappaB inhibitors from *Cinnamomum cassia*. Planta Med. 2004;70(9):823–827.
- [403] Reiter M, Brandt W. Relaxant effects on tracheal and ileal smooth muscles of the guinea pig. Arzneimittelforschung 1985;35(1A):408–414.
- [404] Rodriguez M, Alvarez M, Zayas M. [Microbiological quality of spices consumed in Cuba]. Rev Latinoam Microbiol. 1991;33(2–3):149–151.
- [405] Roffey B, Atwal A, Kubow S. Cinnamon water extracts increase glucose uptake but inhibit adiponectin secretion in 3T3-L1 adipose cells. Mol Nutr Food Res. 2006;50(8):739–745
- [406] Rojas-Grau MA, Avena-Bustillos RJ, Friedman M, Henika PR, Martin-Belloso O, McHugh TH. Mechanical, barrier, and antimicrobial properties of apple puree edible films containing plant essential oils. J Agric Food Chem. 2006;54(24):9262–9267.
- [407] Romaguera C, Grimalt F. Sensitization to cinnamic aldehyde in toothpaste. Contact Dermatitis 1978;4(6):377–378.
- [408] Roselli M, Britti MS, Huerou-Luron I, Marfaing H, Zhu WY, Mengheri E. Effect of different plant extracts and natural substances (PENS) against membrane damage induced by enterotoxigenic *Escherichia coli* K88 in pig intestinal cells. Toxicol In Vitro 2007;21(2):224–229.
- [409] Rosti L, Gastaldi G. Chronic salmonellosis and cinnamon. Pediatrics 2005;116(4):1057.
- [410] Rosti L, Gastaldi G, Frigiola A. Cinnamon and bacterial enteric infections. Indian J Pediatr. 2008;75(5):529–530.
- [411] Roussel AM, Hininger I, Benaraba R, Ziegenfuss TN, Anderson RA. Antioxidant effects of a cinnamon extract in people with impaired fasting glucose that are overweight or obese. J Am Coll Nutr. 2009;28(1):16–21.
- [412] Rudkowska I. Functional foods for health: focus on diabetes. Maturitas 2009;62(3):263–269.
- [413] Rudzki E, Grzywa Z. [Value of a cinnamon and citronellyl oil mixture for the detection of allergy to perfumes]. Przegl Dermatol. 1982;69(1–2):33–38.
- [414] Saeki Y, Ito Y, Shibata M, Sato Y, Okuda K, Takazoe I. Antimicrobial action of natural substances on oral bacteria. Bull Tokyo Dent Coll. 1989;30(3):129–135.
- [415] Sainio EL, Kanerva L. Contact allergens in toothpastes and a review of their hypersensitivity. Contact Dermatitis 1995;33(2):100–105.

- [416] Sakai S, Harada M, Morishita I, Kikuchi T. [Relation between the combination among the chief components of ephedra, Chinese cinnamon, Japanese apricot seeds and their pharmacological actions.]. Yakugaku Zasshi 1964;84:183–187.
- [417] Salam TN, Fowler JF, Jr. Balsam-related systemic contact dermatitis. J Am Acad Dermatol. 2001;45(3):377–381.
- [418] Salmieri S, Lacroix M. Physicochemical properties of alginate/polycaprolactone-based films containing essential oils. J Agric Food Chem 2006;54(26):10205–10214.
- [419] Sambaiah K, Srinivasan K. Influence of spices and spice principles on hepatic mixed function oxygenase system in rats. Indian J Biochem Biophys 1989;26(4):254–258.
- [420] Sambaiah K, Srinivasan K. Effect of cumin, cinnamon, ginger, mustard and tamarind in induced hypercholesterolemic rats. Nahrung 1991;35(1):47–51.
- [421] Samuelsen OB, Brenna J, Solheim E, Scheline RR. Metabolism of the cinnamon constituent o-methoxycinnamaldehyde in the rat. Xenobiotica 1986;16(9):845–852.
- [422] Sanchez-Perez J, Garcia-Diez A. Occupational allergic contact dermatitis from eugenol, oil of cinnamon and oil of cloves in a physiotherapist. Contact Dermatitis 1999;41(6):346– 347
- [423] Sato K, Krist S, Buchbauer G. Antimicrobial effect of trans-cinnamaldehyde, (-)-perillaldehyde, (-)-citronellal, citral, eugenol and carvacrol on airborne microbes using an airwasher. Biol Pharm Bull. 2006;29(11):2292–2294.
- [424] Scheidegger UA, Fluck CE, Scheidegger K, Diem P, Mullis PE. [Role of complementary medicine in type 1 diabetes mellitus in two Swiss centres]. Praxis (Bern) 2009;98(18):1001–1005.
- [425] Schirmer BC, Heiberg R, Eie T, Moretro T, Maugesten T, Carlehog M, Langsrud S. A novel packaging method with a dissolving CO(2) headspace combined with organic acids prolongs the shelf life of fresh salmon. Int J Food Microbiol. 2009;133(1–2):154–160.
- [426] Schnuch A, Lessmann H, Geier J, Frosch PJ, Uter, W. Contact allergy to fragrances: frequencies of sensitization from 1996 to 2002. Results of the IVDK*. Contact Dermatitis 2004;50(2):65–76.
- [427] Schoene NW, Kelly MA, Polansky MM, Anderson RA. Water-soluble polymeric polyphenols from cinnamon inhibit proliferation and alter cell cycle distribution patterns of hematologic tumor cell lines. Cancer Lett. 2005;230(1):134–140.
- [428] Schoene NW, Kelly MA, Polansky MM, Anderson RA. A polyphenol mixture from cinnamon targets p38 MAP kinase-regulated signaling pathways to produce G2/M arrest. J Nutr Biochem. 2009;20(8):614–620.
- [429] Schorr WF. Cinnamic aldehyde allergy. Contact Dermatitis 1975;1(2):108–111.
- [430] Schubert HJ. Skin diseases in workers at a perfume factory. Contact Dermatitis 2006;55(2):81–83.
- [431] Schwab AH, Harpestad AD, Swartzentruber A, Lanier JM, Wentz BA, Duran AP, Barnard RJ, Read RB, Jr. Microbiological quality of some spices and herbs in retail markets. Appl Environ Microbiol. 1982;44(3):627–630.
- [432] Schwartz RH. Cinnamon oil: kids use it to get high. Clin Pediatr. (Phila.) 1990;29(3):196.
- [433] Scully C. Reaction to cinnamon. Br Dent J. 2006;201(8):489.
- [434] Sedghizadeh PP, Allen CM. White plaque of the lateral tongue. J Contemp Dent Pract. 2002;3(3):46–50.
- [435] Sekizawa J, Shibamoto T. Genotoxicity of safrole-related chemicals in microbial test systems. Mutat Res. 1982;101(2):127–140.
- [436] Senhaji O, Faid M, Kalalou I. Inactivation of *Escherichia coli* O157:H7 by essential oil from *Cinnamomum zeylanicum*. Braz J Infect Dis. 2007;11(2):234–236.
- [437] Senma M, Fujiwara N, Sasaki S, Toyama M, Sakaguchi K, Takaoka I. Studies on the cutaneous sensitization reaction of guinea pigs to purified aromatic chemicals. Acta Derm Venereol. 1978;58(2):121–124.
- [438] Seo UK, Lee YJ, Kim JK, Cha BY, Kim DW, Nam KS, Kim CH. Large-scale and effective screening of Korean medicinal plants for inhibitory activity on matrix metalloproteinase-9. J Ethnopharmacol. 2005;97(1):101–106.

- [439] Shah AH, Al Shareef AH, Ageel AM, Qureshi S. Toxicity studies in mice of common spices, Cinnamomum zeylanicum bark and Piper longum fruits. Plant Foods Hum Nutr. 1998;52(3):231–239.
- [440] Shahverdi AR, Monsef-Esfahani HR, Tavasoli F, Zaheri A, Mirjani R. Transcinnamaldehyde from *Cinnamomum zeylanicum* bark essential oil reduces the clindamycin resistance of *Clostridium difficile* in vitro. J Food Sci. 2007;72(1):S055–S058.
- [441] Shakila RJ, Vasundhara TS, Rao DV. Inhibitory effect of spices on in vitro histamine production and histidine decarboxylase activity of *Morganella morganii* and on the biogenic amine formation in mackerel stored at 30 degrees C. Z Lebensm Unters Forsch. 1996;203(1):71–76.
- [442] Shan B, Cai YZ, Brooks JD, Corke H. Antibacterial properties and major bioactive components of cinnamon stick (*Cinnamomum burmannii*): activity against foodborne pathogenic bacteria. J Agric Food Chem. 2007;55(14):5484–5490.
- [443] Shan B, Cai YZ, Sun M, Corke H. Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. J Agric Food Chem. 2005;53(20):7749–7759.
- [444] Shan BE, Yoshida Y, Sugiura T, Yamashita U. Stimulating activity of Chinese medicinal herbs on human lymphocytes in vitro. Int J Immunopharmacol. 1999;21(3):149–159.
- [445] Shan R, Lee KJ, Kwon BM, Lee CH. Protein binding characteristics of 2'-benzoyloxycinnamaldehyde. Drug Dev Ind Pharm. 2005;31(6):545–549.
- [446] Sharififar F, Moshafi MH, Dehghan-Nudehe G, Ameri A, Alishahi F, Pourhemati A. Bioassay screening of the essential oil and various extracts from 4 spices medicinal plants. Pak J Pharm Sci. 2009;22(3):317–322.
- [447] Sharma N, Trikha P, Athar M, Raisuddin S. Inhibition of benzo[a]pyreneand cyclophoshamide-induced mutagenicity by *Cinnamomum cassia*. Mutat Res 2001;480–481:179–188.
- [448] Shaughnessy DT, Schaaper RM, Umbach DM, Demarini DM. Inhibition of spontaneous mutagenesis by vanillin and cinnamaldehyde in *Escherichia coli*: dependence on recombinational repair. Mutat Res. 2006;602(1–2):54–64.
- [449] Shen LR, Li HY, Zhou YG, Gu S, Lou YG. [Ovicidal activity of nine essential oils against *Chrysomya megacephara* in bacon and kipper]. Ying Yong Sheng Tai Xue Bao. 2007;18(10):2343–2346.
- [450] Shen Q, Chen F, Luo J. [Comparison studies on chemical constituents of essential oil from ramulus Cinnamomi and cortex Cinnamomi by GC-MS]. Zhong Yao Cai. 2002;25(4):257–258.
- [451] Shen YC, Chou CJ, Wang YH, Chen CF, Chou YC, Lu MK. Anti-inflammatory activity of the extracts from mycelia of *Antrodia camphorata* cultured with water-soluble fractions from five different *Cinnamomum* species. FEMS Microbiol Lett. 2004;231(1):137–143.
- [452] Shimada Y, Goto H, Kogure T, Kohta K, Shintani T, Itoh T, Terasawa K. Extract prepared from the bark of *Cinnamomum cassia* Blume prevents glutamate-induced neuronal death in cultured cerebellar granule cells. Phytother Res. 2000;14(6):466–468.
- [453] Shimada Y, Yokoyama K, Goto H, Sekiya N, Mantani N, Tahara E, Hikiami H, Terasawa K. Protective effect of keishi-bukuryo-gan and its constituent medicinal plants against nitric oxide donor-induced neuronal death in cultured cerebellar granule cells. Phytomedicine 2004;11(5):404–410.
- [454] Shin DS, Kim J, Han DC, Son KH, Lee CW, Kim HM, Hong SH, Kwon BM. Synthesis and biological evaluation of cinnamyl compounds as potent antitumor agents. Bioorg Med Chem Lett. 2007;17(19):5423–5427.
- [455] Shobana S, Naidu KA. Antioxidant activity of selected Indian spices. Prostaglandins Leukot Essent Fatty Acids 2000;62(2):107–110.
- [456] Si W, Gong J, Chanas C, Cui S, Yu H, Caballero C, Friendship RM. In vitro assessment of antimicrobial activity of carvacrol, thymol and cinnamaldehyde towards *Salmonella* serotype Typhimurium DT104: effects of pig diets and emulsification in hydrocolloids. J Appl Microbiol. 2006;101(6):1282–1291.

- [457] Simic A, Sokovic MD, Ristic M, Grujic-Jovanovic S, Vukojevic J, Marin PD. The chemical composition of some Lauraceae essential oils and their antifungal activities. Phytother Res. 2004;18(9):713–717.
- [458] Singh G, Maurya S, DeLampasona MP, Catalan CA. A comparison of chemical, antioxidant and antimicrobial studies of cinnamon leaf and bark volatile oils, oleoresins and their constituents. Food ChemToxicol. 2007;45(9):1650–1661.
- [459] Smith-Palmer A, Stewart J, Fyfe L. Antimicrobial properties of plant essential oils and essences against five important food-borne pathogens. Lett Appl Microbiol. 1998;26(2):118–122.
- [460] Solomon TP, Blannin AK. Effects of short-term cinnamon ingestion on in vivo glucose tolerance. Diabetes Obes Metab. 2007;9(6):895–901.
- [461] Solomon TP, Blannin AK. Changes in glucose tolerance and insulin sensitivity following 2 weeks of daily cinnamon ingestion in healthy humans. Eur J Appl Physiol. 2009;105(6):969–976.
- [462] Sparks T. Cinnamon oil burn. West J Med. 1985;142(6):835.
- [463] Speer F. Food allergy: the 10 common offenders. Am Fam Physician 1976;13(2):106–112.
- [464] Stager J, Wuthrich B, Johansson SG. Spice allergy in celery-sensitive patients. Allergy 1991;46(6):475–478.
- [465] Strauss RM, Orton DI. Allergic contact cheilitis in the United Kingdom: a retrospective study. Am J Contact Dermat. 2003;14(2):75–77.
- [466] Su MJ, Chen WP, Lo TY, Wu TS. Ionic mechanisms for the antiarrhythmic action of cinnamophilin in rat heart. J Biomed Sci. 1999;6(6):376–386.
- [467] Subash, Babu P, Prabuseenivasan S, Ignacimuthu S. Cinnamaldehyde a potential antidiabetic agent. Phytomedicine 2007;14(1):15–22.
- [468] Subehan, Usia T, Iwata H, Kadota S, Tezuka Y. Mechanism-based inhibition of CYP3A4 and CYP2D6 by Indonesian medicinal plants. J Ethnopharmacol. 2006;105(3):449–455.
- [469] Subehan S, Kadota S, Tezuka Y. In vitro mechanism-based inactivation of cytochrome P450 3A4 by a new constituent of *Cinnamomum burmani*. Planta Med. 2008;74(12):1474–1480.
- [470] Suganthi R, Rajamani S, Ravichandran MK, Anuradha CV. Effect of food seasoning spices mixture on biomarkers of oxidative stress in tissues of fructose-fed insulin-resistant rats. J Med Food 2007;10(1):149–153.
- [471] Sui Y, Qiu D, Xie C, Chen K. [Observation of antiarrhythmic effects of Cinnamo-mum migao H. W. Li on experimental arrhythmia]. Zhongguo Zhong Yao Za Zhi. 1998;23(8):495–497.
- [472] Sun WS, Imai A, Tagami K, Sugiyama M, Furui T, Tamaya T. In vitro stimulation of granulosa cells by a combination of different active ingredients of unkei-to. Am J Chin Med. 2004;32(4):569–578.
- [473] Sun X, Sui Y, Qiu D. [Influence of *Cinnamomum migao* H.W. Li oil on hemodynamic action in anesthetized cats]. Zhongguo Zhong, Yao Za Zhi. 1995;20(10):622–624.
- [474] Suppapitiporn S, Kanpaksi N, Suppapitiporn S. The effect of *Cinnamon cassia* powder in type 2 diabetes mellitus. J Med Assoc Thai. 2006;89(Suppl 3):S200–S205.
- [475] Svedman C, Bruze M, Johansen JD, Andersen KE, Goossens A, Frosch PJ, Lepoittevin JP, Rastogi S, White IR, Menne T. Deodorants: an experimental provocation study with hydroxycitronellal. Contact Dermatitis 2003;48(4):217–223.
- [476] Swanston-Flatt SK, Day C, Bailey CJ, Flatt PR. Evaluation of traditional plant treatments for diabetes: studies in streptozotocin diabetic mice. Acta Diabetol Lat. 1989;26(1):51–55.
- [477] Tabak M, Armon R, Neeman I. Cinnamon extracts' inhibitory effect on *Helicobacter pylori*. J Ethnopharmacol. 1999;67(3):269–277.
- [478] Tabak M, Armon R, Potasman I, Neeman I. In vitro inhibition of *Helicobacter pylori* by extracts of thyme. J Appl Bacteriol. 1996;80(6):667–672.
- [479] Taher M, Abdul Majid FA, Sarmidi MR. Cinnamtannin B1 activity on adipocytes formation. Med J Malaysia 2004;(59 Suppl B):97–98.

- [480] Takenaga M, Hirai A, Terano T, Tamura Y, Kitagawa H, Yoshida S. In vitro effect of cinnamic aldehyde, a main component of Cinnamomi cortex, on human platelet aggregation and arachidonic acid metabolism. J Pharmacobiodyn. 1987;10(5):201–208.
- [481] Talpur N, Echard B, Ingram C, Bagchi D, Preuss H. Effects of a novel formulation of essential oils on glucose-insulin metabolism in diabetic and hypertensive rats: a pilot study. Diabetes Obes Metab. 2005;7(2):193–199.
- [482] Tanaka N, Sekiya N, Hattori M, Goto H, Shibahara N, Shimada Y, Terasawa K. Measurement of plasma procyanidin B-2 and procyanidin B-3 levels after oral administration in rat. Phytomedicine 2003;10(2–3):122–126.
- [483] Tanaka S, Yoon YH, Fukui H, Tabata M, Akira T, Okano K, Iwai M, Iga Y, Yokoyama K. Antiulcerogenic compounds isolated from Chinese cinnamon. Planta Med. 1989;55(3):245–248.
- [484] Tang M, Larson-Meyer DE, Liebman M. Effect of cinnamon and turmeric on urinary oxalate excretion, plasma lipids, and plasma glucose in healthy subjects. Am J Clin Nutr. 2008;87(5):1262–1267.
- [485] Tatrai E, Adamis Z, Ungvary G. The pulmonary toxicity of cinnamon dust in rats. Indian J Med Res. 1995;102:287–292.
- [486] Tayel AA, El Tras WF. Possibility of fighting food borne bacteria by Egyptian folk medicinal herbs and spices extracts. J Egypt Public Health Assoc. 2009;84(1–2):21–32.
- [487] Temesvari E, Soos G, Podanyi B, Kovacs I, Nemeth I. Contact urticaria provoked by balsam of Peru. Contact Dermatitis 1978;4(2):65–68.
- [488] Teuscher E. Ein Handbuch der Gewürze, Gewürzkräuter, Ihrer Ätherischen Öle und Gewürzmischungen. Gewürzdrogen. 2003;423–429.
- [489] Thantsin K, Zhang Q, Yang J, Wang Q. Composition of semivolatile compounds of 10 *Cinnamomum* species from China and Myanmar. Nat Prod Res. 2008;22(7):576–583.
- [490] Thyne G, Young DW, Ferguson MM. Contact stomatitis caused by toothpaste. N Z Dent J. 1989;85(382):124–126.
- [491] Toda S. Inhibitory effects of aromatic herbs on lipid peroxidation and protein oxidative modification by copper. Phytother Res. 2003;17(5):546–548.
- [492] Tonkal AM, Morsy TA. An update review on *Commiphora molmol* and related species. J Egypt Soc Parasitol. 2008;38(3):763–796.
- [493] Tremblay S, Avon SL. Contact allergy to cinnamon: case report. J Can Dent Assoc. 2008;74(5):445–461.
- [494] Tsai PJ, Tsai TH, Yu CH, Ho SC. Evaluation of NO-suppressing activity of several Mediterranean culinary spices. Food Chem Toxicol. 2007;45(3):440–447.
- [495] Tunc S, Chollet E, Chalier P, Preziosi-Belloy L, Gontard N. Combined effect of volatile antimicrobial agents on the growth of *Penicillium notatum*. Int J Food Microbiol. 2007;113(3):263–270.
- [496] Tung YT, Chua MT, Wang SY, Chang ST. Anti-inflammation activities of essential oil and its constituents from indigenous cinnamon (*Cinnamomum osmophloeum*) twigs. Bioresour. Technol. 2008;99(9):3908–3913.
- [497] Turgis M, Han J, Borsa J, Lacroix M. Combined effect of natural essential oils, modified atmosphere packaging, and gamma radiation on the microbial growth on ground beef. J Food Prot. 2008;71(6):1237–1243.
- [498] Ungsurungsie M, Paovalo C, Noonai A. Mutagenicity of extracts from Ceylon cinnamon in the rec assay. Food Chem Toxicol. 1984;22(2):109–112.
- [499] Ungsurungsie M, Suthienkul O, Paovalo C. Mutagenicity screening of popular Thai spices. Food Chem Toxicol. 1982;20(5):527–530.
- [500] Uragoda CG. Asthma and other symptoms in cinnamon workers. Br J Ind Med. 1984;41(2):224–227.
- [501] Uragoda CG. Symptoms in spice workers. J Trop Med Hyg. 1992;95(2):136–139.
- [502] Usta J, Kreydiyyeh S, Bajakian K, Nakkash-Chmaisse H. In vitro effect of eugenol and cinnamaldehyde on membrane potential and respiratory chain complexes in isolated rat liver mitochondria. Food Chem Toxicol. 2002;40(7):935–940.

- [503] Usta J, Kreydiyyeh S, Barnabe P, Bou-Moughlabay Y, Nakkash-Chmaisse H. Comparative study on the effect of cinnamon and clove extracts and their main components on different types of ATPases. Hum Exp Toxicol. 2003;22(7):355–362.
- [504] Valcarcel R, Bennett JW, Vitanza J. Effect of selected inhibitors on growth, pigmentation, and aflatoxin production by *Aspergillus parasiticus*. Mycopathologia 1986;94(1):7–10.
- [505] Valero M, Frances E. Synergistic bactericidal effect of carvacrol, cinnamaldehyde or thymol and refrigeration to inhibit *Bacillus cereus* in carrot broth. Food Microbiol. 2006;23(1):68–73.
- [506] Valero M, Giner MJ. Effects of antimicrobial components of essential oils on growth of Bacillus cereus INRA L2104 in and the sensory qualities of carrot broth. Int J Food Microbiol. 2006;106(1):90–94.
- [507] Valero M, Salmeron MC. Antibacterial activity of 11 essential oils against *Bacillus cereus* in tyndallized carrot broth. Int J Food Microbiol. 2003;85(1–2):73–81.
- [508] Van den Akker TW, Roesyanto-Mahadi ID, van Toorenenbergen AW, van Joost T. Contact allergy to spices. Contact Dermatitis 1990;22(5):267–272.
- [509] Van Tol RW, Swarts HJ, van der Linden A, Visser JH. Repellence of the red bud borer Resseliella oculiperda from grafted apple trees by impregnation of rubber budding strips with essential oils. Pest Manag Sci. 2007;63(5):483–490.
- [510] Vanschoonbeek K, Thomassen BJ, Senden JM, Wodzig WK, van Loon LJ. Cinnamon supplementation does not improve glycemic control in postmenopausal type 2 diabetes patients. J Nutr. 2006;136(4):977–980.
- [511] Veal L. The potential effectiveness of essential oils as a treatment for headlice, *Pediculus humanus capitis*. Complement Ther Nurs Midwifery 1996;2(4):97–101.
- [512] Wang G, Wang L, Xiong ZY, Mao B, Li TQ. Compound salvia pellet, a traditional Chinese medicine, for the treatment of chronic stable angina pectoris compared with nitrates: a meta-analysis. Med Sci Monit. 2006;12(1):SR1–SR7.
- [513] Wang HF, Wang YK, Yih KH. DPPH free-radical scavenging ability, total phenolic content, and chemical composition analysis of forty-five kinds of essential oils. J Cosmet Sci. 2008;59(6):509–522.
- [514] Wang JG, Anderson RA, Graham GM, III, Chu MC, Sauer MV, Guarnaccia MM, Lobo RA. The effect of cinnamon extract on insulin resistance parameters in polycystic ovary syndrome: a pilot study. Fertil Steril 2007;88(1):240–243.
- [515] Wang SY, Chen PF, Chang ST. Antifungal activities of essential oils and their constituents from indigenous cinnamon (*Cinnamomum osmophloeum*) leaves against wood decay fungi. Bioresour Technol. 2005;96(7):813–818.
- [516] Wang SY, Yang CW, Liao JW, Zhen WW, Chu FH, Chang ST. Essential oil from leaves of *Cinnamomum osmophloeum* acts as a xanthine oxidase inhibitor and reduces the serum uric acid levels in oxonate-induced mice. Phytomedicine 2008 [Epub ahead of print].
- [517] Wang WK, Hsu TL, Huang ZY, Wang YY. Collective effect of a Chinese formula a study of xiao-jian-zhong-tang. Am J Chin Med. 1995;23(3–4):299–304.
- [518] Wannissorn B, Jarikasem S, Siriwangchai T, Thubthimthed S. Antibacterial properties of essential oils from Thai medicinal plants. Fitoterapia 2005;76(2):233–236.
- [519] Warin RP, Smith RJ. Chronic urticaria investigations with patch and challenge tests. Contact Dermatitis 1982;8(2):117–121.
- [520] Warnke PH, Becker ST, Podschun R, Sivananthan S, Springer IN, Russo PA, Wiltfang J, Fickenscher H, Sherry E. The battle against multi-resistant strains: renaissance of antimicrobial essential oils as a promising force to fight hospital-acquired infections. J Craniomaxillofac Surg. 2009;37(7):392–397.
- [521] Warshaw EM, Furda LM, Maibach HI, Rietschel RL, Fowler JF, Jr, Belsito DV, Zug KA, DeLeo VA, Marks JG, Jr, Mathias CG, Pratt MD, Sasseville D, Storrs FJ, Taylor JS. Anogenital dermatitis in patients referred for patch testing: retrospective analysis of cross-sectional data from the North American Contact Dermatitis Group, 1994–2004. Arch Dermatol. 2008;144(6):749–755.
- [522] Watt K, Christofi N, Young R. The detection of antibacterial actions of whole herb tinctures using luminescent *Escherichia coli*. Phytother Res. 2007;21(12):1193–1199.

- [523] Wei A, Shibamoto T. Antioxidant activities of essential oil mixtures toward skin lipid squalene oxidized by UV irradiation. Cutan Ocul Toxicol. 2007;26(3):227–233.
- [524] Weibel H, Hansen J, Andersen KE. Cross-sensitization patterns in guinea pigs between cinnamaldehyde, cinnamyl alcohol and cinnamic acid. Acta Derm Venereol. 1989;69(4):302–307.
- [525] Westra WH, McMurray JS, Califano J, Flint PW, Corio RL. Squamous cell carcinoma of the tongue associated with cinnamon gum use: a case report. Head Neck 1998;20(5):430–433.
- [526] White A, Nunes C, Escudier M, Lomer MC, Barnard K, Shirlaw P, Challacombe SJ, Sanderson JD. Improvement in orofacial granulomatosis on a cinnamon- and benzoatefree diet. Inflamm Bowel Dis. 2006;12(6):508–514.
- [527] Wickramasinghe RH, Muller G, Norpoth K. Spectral evidence of interaction of spice constituents with hepatic microsomal cytochrome P-450. Cytobios 1980;29(113):25– 27
- [528] Wijesekera RO. Historical overview of the cinnamon industry. CRC Crit Rev Food Sci Nutr. 1978;10(1):1–30.
- [529] Wondrak GT, Cabello CM, Villeneuve NF, Zhang S, Ley S, Li Y, Sun Z, Zhang DD. Cinnamoyl-based Nrf2-activators targeting human skin cell photo-oxidative stress. Free Radic Biol Med. 2008;45(4):385–395.
- [530] Wong SY, Grant IR, Friedman M, Elliott CT, Situ C. Antibacterial activities of naturally occurring compounds against *Mycobacterium avium* subsp. paratuberculosis. Appl Environ Microbiol. 2008;74(19):5986–5990.
- [531] Woodruff RC, Mason JM, Valencia R, Zimmering S. Chemical mutagenesis testing in Drosophila: V. Results of 53 coded compounds tested for the National Toxicology Program. Environ Mutagen. 1985;7(5):677–702.
- [532] Wortelboer HM, Usta M, van Zanden JJ, van Bladeren PJ, Rietjens IM, Cnubben NH. Inhibition of multidrug resistance proteins MRP1 and MRP2 by a series of alpha,beta-unsaturated carbonyl compounds. Biochem Pharmacol. 2005;69(12):1879–1890.
- [533] Wu SJ, Ng LT. MAPK inhibitors and pifithrin-alpha block cinnamaldehyde-induced apoptosis in human PLC/PRF/5 cells. Food Chem Toxicol. 2007;45(12):2446–2453.
- [534] Wu SJ, Ng LT, Lin CC. Effects of vitamin E on the cinnamaldehyde-induced apoptotic mechanism in human PLC/PRF/5 cells. Clin Exp Pharmacol Physiol. 2004;31(11):770–776.
- [535] Xiao X, Satake M. [Calcium oxalate crystals in several kinds of Cinnamon bark]. Zhongguo Zhong Yao Za Zhi. 1998;23(9):515–518, 574.
- [536] Yang SS, Hou WC, Huang LW, Lee TH. A new gamma-lactone from the leaves of Cinnamomum kotoense. Nat Prod Res. 2006;20(13):1246–1250.
- [537] Yang ZY, Pei J, Liu RM, Cheng J, Wan DG, Hu R. [Study on relative bioavailability of ferulic acid of combination radix *Angelicae sinensis* and cortex Cinnamomi in mice]. Zhongguo Zhong Yao Za Zhi. 2006;31(12):1012–1015.
- [538] Yeh RY, Shiu YL, Shei SC, Cheng SC, Huang SY, Lin JC, Liu CH. Evaluation of the antibacterial activity of leaf and twig extracts of stout camphor tree, *Cinnamomum kanehirae*, and the effects on immunity and disease resistance of white shrimp, *Litopenaeus vannamei*. Fish Shellfish Immunol. 2009;27(1):26–32.
- [539] Yin XJ, Liu DX, Wang HC, Zhou Y. A study on the mutagenicity of 102 raw pharmaceuticals used in Chinese traditional medicine. Mutat Res. 1991;260(1):73–82.
- [540] Youn HS, Lee JK, Choi YJ, Saitoh SI, Miyake K, Hwang DH, Lee JY. Cinnamaldehyde suppresses toll-like receptor 4 activation mediated through the inhibition of receptor oligomerization. Biochem Pharmacol. 2008;75(2):494–502.
- [541] Yousef RT, Tawil GG. Antimicrobial activity of volatile oils. Pharmazie 1980;35(11):698–701.
- [542] Yu HS, Lee SY, Jang CG. Involvement of 5-HT1A and GABAA receptors in the anxiolytic-like effects of *Cinnamomum cassia* in mice. Pharmacol Biochem Behav. 2007;87(1):164–170.

- [543] Yu SM, Wu TS, Teng CM. Pharmacological characterization of cinnamophilin, a novel dual inhibitor of thromboxane synthase and thromboxane A2 receptor. Br J Pharmacol. 1994;111(3):906–912.
- [544] Yuste J, Fung DY. Inactivation of *Listeria monocytogenes* Scott A 49594 in apple juice supplemented with cinnamon. J Food Prot. 2002;65(10):1663–1666.
- [545] Yuste J, Fung DY. Inactivation of *Salmonella typhimurium* and *Escherichia coli* O157:H7 in apple juice by a combination of nisin and cinnamon. J Food Prot. 2004;67(2):371–377.
- [546] Zanotto KL, Iodi, Carstens M., Carstens E. Cross-desensitization of responses of rat trigeminal subnucleus caudalis neurons to cinnamaldehyde and menthol. Neurosci Lett. 2008;430(1):29–33.
- [547] Zanotto KL, Merrill AW, Carstens MI, Carstens E. Neurons in superficial trigeminal subnucleus caudalis responsive to oral cooling, menthol, and other irritant stimuli. J Neurophysiol 2007;97(2):966–978.
- [548] Zee-Cheng RK. Shi-quan-da-bu-tang (ten significant tonic decoction), SQT. A potent Chinese biological response modifier in cancer immunotherapy, potentiation and detoxification of anticancer drugs. Methods Find Exp Clin Pharmacol. 1992;14(9):725–736.
- [549] Zenner L, Callait MP, Granier C, Chauve, C. In vitro effect of essential oils from Cinnamomum aromaticum, Citrus limon and Allium sativum on two intestinal flagellates of poultry, Tetratrichomonas gallinarum and Histomonas meleagridis. Parasite 2003;10(2):153–157.
- [550] Zhang BG, Liu QF. [Clinic utilization of Guizhi decoction in modern times]. Zhongguo Zhong Yao Za Zhi. 2007;32(8):757–760.
- [551] Zhao JX, Li P, Sheng X, Liu X, Liang DY. [Effects of cinnamyl aldehyde on cell cycle and relafeol proteins expression in NIH3T3 cells]. Zhongguo Zhong Yao Za Zhi. 2007;32(16):1692–1694.
- [552] Zhao X, Zhu JX, Mo SF, Pan Y, Kong LD. Effects of cassia oil on serum and hepatic uric acid levels in oxonate-induced mice and xanthine dehydrogenase and xanthine oxidase activities in mouse liver. J Ethnopharmacol. 2006;103(3):357–365.
- [553] Zhou L, Chen ZX, Chen JY. [Effect of wu lin powder and its ingredients on atrial natriuretic factor level in mice]. Zhongguo Zhong Xi Yi Jie He Za Zhi. 1995;15(1):36–37.
- [554] Zhou Q, Liang G. [Effect of plant alcohol extracts on vegetable aphids and their parasitoids]. Ying Yong Sheng Tai Xue Bao. 2003;14(2):249–252.
- [555] Zhu S, Liu P, Sun S. [Identification of cortex Cinnamomi and its confused species "yin xiang" by FT-Raman spectroscopy]. Zhong Yao Cai. 2001;24(9):636–637.
- [556] Zhu YJ, Song KK, Li ZC, Pan ZZ, Guo YJ, Zhou JJ, Wang Q, Liu B, Chen QX. Antity-rosinase and antimicrobial activities of trans-cinnamaldehyde thiosemicarbazone. J Agric Food Chem. 2009;57(12):5518–5523.
- [557] Zhu ZP, Zhang MF, Shen YQ, Chen GJ. [Pharmacological study on spleen-stomach warming and analgesic action of *Cinnamomum cassia* Presl]. Zhongguo Zhong Yao Za Zhi. 1993;18(9):553–555.
- [558] Ziegenfuss TN, Hofheins JE, Mendel RW, Landis J, Anderson RA. Effects of a water-soluble cinnamon extract on body composition and features of the metabolic syndrome in pre-diabetic men and women. J Int Soc Sports Nutr. 2006;3:45–53.
- [559] Ziment I. History of the treatment of chronic bronchitis. Respiration 1991;58(Suppl 1):37–42.
- [560] Zuskin E, Kanceljak B, Skuric Z, Pokrajac D, Schachter EN, Witek TJ, Maayani S. Immunological and respiratory findings in spice-factory workers. Environ Res. 1988;47(1):95–108.

Copyright of Journal of Dietary Supplements is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.